

Final report



Carbon & Energy Management
ALBERTA RESEARCH COUNCIL
250 KARL CLARK ROAD
EDMONTON, ALBERTA T6N 1E4

Development of a preliminary database of digestate chemistry, heavy metal and pathogen content to assist in Alberta regulation compliance

Part A

Prepared by
Ruth Eckford

Part B

Prepared by Tiejun TJ Gao
and
edited by Ruth Eckford

for

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ABSTRACT

This study was used to report salt concentrations, heavy metal concentrations, fecal coliform and *Salmonella* sp. in pre-digested biowaste substrates and post-digested products. Heavy metal and salt concentrations were followed for digestate separated liquid and solid fractions. Batch anaerobic cultures were used for 25 manure and non-manure substrates and two substrates were chosen to be used in a continuously stirred tank reactor Pilot Plant digestion system. The results showed that manures contained the highest concentrations for heavy metals and that the manure substrates exceeded the Canadian Council of Ministers of the Environment (CCME) guidelines for soil application. Some non-manure substrates including grocery waste, poultry slaughterhouse waste and thermally hydrolyzed waste exceeded the heavy metal soil guideline concentrations from the CCME. Manure and non-manure substrates showed heavy metal concentrations that exceeded the CCME water guidelines for agricultural use. Salt concentrations were highest in manure substrates and the non-manure substrates that contained high salt concentrations were poultry slaughterhouse waste, hog slaughterhouse peptone waste, potato process waste and potato plant vines. Heavy metal and salt concentrations were increased with digestion and the separated liquid fractions contained higher concentrations than the separated solid fractions. Results showed that heavy metals may be retained in large digester systems due to incomplete purging of substrates and precipitation. Levels above the US Environmental Protection Agency (USEPA) Class A and Class B biosolids and CCME compost guidelines for fecal coliform were found in freshly collected dairy and chicken manures. *Salmonella* sp. above the accepted USEPA for Class A biosolids was found in one freshly collected dairy manure. Fecal coliform numbers were decreased on storage of dairy manure. Anaerobic digestion at 55° and 14-day hydraulic retention time using a continuously stirred tank reactor system reduced fecal coliform and *Salmonella* sp.

TABLE OF CONTENTS

PART A: DATA BASE OF DIGESTATE CHEMISTRY	1
1.0 Introduction	1
2.0 Methods	2
2.1 <i>Methane potential using batch culture</i>	2
2.2 <i>Post-digestion separation of digestate into liquid and solid fractions</i>	4
2.3 <i>Chemical analyses</i>	4
2.4 <i>CCME soil and water quality guidelines</i>	8
3.0 Biowaste Substrates	12
4.0 Results	13
4.1 <i>Methane potential using batch culture</i>	13
4.2 <i>Chemical characteristics for biowaste substrates and batch cultures</i>	14
4.3 <i>Chemical characteristics for substrates used in the ARC Pilot Plant</i>	43
4.4 <i>Chemical characteristics for inoculum used in this study</i>	44
4.5 <i>Heavy metals for biowaste substrates and batch cultures</i>	45
4.6 <i>Heavy metals for substrates used in the ARC Pilot Plant</i>	93
4.7 <i>Heavy metals for inoculum used in this study</i>	97
5.0 Heavy Metal Results Compared to CCME Soil and Water Guidelines	100
6.0 Summary	119
7.0 Acknowledgements	120
8.0 References	120
PART B: EVALUATION OF PATHOGEN IN BIOWASTE AND DIGESTATE	122
1.0 Introduction	122
2.0 Objectives	123
3.0 Biowaste and Sample Collection	124
4.0 Testing Methods	125
4.1 <i>Total solid measurements of biowaste</i>	125
4.2 <i>Testing for fecal coliform</i>	125
4.3 <i>Testing for Salmonella sp.</i>	126
4.4 <i>Quality control</i>	126
5.0 Results and Discussion	127
6.0 Summary	129
7.0 Acknowledgements	130
8.0 References	130

LIST OF TABLES

PART A. DATA BASE OF DIGESTATE CHEMISTRY

Table 2. 1: Explanation of chemical parameters analyzed for this study	7
Table 2. 2: CCME soil quality guidelines.....	10
Table 2. 3: CCME water quality guidelines.....	11

Chemical characteristics of substrates

Table 4. 1: Cattle feedlot bed pack manure.....	19
Table 4. 2: Dairy manure #1	20
Table 4. 3: Potato tuber waste.....	21
Table 4. 4: Chicken broiler manure	22
Table 4. 5: Chicken laying hen manure	23
Table 4. 6: Dairy manure #2	24
Table 4. 7: Potato process waste.....	25
Table 4. 8: Potato stems and leaves	26
Table 4. 9: Poultry slaughter waste.....	27
Table 4. 10: Cattle carcass	28
Table 4. 11: Sugar beet tailings.....	29
Table 4. 12: Grocery waste	30
Table 4. 13: Thermally hydrolyzed biowaste #1	31
Table 4. 14: Thermally hydrolyzed biowaste #2	32
Table 4. 15: Thin stillage	33
Table 4. 16: Glycerine with methanol.....	34
Table 4. 17: Cheese whey	35
Table 4. 18: Wet distillers grains	36
Table 4. 19: Cattle slaughterhouse waste - paunch.....	37
Table 4. 20: Solid hog manure.....	38
Table 4. 21: Sewage sludge	39
Table 4. 22: Liquid hog.....	40
Table 4. 23: Liquid sewage.....	41
Table 4. 24: Hog slaughterhouse waste	42

Chemical characteristics for substrates used in the ARC Pilot Plant

Table 4. 25: ARC Pilot Plant substrates.....	43
--	----

Chemical characteristics for inoculum used in this study

Table 4. 26: Inoculum.....	44
----------------------------	----

Heavy metals for biowaste substrates and cultures

Table 4. 27: Heavy metals for cattle feedlot bed pack manure.....	45
Table 4. 28: Heavy metals for cattle feedlot bed pack manure culture.....	46
Table 4. 29: Heavy metals for dairy manure #1.....	47
Table 4. 30: Heavy metals for dairy manure #1 culture	48
Table 4. 31: Heavy metals for potato tuber waste	49
Table 4. 32: Heavy metals for potato tuber waste culture	50
Table 4. 33: Heavy metals for chicken broiler manure.....	51

Table 4. 34: Heavy metals for chicken broiler manure culture.....	52
Table 4. 35: Heavy metals for chicken laying hen manure.....	53
Table 4. 36: Heavy metals for chicken laying hen manure culture	54
Table 4. 37: Heavy metals for dairy manure #2.....	55
Table 4. 38: Heavy metals for dairy manure #2 culture	56
Table 4. 39: Heavy metals for potato process waste.....	57
Table 4. 40: Heavy metals for potato process waste culture.....	58
Table 4. 41: Heavy metals for potato stems and leaves.....	59
Table 4. 42: Heavy metals for potato stems and leaves culture.....	60
Table 4. 43: Heavy metals for poultry slaughter waste	61
Table 4. 44: Heavy metals for poultry slaughter waste culture	62
Table 4. 45: Heavy metals for cattle carcass.....	63
Table 4. 46: Heavy metals for cattle carcass culture	64
Table 4. 47: Heavy metals for sugar beet tailings.....	65
Table 4. 48: Heavy metals for sugar beet tailings culture.....	66
Table 4. 49: Heavy metals for grocery waste	67
Table 4. 50: Heavy metals for grocery waste culture	68
Table 4. 51: Heavy metals for thermally hydrolyzed biowaste (#1).....	69
Table 4. 52: Heavy metals for thermally hydrolyzed biowaste (#1) culture	70
Table 4. 53: Heavy metals for thermally hydrolyzed biowaste (#2).....	71
Table 4. 54: Heavy metals for thermally hydrolyzed biowaste (#2) culture	72
Table 4. 55: Heavy metals for thin stillage	73
Table 4. 56: Heavy metals for thin stillage culture	74
Table 4. 57: Heavy metals for glycerine with methanol.....	75
Table 4. 58: Heavy metals for glycerine with methanol culture.....	76
Table 4. 59: Heavy metals for hog slaughterhouse sludge (DAF).....	77
Table 4. 60: Heavy metals for hog slaughterhouse peptone	78
Table 4. 61: Heavy metals for wet distillers grains	79
Table 4. 62: Heavy metals for wet distillers grains culture	80
Table 4. 63: Heavy metals for cheese whey	81
Table 4. 64: Heavy metals for cheese whey culture	82
Table 4. 65: Heavy metals for cattle slaughterhouse waste - paunch	83
Table 4. 66: Heavy metals for cattle slaughterhouse waste - paunch culture	84
Table 4. 67: Heavy metals for solid hog manure	85
Table 4. 68: Heavy metals for solid hog manure culture.....	86
Table 4. 69: Heavy metals for sewage sludge.....	87
Table 4. 70: Heavy metals for sewage sludge culture	88
Table 4. 71: Heavy metals for liquid hog manure	89
Table 4. 72: Heavy metals for liquid hog culture	90
Table 4. 73: Heavy metals for liquid sewage.....	91
Table 4. 74: Heavy metals for liquid sewage culture.....	92

Heavy metals for substrates used in the ARC Pilot Plant

Table 4. 75: Heavy metals for ARC Pilot Plant (1) mixture.....	93
Table 4. 76: Heavy metals for ARC Pilot Plant (1) culture	94
Table 4. 77: Heavy metals for ARC Pilot Plant (2) mixture.....	95

Table 4. 78: Heavy metals for ARC Pilot Plant (2) culture	96
---	----

Heavy metals for inoculum used in this study

Table 4. 79: Heavy metals for inoculum.....	97
---	----

Table 4. 80: Inoculum mixture and post digestion separation fractions, 44-day	98
---	----

Table 4. 81: Inoculum mixture and post digestion separation fractions, 30-day	99
---	----

Comparison of results to CCME soil and water guidelines

Table 5. 1: CCME soil guideline comparison for manures.	104
--	-----

Table 5. 2: CCME soil guideline comparison for vegetable/fruit/grain.	106
--	-----

Table 5. 3: CCME soil guideline comparison for human and other animal waste.	108
---	-----

Table 5. 4: CCME soil guideline comparison for ARC Pilot Plant samples.	110
--	-----

Table 5. 5: CCME soil guideline comparison for inoculum.....	111
--	-----

Table 5. 6: CCME water guideline comparison for manures	112
---	-----

Table 5. 7: CCME water guideline comparison for vegetable/fruit/grain	113
---	-----

Table 5. 8: CCME water guideline comparison for human and other animal	115
--	-----

Table 5. 9: CCME water guideline comparison for ARC Pilot Plant samples.....	117
--	-----

Table 5. 10: CCME water guideline comparison for inoculum.....	118
--	-----

Part B. Evaluation of pathogen in Biowaste and AD Digestate

Table 5. 1: Summary of microbiology testing results of selected biowaste samples	127
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PART A: DATA BASE OF DIGESTATE CHEMISTRY

1.0 Introduction

For Alberta, there is an ongoing investigation into the possibility of designing biogas facilities that will use various biowaste feedstocks or substrates. Currently, digestate from biowaste substrates other than manure is regulated under sewage sludge. When digestate slurry and separated liquid and solid fractions from anaerobic digesters are designated as sewage sludge, there is an automatic assumption as to the characteristics, and these assumptions may not be accurate. This designation could present economical challenges to an emerging anaerobic digestion industry.

Work that was previously done by the Alberta Research Council showed that elevated heavy metal content is found mostly in manure and slaughterhouse waste and not in general biowaste. However, the limited data from Alberta Research Council does not give Alberta Environment enough information to set guidelines for handling the digestate generated from biogas plants using various biowastes. There is a need to determine the levels of heavy metals, salts, as well as nitrogen, phosphorous and potassium in feedstock biowastes used for anaerobic digestion and the post-digestion digestates, separated liquids and solids. As well, there is a need to determine levels and destruction of pathogens in biowaste during anaerobic digestion. The combined information will provide data for Alberta Environment to develop guidelines for the biogas industry.

Two objectives were used for this study:

- to determine the chemical characteristics, including nutrient, heavy metal and salt content of various biowaste feedstocks or substrates and the chemical characteristics of their digestates and digestates separated into liquids and solids,
- to perform a preliminary assessment of pathogen content and destruction through anaerobic digestion.

2.0 Methods

2.1 Methane potential using batch culture

Methane potential or productivity measurements are used to indicate the yield of methane per unit of a variable. For this trial, the methane potential was performed using batch cultures with chosen substrates and a methanogenic inoculum. The batch culture method was used to determine whether stable methanogenic conditions were obtained for each substrate during anaerobic digestion.

In order to maintain consistent results for this study, the total amount of inoculum required for batch cultures using all substrates was collected in pails from a large digestion system operating under stable conditions. There were no adequate resources available to remix the collected inoculum. However, since the inoculum was obtained from one collection period, it was assumed that the inoculum in each pail did not differ to a large degree. The inoculum was immediately transferred to Alberta Research Council, Biowaste Resource Recovery Laboratory in Vegreville and frozen at -20°C. When a substrate was ready to be tested during the study, the appropriate amount of inoculum was gently thawed, ground, mixed and incubated for 7 days at 55°C to determine its viability, stimulate activity and remove as much excess carbon as possible. Ten sets of substrates with 2 or 3 substrates per test set were used for this study. For each test set, triplicate cultures with only inoculum were set up. At the same time, the quality of the inoculum was evaluated with cellulose (Alfa Aesar, Ward Hill, MA) using triplicate 5% total solids (TS), 300-ml cultures in 1-L Corning™ bottles with black butyl stoppers (Fisher Scientific, Canada).

Methane potential for a chosen substrate was obtained at a predetermined substrate/inoculum VS ratio, temperature and incubation period. Pretesting of substrate was done using 250-ml Wheaton™ culture bottles with cut-out top and butyl inserts (Fisher Scientific, Canada) containing 8% TS, the same TS that was used for the substrates in this study. Various mixtures of substrate and inoculum were evaluated in order to determine the VS ratio that would be used for each substrate.

The method used for methane potential is consistent with the standard methods ISO 11734 and DIN 38414 Part 8. The production of biogas components such as methane, carbon dioxide and hydrogen sulfide were measured. Briefly, substrate was

added to inoculum and the mixture was incubated at 55°C in sealed glass culture bottles for 30 or 45 days. The cultures are considered finished when the daily biogas production was approximately 1% of the total biogas production. The extended incubation period was needed in some cases in order to meet that requirement. During incubation, biogas volume was calculated from pressure readings and biogas components such as methane, carbon dioxide and hydrogen sulfide were determined using gas chromatography (GC). The results for the aforementioned biogases that were produced in cultures containing only inoculum were subtracted from the same biogas results for cultures with substrate plus inoculum. The biogas volume results were normalized to standard conditions.

Over the incubation period, all bottles for each substrate were monitored for pressure. Due to time restraints, gases, including hydrogen, oxygen, nitrogen, methane, carbon dioxide and hydrogen sulfide, were monitored by GC on 5 bottles for each substrate. The methods for batch culture anaerobic digestion are described in the Biowaste Resource Recovery Laboratory manual (1).

After the digestion period, digestate from each culture mixture was separated into liquid and solid fractions. In order to obtain sufficient separated solids from the digestate after incubation, several 2-L Corning™ bottles with black butyl stoppers (Fisher Scientific, Canada) containing 8% TS and 800 ml of culture were used. For the methane potential of the first four test sets of substrates using cattle feedlot bed pack manure, dairy manure (2 types), potato tuber waste, broiler chicken manure, laying hen manure, hog slaughterhouse waste (sludge and peptone), unstripped glycerine, potato process waste and potato vine, 10 to 12 bottles were used. For the remaining test sets of substrates including cattle carcass, poultry slaughterhouse, sugar beet waste, thermally hydrolyzed biowaste (2 types), grocery waste, thin stillage, wet distillers grains, cheese whey, sewage sludge, beef slaughterhouse waste and solid hog manure, it was found that nine bottles produced an adequate amount of separated solids. For the liquid hog and liquid sewage substrates, the TS results were low, and 14 bottles with 5% TS were needed.

The cultures were incubated in New Brunswick Scientific Classic Incubator Shakers (Edison, NJ), C-24 for small culture bottles and E-25R for large bottles. The bottles in the incubators were covered with an insulation jacket to prevent heat loss during pressure and GC analyses. The gas chromatographic instrument was a Varian

CP-4900 Micro-GC (Walnut Creek, CA) equipped with a Molsieve 5Å PLOT (10 m) for analyzing hydrogen, oxygen, nitrogen, and methane and a Poraplot U (10 m) for carbon dioxide and hydrogen sulfide. Pressure readings were done using a SRP PT420 transducer (SRP Control Systems, Mississauga, Ont.) with a 0-25 psig range and readout pressure meter.

2.2 *Post-digestion separation of digestate into liquid and solid fractions*

The overall plan for large-scale anaerobic digestion facilities must include the handling of the post-digestion product or digestate from digesters. Often, the digestate is separated into liquid and solid fractions. There is a need to evaluate digestate and the separated fractions in order to make environmentally safe decisions as to how they will be used.

Laboratory equipment was designed to simulate large-scale anaerobic digestion separators. The equipment was used to separate digestate from batch cultures into liquid and solid fractions after anaerobic digestion for 30 or 45 days. The sieve assembly consisted of three pieces, a nylon bottom fluted piece, a sieve (USA Standard Test Sieve, ASTM E11 specification, 500 µm, #35) to fit over the raised part of the bottom and a nylon top piece to fit inside the sieve. The separation process was done after digestate was scraped from batch culture bottles and mixed. Once the sieve was assembled, the digestate was slowly poured through the sieve and then pressure was applied using a hydraulic press. Liquid was extruded from the solids until the consistency was similar to separated solids from large-scale facilities.

2.3 *Chemical analyses*

Chemical parameters are important indicators for anaerobic digestion processes. They are used to determine the suitability of substrates for digestion, potential toxic conditions that may arise during digestion and what nutrient resources remain after digestion. Optimum conditions for ultimate methane yield may not be achieved because of inhibitory factors contained in certain substrates. Therefore, it is beneficial to test new substrates for factors that may inhibit methanogenic activity.

Each biowaste substrate was collected at the place of origin by non-ARC staff into containers supplied by the company or into new clean containers supplied by ARC. The samples were immediately shipped to ARC by courier or other means. Upon arrival at ARC, the substrates were carefully mixed and ground. They were then proportioned into new clean small containers and analyzed immediately or frozen at -20°C and analyzed as soon as possible. The methods used for handling substrates after arrival and preparation for analyses are described in the Biowaste Resource Recovery Laboratory Manual (1).

Due to time and economic restraints, the inoculum was completely analyzed for chemical parameters and heavy metals only at the beginning of the study. The results were used to determine the amount of heavy metals that the inoculum would contribute to a digestion mixture. After the inoculum was incubated, complete results for chemical parameters and heavy metals for final cultures and separated fractions were obtained for the inoculum for 30-day and 45-day incubation periods. The 45-day results were obtained at the beginning of the study and the 30-day results were obtained near the end of the study. During the course of the study, when each of the substrate test sets was done, the inoculum was analyzed for TS, total carbon (Total C) and total nitrogen (Total N). These results were used to determine the consistency of the inoculum that was collected for the study.

Table 2.1 gives the list of chemical parameters and a brief explanation of the method used for each parameter. All samples were prepared at the Biowaste Resource Recovery Laboratory at Alberta Research Council in Vegreville, Alberta. Most analyses were done at the Biowaste Resource Recovery Laboratory at Alberta Research Council in Vegreville, Alberta. The ICP-MS analyses for heavy metals and phosphorous were done at the Analytical Chemistry Laboratory, Environmental Management Group at Alberta Research Council in Vegreville, Alberta. The available nitrogen colorimetry and potassium atomic absorption were done at the Natural Resources Analytical Laboratory at the University of Alberta, Edmonton and the Karl Fisher moisture content was done at the Advanced Materials Unit, Alberta Research Council, Devon, Alberta. All samples were analyzed within a very short period of time after they were prepared. If analyses were not done quickly, the samples were frozen at -20°C. Generally, triplicate analyses

were done for each parameter except for Total N, Total C and total sulphur. These were done on 10 replicates for Total N and Total C and 5 replicates for sulphur.

Samples for heavy metal analysis of substrate, initial culture, final culture, liquid and solid were dried at 70°C for 48 h and one mixed sample was sent for analysis. Samples for available heavy metals, nitrogen, phosphorous and potassium analyses were done in triplicate, diluted in distilled water if needed and sent for analysis. There was no need for dilution of the two thermally hydrolyzed biowaste substrates, cheese whey, liquid hog, liquid sewage and two ARC Pilot Plant substrates.

The heavy metals were analyzed as samples were prepared. The available heavy metals were done shortly after the substrates arrived at the laboratory. The heavy metals for the initial culture mixtures were done as soon as the batch cultures were set up and the final digestate and separated liquid and solid samples were sent for analysis at the same time after the incubation period.

Electrical conductivity (EC) and pH were performed on substrates and samples diluted with distilled water to give 2 g TS slurry in 100 ml. The EC results were used to give an indication of the salt content for a substrate and resulting digestate as well as separated liquid and solid fractions. If the TS of the substrate or sample was <2%, no dilution was done. Due to convenience, EC and pH were performed on the same slurry sample. Initial investigation of the EC method showed that there was no linear relationship for EC results when various dilutions were compared for one substrate. Since the substrates had diverse matrices and many substrates required dilution to obtain a slurry for the probe measurement, a consistent dilution of 2 g TS in 100 ml solution was chosen. This allowed comparison of EC results for the substrates used in this study.

Biowaste substrates are not always homogeneous even after rigorous mixing. Therefore, results were difficult to obtain in some cases and more samples were analyzed in order to get a range of results, especially when wet samples were used. These samples are noted in the tables of results. Unless otherwise indicated, results were within 15% of the mean as recommended in the Recommended Methods of Manure Analysis (2).

Table 2. 1: Explanation of chemical parameters analyzed for this study

Parameter	Explanation
Total solids (TS)	Forced-air oven-drying method at 70°C for 48 h; assumed only water removed (3)
Volatile solids (VS)	Muffle furnace at 550°C for 2.5 h giving an indication of organic content (3)
Total nitrogen (Total N)	Leco TruSpec CN method on wet sample
Total carbon (Total C)	Leco TruSpec CN method on wet sample
Total potassium (Total K) ^a	Acid digestion and ICP-MS analysis on dried sample
Total phosphorus (Total P) ^a	Acid digestion and ICP-MS analysis on dried sample
Total sulphur (Total S)	Leco TruSpec S method on dried sample
Available nitrogen (Available NH ₄ ⁺ -N)	Water extracted ammonium-N (NH ₄ ⁺ -N) on wet sample and measured by colorimetry (3)
Available potassium (Available K)	Water extracted potassium on wet sample and measured by atomic absorption spectroscopy
Available phosphorus (Available P)	Water extracted phosphorus on wet sample and measured by ICP-MS ^b
Electrical conductivity (EC) ^c	Probe was used; gives an indication of dissolved salts and ions
pH ^c	Probe was used; determines acid or basic characteristic
Heavy metal analyses ^d measured by ICP-MS ^a	Water extracted available heavy metals on wet sample and acid digested extractable heavy metals on dried sample

^a The biowaste material was oven dried at 70°C for 48 hours. Samples were acid digested using an EPA method (4). Elemental analysis was done using a Perkin Elmer Elan DRC-II Inductively Coupled Plasma Mass Spectrometer (ICP-MS) following an EPA method (5).

^b ICP-MS was done using a Perkin Elmer Elan DRC-II Inductively Coupled Plasma Mass Spectrometer (ICP-MS) following an EPA method (5)

^c Performed on samples diluted with distilled water to give 2 g TS slurry in 100 ml. EC was done using a YSI Model 32 conductance meter and probe (Yellow Springs Inst. Co. Inc., Yellow Springs, OH). The pH was done using probe Model IQ 150, Spectrum Technologies Inc. (Plainfield, IL). EC and pH were performed on the same slurry.

^d For aqueous sample, water extractable dissolved heavy metals were analyzed with no digestion of the wet samples. For dried sample, the sample was dried at 70°C for 48 hours and digested with nitric acid before heavy metals were analyzed.

The names and symbols for the heavy metals analyzed in this study are: Ag-Silver; Al-Aluminum; As-Arsenic; B-Boron; Ba-Barium; Be-Beryllium; Bi-Bismuth; Ca-Calcium; Cd-Cadmium; Cl-Chloride; Co-Cobalt; Cr-Chromium; Cu-Copper; Fe-Iron; Hg-Mercury; Li-Lithium; Mg-Magnesium; Mn-Manganese; Mo-Molybdenum; Na-Sodium; Ni-Nickel; Pb-Lead; Sb-Antimony; Se-Selenium; Sn-Tin; Sr-Strontium; Th-Thorium; Ti-Titanium; Tl-Thallium; U-Uranium; V-Vanadium; Zn-Zinc.

Two sets of biowaste substrates were chosen to be evaluated using the fully automated ARC Pilot Plant system in Vegreville, Alberta (referred to as ARC Pilot Plant from this point). The system includes 80-L continuous stirred tank reactor digesters. This

part of the study was important in order to simulate the digestion process that occurs in large-scale systems. The first ARC Pilot Plant test used a dairy manure (labeled #1 in Section 3.0) plus glycerine and the second ARC Pilot Plant test used the wet distillers grain. For the ARC Pilot Plant substrates, the initial digestion mixture, digestate and separated liquid and solid fractions were analyzed, and the results are in the Pilot Plant section of this report. The substrates were also used in the batch cultures, and the chemical parameter and heavy metal results for these substrates are included with the batch culture results.

For batch cultures in our laboratory, substrate is added to inoculum, placed in an anaerobic environment and monitored for an incubation period. The mixture is not disturbed during the incubation period other than a swirling mix when gases are released and monitored. In the continuous stirred tank reactor digesters at the ARC Pilot Plant, substrate is fed, mixed and digestate removed at predetermined intervals over a 14-day hydraulic retention time. The biogas is continuously monitored using an automatic system. The continuous stirred tank reactor system cannot handle a large number of different substrates over a short period of time. The batch culture, on the other hand, is valuable for obtaining methane potential of a substrate, is less expensive for studies that include a large number of different substrates and can evaluate several substrates at one time. We have found in our work at ARC that these two systems work very well together for the evaluation of substrates.

2.4 CCME soil and water quality guidelines

Excerpts from the Canadian Council of Ministers of the Environment (CCME) guidelines that pertain to heavy metal concentrations in soil from agriculture, parks, residential, commercial and industrial sites and in irrigation and livestock water used for agriculture are presented in Tables 2.2 and 2.3. For each substrate, water extracted available heavy metals on wet substrate and acid digested extractable heavy metals on dried substrate, initial culture mixture, final culture mixture and separated liquid and solid from digestate were compared to the concentrations in the guidelines.

Due to time and financial restraints, only the heavy metals from Table 1 in the Canadian Soil Quality Guidelines for the Protection of Environmental and Human

Health, update 7.0, September 2007 and the footnotes that accompany the heavy metals in the guidelines were used in this study. They are presented in Table 2.2. The conductivity guideline shown in Table 2.2 is from Table 2 in the Canadian Soil Quality Guidelines for the Protection of Environmental and Human Health, update 7.0, September 2007. Conductivity is used to evaluate salt content.

The water quality guidelines for heavy metals are from the Canadian Water Quality Guidelines for the Protection of Agricultural Water Uses, Summary Table, update October 2005. The water quality guidelines use total dissolved solids for salt guidelines. This parameter was not measured in our study.

The reader is encouraged to visit the CCME website to obtain more information about the guidelines and guidelines for other heavy metals from the Canadian Soil Quality Guidelines Table 2 that were released in 1991. The website location is <http://www.ccme.ca>.

Table 2. 2: CCME soil quality guidelines

CCME soil quality guidelines for the protection of environmental and human health (heavy metals given in µg/g and conductivity or salts given in dS/m) for application to soils from CCME Table 1, update 7.0, September 2007 for metals and Table 2 for Conductivity (6)									
Heavy metal values, footnote references from CCME document and year guideline set									
Parameter	Agriculture		Parks/Residential		Commercial		Industrial		Year guideline set
As	12	b	12	b	12	b	12	b	1997
Ba	750	c	500	c	2000	c	2000	c	1999
Cd	1.4	b	10	g	22	b	22	b	1999
Cr (total)	64	b	64	b	87	b	87	b	1997/99
Cu	63	b	63	b	91	b	91	b	1999
Hg	6.6	b	6.6	b	24	b	50	b	1999
Ni	50	l	50	l	50	l	50	l	1999
Pb	70	b	140	b	260	b	600	b	1999
Se	1	b	1	b	2.9	b	2.9	b	2007
TI	1	n	1	o	1	o	1	o	1999
U	23	t	23	t	33	t	300	t	2007
V	130	l	130	l	130	i	130	i	1997
Zn	200	l	200	l	360	l	360	l	1999
Conductivity	2		2		4		4		1991

* For guidelines derived prior to 2004, differentiation between soil texture (coarse/fine) is not applicable.

- b Data are sufficient and adequate to calculate an SQGHH and an SQGE. Therefore the soil quality guideline is the lower of the two and represents a fully integrated *de novo* guideline for this land use, derived in accordance with the soil protocol (CCME 1996; 2006). The corresponding interim soil quality criterion (CCME 1991) is superseded by the soil quality guideline.
- c Data are insufficient/inadequate to calculate an SQGHH, a provisional SQGHH, an SQGE, or a provisional SQGE. Therefore the interim soil quality criterion (CCME 1991) is retained as the soil quality guideline for this land use.
- g The soil–plant–human pathway was not considered in the guideline derivation. If produce gardens are present or planned, a site-specific objective must be derived to take into account the bioaccumulation potential (e.g., adopt the agricultural guideline as objective). The off-site migration check should be recalculated accordingly.
- i Data are sufficient and adequate to calculate only an SQGE. An interim soil quality criterion (CCME 1991) was not established for this land use, therefore the SQGE becomes the soil quality guideline.
- l Data are sufficient and adequate to calculate only an SQGE, which is less than the interim soil quality criterion (CCME 1991) for this land use. Therefore the SQGE becomes the soil quality guideline, which supersedes the interim soil quality criterion for this land use.
- n Data are sufficient and adequate to calculate a provisional SQGHH and an SQGE. The provisional SQGHH is equal to the SQGE and to the existing interim soil quality criterion (CCME 1991) and thus becomes the soil quality guideline, which supersedes the interim soil quality criterion for this land use.
- o Data are sufficient and adequate to calculate a provisional SQGHH and an SQGE. The provisional SQGHH is less than the SQGE and thus becomes the soil quality guideline for this land use.
- t Data are sufficient and adequate to calculate an SQGHH and an SQGE. Therefore the soil quality guideline is the lower of the two and represents a fully integrated *de novo* guideline for this land use.
- u Author's note: Data for conductivity is given in µS/cm in Tables related to substrates used in this study. The conversion factor is µS/cm ÷ 1000 = dS/m.

Table 2. 3: CCME water quality guidelines

CCME water guidelines for the protection of agricultural water uses, Summary table, updated October 2005, in µg/L for agricultural applications (7)							
Heavy metal values, footnote references from CCME document and year guideline set							
Metal		Irrigation		Year	Livestock		Year
Al	d	5000		1987	5000		1987
As	e	100	f	1997	25	f	1997
B	d	500-6000	h	1987	5000		1987
Be	d	100		1987	100	f	1987
Ca	d				1 000 000		1987
Cd		5.1	i,j	1996	80		1996
Cl	d	100 000-700 000	k	1987			
Co	d	50			1000		1987
Cu	d	200-1000	o	1987	500-5000	p	1987
Fe	d	5000		1987			
Hg	d				3		1987
Li	d	2500		1987			
Mn	d	200		1987			
Mo	d	10-50	r	1987	500		1987
Ni	d	200		1987	1000		1987
Pb	d	200		1987	100		1987
Se	d	20-50	s	1987	50		1987
U	d	10	f	1987	200		1987
V	d	100		1987	100		1987
Zn	d	1000-5000	u	1987	50 000		1987

d No fact sheet created.

e The technical document for the guideline is available from the Ontario Ministry of the Environment.

f Interim guideline.

h Boron guideline = 500 µg·L⁻¹ for blackberries = 500–1000 µg·L⁻¹ for peaches, cherries, plums, grapes, cowpeas, onions, garlic, sweet potatoes, wheat, barley, sunflowers, mung beans, sesame, lupins, strawberries, Jerusalem artichokes, kidney beans, and lima beans = 1000–2000 µg·L⁻¹ for red peppers, peas, carrots, radishes, potatoes, and cucumbers = 2000–4000 µg·L⁻¹ for lettuce, cabbage, celery, turnips, Kentucky bluegrass, oats, corn, artichokes, tobacco, mustard, clover, squash, and muskmelons = 4000–6000 µg·L⁻¹ for sorghum, tomatoes, alfalfa, purple vetch, parsley, red beets, and sugar beets = 6000 µg·L⁻¹ for asparagus

i Guideline value slightly modified from CCREM 1987 + Appendixes due to re-evaluation of the significant figures.

j Guideline is crop-specific (see fact sheet).

k Chloride guideline Foliar damage = 100–178 mg·L⁻¹ for almond apricots and plums = 178–355 mg·L⁻¹ for grapes, peppers, potatoes, and tomatoes = 355–710 mg·L⁻¹ for alfalfa, barley, corn, and cucumbers >710 mg·L⁻¹ for cauliflower, cotton, safflower, sesame, sorghum, sugar beets, and sunflowers Rootstocks = 180–600 mg·L⁻¹ for stone fruit (peaches, plums, etc.) = 710–900 mg·L⁻¹ for grapes Cultivars = 110–180 mg·L⁻¹ for strawberries = 230–460 mg·L⁻¹ for grapes = 250 mg·L⁻¹ for boysenberries, blackberries, and raspberries

o Copper guideline = 200 µg·L⁻¹ for cereals = 1000 µg·L⁻¹ for tolerant crops

p Copper guideline = 500 µg·L⁻¹ for sheep, 1000 µg·L⁻¹ for cattle, 5000 µg·L⁻¹ for swine and poultry.

r Molybdenum guideline = 50 µg·L⁻¹ for short-term use on acidic soils

s Selenium guideline = 20 µg·L⁻¹ for continuous use = 50 µg·L⁻¹ for intermittent use

u Zinc guideline = 1000 µg·L⁻¹ when soil pH < 6.5 = 5000 µg·L⁻¹ when soil pH > 6.5

3.0 Biowaste Substrates

The following gives the list of substrates used for this project. The sewage sludge and liquid sewage wastewater were included for heavy metal comparisons. The samples were collected from various producers throughout central and southern Alberta.

1. Cattle feedlot bed pack manure
2. Dairy manure #1
3. Chicken laying hen manure
4. Chicken broiler manure
5. Potato tuber waste
6. Dairy manure #2
7. Potato process waste
8. Potato plant vines
9. Poultry slaughter waste
10. Cattle carcass
11. Sugar beet tailings
12. Grocery waste
13. Thermally hydrolyzed biowaste (#1)
14. Thermally hydrolyzed biowaste (#2)
15. Thin stillage (ethanol production from wheat)
16. Glycerine with methanol
17. Hog slaughterhouse waste; peptone
18. Hog slaughterhouse waste; sludge (DAF)
19. Wet distillers grains (ethanol production from wheat)
20. Cheese whey
21. Beef slaughterhouse waste (paunch)
22. Solid hog manure (manure with bed pack)
23. Liquid hog manure
24. Sewage sludge
25. Liquid sewage wastewater
26. Sample 1 from the ARC Pilot Plant (Dairy #1 and glycerine)
27. Sample 2 from the ARC Pilot Plant (Wet distillers grain)

4.0 Results

4.1 Methane potential using batch culture

In order to determine whether stable methanogenic conditions were reached in the batch cultures, methane potential was monitored for each substrate. Methane potential for manures depends on animal species, feed ration, manure age, manure collection methods, manure storage, foreign material incorporated in the manure and chemicals in the manure (8,9). The same conditions likely apply to other biowaste substrates. Although the biogas and methane potential results were monitored, the results are not given in this report.

The inoculum that was used for each test set was evaluated with cellulose and in each case, met the cellulose conversion requirement to methane and carbon dioxide in 7 days. The biogas and methane results showed that all the biowaste substrates in the batch cultures obtained stable methane production conditions except for the hog slaughterhouse sludge and peptone substrates.

The hog slaughterhouse substrates were tested at several VS ratios. In each case, stable methanogenic conditions were not observed. It was thought that something in the substrates was producing toxic conditions for the batch cultures. The peptone culture produced high concentrations of hydrogen sulfide gas early in the incubation period and the cultures became unstable with very little biogas production. Previous results from the ARC Pilot Plant showed that sulfide in digestion liquid can produce toxic conditions that result in decreased methane production. Sulfide toxicity is substrate, pH and temperature dependent and is related to the partitioning of hydrogen sulfide gas in the headspace to total soluble sulfide (TSS) in the liquid (personal communication with Earl Jenson, Pilot Plant operational manager). Given that the total sulphur for hog slaughterhouse peptone sample was 3% (Table 4.26), it is likely that there was a high concentration of TSS for the culture, although no measurement was taken. Dilution of the peptone substrate with inoculum did not result in stable methane production in batch culture. Information from the substrate supplier revealed that a polymer was added during the production of the hog slaughterhouse sludge and peptone, and this addition could have been toxic for methanogenic activity. No further investigation was made to reveal the exact cause for the low biogas production results.

4.2 *Chemical characteristics for biowaste substrates and batch cultures*

Because of sample matrix problems, the method for analyzing available phosphorous was changed after the start of this study and could only be done on a few select substrates. The substrates were dairy manure, chicken broiler manure, thermally hydrolyzed biowaste, wet distillers grain and inoculum.

In order to follow changes in chemical parameters and heavy metals during anaerobic digestion, each substrate for batch culture tests was analyzed as:

- Liquid substrate, diluted or undiluted depending on TS (for available heavy metals and shown in heavy metal tables as Aqueous Substrate),
- Substrate (shown in tables as Substrate or Dried Substrate)
- Initial batch culture mixture with substrate and inoculum, before incubation (shown in tables as Culture, initial),
- Final batch culture mixture with substrate and inoculum, after incubation (shown in tables as Culture, final),
- Post-digestion batch culture mixture separated into liquid and solid fractions (shown in tables as Post-digestion separation fractions, liquid or solid)

For the ARC Pilot Plant substrates, each was analyzed as:

- Initial liquid substrate mixture (for available heavy metals and shown in heavy metal tables as Aqueous mixture)
- Initial substrate mixture before digestion (shown in tables as Culture, initial)
- Final substrate after digestion (shown in table as Culture, final)
- Post-digestion mixture separated into liquid and solid fractions (shown in tables as Post-digestion separation fractions, liquid or solid)

For the ten substrate test sets used in this study, the inoculum was evaluated for quality and consistency. For each set, the analyses were done on the inoculum used for the initial culture mixture and after incubation. For all the test sets, the inoculum converted 70% or more of the cellulose within 7 days of incubation, showing good quality.

An average was taken for the Total N, Total C and TS results and the individual values for each test set parameter were compared to the average value of that test set. For the 10 inoculum samples that were used in the initial culture mixture (not incubated), the

TS results were 5% or less, the Total N results were 10% or less and the Total C results were 5% or less compared to the average result. For the inoculum that was incubated for 45 days (2 samples), the TS and Total C results were 2% and the Total N results were 6% from the average result. For the inoculum that was incubated for 30 days (8 samples), the TS results were 1% or less, the Total N results were 11% or less and the Total C results were 6% or less than the average result. The chemical parameters for the 45-day incubation final inoculum culture were obtained at the beginning of the study and the parameters for the 30-day incubation final inoculum culture were obtained close to the end of the study. The results were fairly similar except for the available NH_4^+ -N (Table 4.28). It was concluded that available NH_4^+ -N was lost on storage over a 9-month period. The loss was about 0.1%. The results from this evaluation of the inoculum showed that throughout the testing period, the inoculum was of good quality and was sufficiently consistent between collection pails.

In this report, Sections 4.2 to 4.7 and Tables 4.1 to 4.81 give results for chemical parameter and heavy metal results for substrates used in this study. Please refer to the table of contents for locations of specific results. Abbreviations that have been used in the tables are as follows: ww – wet weight; dw – dry weight; St dev – Standard deviation; BD- Below Detection for the instrument. Where no standard deviation is given for a result, there was one value reported, the other replicates were BD.

Generally, the manure substrates contained sufficient amounts of carbon, nitrogen, phosphorous and potassium for anaerobic digestion, with these nutrients remaining in the post-digestion substrates. Of course, high values of nitrogen could mean toxicity in anaerobic digestion systems, as seen for thermally hydrolyzed biowaste substrates (Tables 4.13 and 4.14), and low VS values, as seen with the laying hen chicken manure (Table 4.5), could indicate that a substrate has low organic carbon concentrations. Most non-manure substrates had good amounts of carbon, nitrogen, phosphorous and potassium. However, some non-manure substrates such as glycerine (Table 4.16) and hog slaughterhouse sludge (DAF; Table 4.24) were deficient or low in these parameters. As well, hog slaughterhouse peptone (Table 4.24) and thin stillage substrate (Table 4.15) had total sulfur values that were 1% and above, indicating that high hydrogen sulfide gas

concentrations will occur during digestion. These observations show that care must be taken when using substrates in large-scale anaerobic digestion systems.

Increased salts in soils can adversely affect the growth of many plants (10). “Soil salinity refers to the presence of the major dissolved inorganic solutes in the aqueous phase consisting of soluble and readily dissolvable salts in soil, including charged species like Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Cl^- , HCO_3^- , NO_3^- , SO_4^{2-} , CO_3^{2-} , nonionic solutes, and ions that combine to form ion pairs. Soil salinity is quantified in terms of the total concentration of the soluble salts as measured by the EC of a solution” and is traditionally measured using the saturation paste method. Another variant of this method is EC measurement on solution extracted from a 1:2 soil-water mixture (11). Using the traditional soil methods for measuring EC, an EC value of 2.0 dS/m may be considered high for salt-sensitive fruit and vegetable crops and values <4.0 dS/m are given for normal soils with pH <8.5 (10). The CCME guidelines (Table 2.2) state that soil for agricultural, parks and recreational use should have conductivity values no higher than 2 dS/m and soils for commercial and industrial use, no higher than 4 dS/m.

For the substrates used in this study, the manures generally had the highest EC results. However, the poultry slaughterhouse waste had a higher EC result than most of the manures and some non-manure vegetable/grain substrates and other non-manure substrates had surprisingly high results (See following tables). Comparatively, the vegetable/grain substrates had the lowest EC results.

The EC analysis was performed on a 2% TS slurry. This method was followed in order to be able to compare the EC results for the biowaste substrates and digestate samples used in this study. Comparison of EC results using various dilutions of one substrate cannot be done since there is no linear relationship among the dilutions. As mentioned above, the classical method for soil EC analysis uses the saturated paste method. Therefore, direct comparison of the EC results from this study to soil EC results cannot be done. The CCME soil guideline tables (6) do not mention which method was used for conductivity. One can only assume that a result for 2% TS would be lower than a result for saturated paste. We performed this comparison on a soil sample and discovered this to be true.

Given the fact that direct comparisons cannot be done, our results showed that there were high EC or salt concentrations associated with substrates used in this study, and caution must be exercised when using post-digestion products from these substrates. There are many environmental conditions that affect the concentration of salt in soils including the type of application and rate of application for organic fertilizers, soil location and topography, climate, soil type and irrigation. If a post-digestion product is applied to land without proper monitoring, salts could accumulate (11).

The EC ranges for substrates that produced methane in this study were from 1.0 dS/m for cattle carcass (Table 4.10) to 12.0 dS/m for liquid hog manure (Table 4.22). The sewage sludge substrate fell in this range at 1.3 dS/m (Table 4.21). The range for the digestates was 4.3 dS/m for cattle feedlot bed pack manure (Table 4.1) to 7.6 dS/m for chicken broiler manure (Table 4.4). For the separated fractions, the liquid ranged from 7.0 dS/m for thermally hydrolyzed biowaste #2 and glycerine (Tables 4.14 and 4.16) to 11.0 dS/m for chicken laying hen manure (Table 4.5). The solid fraction ranged from 0.6 dS/m for chicken laying hen manure (Table 4.5) to 1.7 dS/m for thermally hydrolyzed biowaste #1 (Table 4.13).

Of course, the digestate results for the substrates used in batch cultures would be affected by the inoculum EC results. For this study, the inoculum incubated for 30 days had EC values of approximately 6.1 dS/m after incubation, 8.3 dS/m for the separated liquid and 1.5 dS/m for the separated solid. The inoculum incubated for 45 days had EC values of 6.6 dS/m after incubation, 9.2 dS/m for the separated liquid and 1.0 dS/m for the separated solid (Table 4.26). The inoculum results show that salt concentrations increase as incubation times increase.

There was no attempt made to subtract the inoculum EC results from the inoculum plus substrate EC results for the biowaste batch cultures. The results from all the batch culture digestions and the Pilot Plant samples showed an increase in salt concentration after incubation. Comparison of results for the liquid and solid samples clearly showed that the salts are more concentrated in the liquid fraction of the separated digestate and that salts stay with the liquid fraction during separation.

Work at ARC has shown that salt concentrations for the separated liquid fraction can be reduced to approximately 0.2 dS/m using wastewater treatment processes. Given

that salts may accumulate with unmonitored application of biosolids to land, more work is recommended concerning different Alberta soil types and climates after repeated application of biowaste solids. It may be necessary to reduce salt concentrations in solids before land application.

Table 4. 1: Cattle feedlot bed pack manure

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	29	± 0.1	8	± 0.01	6	± 0.1	4	± 0.004	22	± 0.2
VS (% dw)	78	± 0.4	73	± 0.7	64	± 0.1	56	± 0.2	75	± 0.5
Total C (% dw)	40	± 1	36	± 2	34	± 2	34	± 1	40	± 1
Total N (% dw)	3.0	± 0.1	3.8	± 0.2	4.8	± 0.2	6.7	± 0.3	1.9	± 0.1
Total K (% dw)	2.1	± 0.1	2.7	± 0.1	3.8	± 0.1	5.5	± 0.2	1.1	± 0.03
Total P (% dw)	0.9	± 0.01	0.9	± 0.02	1.1	± 0.03	1.3	± 0.04	0.7	± 0.02
Total S (% dw)	0.5	± 0.004	0.5	± 0.01	0.6	± 0.02	0.6	± 0.03	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	9	± 0.1	16	± 0.4	34	± 1	50	± 3	7	± 0.1
Available K (mg/g dw)	19	± 0.3	26	± 1	38	± 0.4	56	± 6	8	± 0.2
Available P (mg/g dw)	Not done									
EC (dS/m)	3.0	± 0.1	3.7	± 0.4	4.3 ^a	±	9.5	± 0.1	1.2	± 0.5
pH	7.6	± 0.1	8.2	± 0.01	8.7 ^a	±	8.0	± 0.01	8.9	± 0.02

^a Only one analysis done

Table 4. 2: Dairy manure #1

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	16	± 0.1	8	± 0.2	7	± 0.3	4	± 0.004	23	± 0.1
VS (% dw)	85	± 0.2	74	± 0.6	69	± 1	57	± 0.2	84	± 0.2
Total C (% dw)	45	± 1	36	± 2	36	± 5	37	± 2	43	± 2
Total N (% dw)	3.7	± 0.1	4.3	± 0.1	4.9	± 0.1	7.2	± 0.3	2	± 0.1
Total K (% dw)	2.3	± 0.1	2.8	± 0.3	3.7	± 0.1	6.0	± 0.4	1.2	± 0.04
Total P (% dw)	0.6	± 0.02	0.9	± 0.1	0.9	± 0.04	1.4	± 0.1	0.5	± 0.003
Total S (% dw)	0.5	± 0.02	0.5	± 0.01	0.6	± 0.004	0.7	± 0.01	0.5	± 0.02
Available NH ₄ ⁺ -N (mg/g dw)	21	± 2	23	± 0.2	35	± 0.3	55	± 0.3	8	± 0.1
Available K (mg/g dw)	25	± 0.3	34	± 2	38	± 0.8	60	± 0.8	8	± 0.03
Available P (mg/g dw)	0.2	± 0.03	0.3	± 0.01	0.4	± 0	0.8	± 0.01	0.6	± 0.003
EC (dS/m)	3.6	± 0.1	4.2	± 0.4	5.8	± 0.4	9.7	± 0.1	1.2	± 0.02
pH	7.9	± 0.02	8.4	± 0	8.2	± 0.01	8.2	± 0.01	8.9	± 0.03

Table 4. 3: Potato tuber waste

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	21	± 0.1	8	± 0.03	6	± 0.01	4	± 0.002	19	± 0.1
VS (% dw)	94	± 0.1	73	± 0.04	63	± 0.3	57	± 0.2	76	± 0.2
Total C (% dw)	40	± 0.9	38	± 0.7	35	± 1	33	± 1	41	± 1
Total N (% dw)	2.0	± 0.04	3.9	± 0.1	5.2	± 0.1	6.7	± 0.2	2.0	± 0.1
Total K (% dw)	2.2	± 0.03	3.4	± 0.03	4.8	± 0.1	6.0	± 0.1	1.3	± 0.03
Total P (% dw)	0.2	± 0.01	0.8	± 0.005	1.1	± 0.02	1.3	± 0.02	1.2	± 0.02
Total S (% dw)	0.2	± 0.005	0.5	± 0.03	0.6	± 0.01	0.7	± 0.01	0.4	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	3	± 0.1	23	± 0.3	19	± 1	29	± 3	5	± 0.2
Available K (mg/g dw)	23	± 0.6	31	± 0.3	42	± 0.5	56	± 0.7	11	± 0.1
Available P (mg/g dw)	Not done									
EC (dS/m)	1.5	± 0.04	3.8	± 0.2	6.8	± 0.2	9.1	± 0.1	1.3	± 0.04
pH	5.7	± 0.04	8.5	± 0.01	8.1	± 0.01	8.1	± 0.01	9.0	± 0.01

Table 4. 4: Chicken broiler manure

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	77	± 0.03	7	± 0.05	6	± 0.01	4	± 0.004	24	± 0.2
VS (% dw)	83	± 0.09	71	± 0.3	62	± 0.08	56	± 0.1	73	± 0.5
Total C (% dw)	40	± 0.3	38	± 2	36	± 0.9	34	± 0.6	39	± 0.7
Total N (% dw)	5.0	± 0.09	4.8	± 0.1	5.9	± 0.3	8.0	± 0.2	2.3	± 0.1
Total K (% dw)	1.6	± 0.04	3.3	± 0.04	4.2	± 0.07	5.0	± 0.04	1.0	± 0.01
Total P (% dw)	2.1	± 0.02	1.3	± 0.01	2.0	± 0.02	2.0	± 0.03	2.3	± 0.03
Total S (% dw)	0.6	± 0.01	0.6	± 0.02	0.6	± 0.02	0.7	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	6	± 0.1	22	± 0.4	26	± 0.6	36	± 3	5	± 0.2
Available K (mg/g dw)	12	± 0.1	26	± 0.3	38	± 0.8	50	± 0.9	7	± 0.1
Available P (mg/g dw)	0.6	± 0.2	0.5	± 0.003	1	± 0.06	2	± 0.07	0.8	± 0.03
EC (dS/m)	1.9	± 0.01	4.0	± 0.01	7.6	± 0.1	10.1	± 0.1	1.0	± 0.03
pH	7.6	± 0.01	8.3	± 0	8.1	± 0.01	8.0	± 0.01	9.0	± 0.06

Table 4. 5: Chicken laying hen manure

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	35	± 0.5	7	± 0.2	6	± 0.2	4	± 0.02	37	± 0.2
VS (% dw)	38	± 0.7	57	± 1	49	± 2	53	± 0.5	33	± 0.6
Total C (% dw)	25	± 0.4	31	± 2	30	± 2	32	± 1	23	± 1
Total N (% dw)	4.1	± 0.2	4.5	± 0.2	5.4	± 0.2	8.6	± 0.2	1.1	± 0.07
Total K (% dw)	1.4	± 0.04	3.0	± 0.04	3.4	± 0.07	4.9	± 0.08	0.4	± 0.01
Total P (% dw)	2.6	± 0.02	1.9	± 0.01	2.7	± 0.05	2.7	± 0.04	1.8	± 0.02
Total S (% dw)	0.4	± 0.01	0.4	± 0.02	0.6	± 0.06	0.7	± 0.01	0.3 ^a	± 0.04
Available NH ₄ ⁺ -N (mg/g dw)	27	± 0.1	29	± 0.3	25	± 0.3	41	± 3	2	± 0.1
Available K (mg/g dw)	16	± 0.5	25	± 0.3	33	± 0.5	53	± 0.7	4	± 0.2
Available P (mg/g dw)	Not done									
EC (dS/m)	2.8	± 0.1	5.2	± 0.3	6.9	± 0.1	11.0	± 0.1	0.6	± 0.01
pH	8.0	± 0.01	8.3	± 0.01	8.2	± 0.01	8.1	± 0.01	8.9	± 0.03

^a Five replicate results were within 20% of the mean

Table 4. 6: Dairy manure #2

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	9	± 0.01	8	± 0.02	7	± 0.03	4	± 0.01	27	± 0.3
VS (% dw)	85	± 0.1	74	± 0.1	70	± 0.1	58	± 0.2	84	± 0.01
Total C (% dw)	45	± 1	39	± 0.8	38	± 2	34	± 0.5	43	± 2
Total N (% dw)	2.9	± 0.1	4.0	± 0.3	4.6	± 0.5	7.1	± 0.5	1.4	± 0.1
Total K (% dw)	2.3	± 0.03	3.4	± 0.04	4.1	± 0.1	6.3	± 0.1	1.1	± 0.02
Total P (% dw)	0.6	± 0.01	0.7	± 0.01	0.9	± 0.005	1.2	± 0.002	0.4	± 0.002
Total S (% dw)	0.4	± 0.02	0.5	± 0.01	0.5	± 0.01	0.6	± 0.01	0.4	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	14	± 0.04	20	± 0.2	25	± 0.5	40	± 0.1	5	± 0.2
Available K (mg/g dw)	22	± 2	27	± 2	34	± 0.5	54	± 0.4	7	± 0.3
Available P (mg/g dw)	Not done									
EC (dS/m)	4.6	± 0.1	4.9	± 0.1	6.4	± 0.1	9.6	± 0.1	1.0	± 0.03
pH	7.3	± 0.02	8.1	± 0.01	8.2	± 0.02	8.1	± 0.02	9.0	± 0.02

Table 4. 7: Potato process waste

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	52	± 1	8	± 0.04	6	± 0.1	4	± 0.004	23	± 0.3
VS (% dw)	91	± 0.3	73	± 0.1	62	± 0.2	55	± 0.03	77	± 0.2
Total C (% dw)	51	± 0.5	41	± 0.8	35	± 2	31	± 0.6	39	± 1
Total N (% dw)	0.8	± 0.04	3.6	± 0.1	5.0	± 0.2	5.7	± 0.4	1.7	± 0.1
Total K (% dw)	1.0	± 0.02	3.6	± 0.1	4.4	± 0.1	6.3	± 0.03	1.2	± 0.01
Total P (% dw)	0.1	± 0.001	0.9	± 0.01	1.0	± 0.01	1.1	± 0.01	1.2	± 0.01
Total S (% dw)	0.1	± 0.002	0.6	± 0.04	0.6	± 0.002	0.6	± 0.02	0.4	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	0.6	± 0.01	16	± 0.2	22	± 0.2	28	± 0.7	5	± 0.2
Available K (mg/g dw)	10	± 0.2	27	± 0.1	41	± 3	55	± 1	8	± 0.1
Available P (mg/g dw)	Not done									
EC (dS/m)	3.1	± 0.3	5.1	± 0.1	7.4	± 0.1	10.3	± 0.1	1.5	± 0.01
pH	5.6	± 0.2	8.3	± 0.02	8.1	± 0.03	8.2	± 0.01	8.9	± 0.04

Table 4. 8: Potato stems and leaves

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	12	± 0.02	8	± 0.01	7	± 0.02	5	± 0.001	28	± 0.1
VS (% dw)	58	± 0.1	63	± 0.1	57	± 0.1	50	± 0.2	72	± 0.2
Total C (% dw)	32	± 1	33	± 0.8	31	± 1	32	± 2	39	± 1
Total N (% dw)	2.4	± 0.1	3.7	± 0.1	4.3	± 0.2	5.6	± 0.2	1.4	± 0.1
Total K (% dw)	4.3	± 0.1	4.3	± 0.1	4.5	± 0.03	6.3	± 0.1	1.3	± 0.01
Total P (% dw)	0.1	± 0.001	0.7	± 0.01	0.8	± 0.01	1.0	± 0.01	0.5	± 0.01
Total S (% dw)	0.2	± 0.002	0.6	± 0.1	0.5	± 0.02	0.6	± 0.003	0.4	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	4	± 0.1	15	± 0.2	19	± 0.7	25	± 1	4	± 0.04
Available K (mg/g dw)	36	± 0.4	35	± 0.5	46	± 2	49	± 5	8	± 0.2
Available P (mg/g dw)	Not done									
EC (dS/m)	3.2	± 0.5	4.1	± 0.5	6.3	± 0.1	8.3	± 0.1	1.1	± 0.01
pH	6.9	± 0.05	8.2	± 0.01	8.2	± 0.01	8.2	± 0.01	9.0	± 0.04

Table 4. 9: Poultry slaughter waste

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	47	± 1	8	± 0.02	7	± 0.01	5	± 0.02	23	± 0.2
VS (% dw)	95	± 0.1	68	± 0.1	63	± 0.2	56	± 0.1	77	± 0.03
Total C (% dw)	57	± 1	36	± 0.7	36	± 1	35	± 0.4	42	± 1
Total N (% dw)	4.3	± 0.1	4.6	± 0.1	5.0	± 0.1	6.7	± 0.2	1.9	± 0.1
Total K (% dw)	0.6	± 0.01	3.7	± 0.1	4.2	± 0.1	5.7	± 0.1	1.3	± 0.02
Total P (% dw)	0.3	± 0.01	1.0	± 0.01	1.2	± 0.01	1.6	± 0.01	0.7	± 0.003
Total S (% dw)	0.4	± 0.01	0.6	± 0.003	0.7	± 0.01	0.7	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	3 ^a	± 0.4	18	± 0.2	23	± 0.6	28	± 3	6	± 0.1
Available K (mg/g dw)	4	± 0.5	36	± 2	38	± 0.7	52	± 0.6	8	± 0.02
Available P (mg/g dw)	Not done									
EC (dS/m)	6.5	± 0.5	4.7	± 0.2	6.5	± 0.1	8.6	± 0.2	1.4	± 0.04
pH	6.1	± 0.05	8.3	± 0.02	8.2	± 0.01	8.2	± 0.01	8.8	± 0.01

^a Triplicate results were within 20% of the mean

Table 4. 10: Cattle carcass

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	34	± 0.6	8	± 0.04	6	± 0.02	5	± 0.001	24	± 0.1
VS (% dw)	94	± 0.2	70	± 0.3	63	± 0.1	56	± 0.1	76	± 0.4
Total C (% dw)	59	± 4	39	± 0.7	38	± 0.9	36	± 0.7	42	± 0.9
Total N (% dw)	8.5	± 0.2	5.2	± 0.1	5.9	± 0.2	8.3	± 0.3	1.9	± 0.1
Total K (% dw)	1.1	± 0.02	3.4	± 0.03	4.2	± 0.1	6.0	± 0.1	1.2	± 0.03
Total P (% dw)	0.5	± 0.01	1.0	± 0.01	1.2	± 0.01	1.7	± 0.03	0.7	± 0.01
Total S (% dw)	0.4	± 0.04	0.6	± 0.004	0.7	± 0.01	0.7	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	1	± 0.1	15	± 0.3	32	± 0.3	46	± 7	8	± 0.1
Available K (mg/g dw)	5	± 0.7	32	± 0.4	40	± 0.8	55	± 2	8	± 0.3
Available P (mg/g dw)	Not done									
EC (dS/m)	1.0	± 0.1	4.6	± 0.1	7.4	± 0.1	10.1	± 0.2	1.4	± 0.02
pH	6.1	± 0.03	8.4	± 0.03	8.2	± 0.01	8.2	± 0.01	8.7	± 0

Table 4. 11: Sugar beet tailings

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	16	± 0.01	8	± 0.04	6	± 0.02	5	± 0.01	23	± 0.02
VS (% dw)	82	± 0.03	69	± 0.1	62	± 0.2	55	± 0.1	75	± 0.2
Total C (% dw)	40	± 0.5	37	± 0.3	36	± 0.8	35	± 0.5	41	± 1
Total N (% dw)	1.7	± 0.1	4.3	± 0.1	5.0	± 0.2	6.9	± 0.3	1.8	± 0.1
Total K (% dw)	1.3	± 0.03	3.7	± 0.03	4.8	± 0.1	5.9	± 0.1	1.2	± 0.02
Total P (% dw)	0.2	± 0.002	1.0	± 0.01	1.2	± 0.01	1.5	± 0.02	0.8	± 0.01
Total S (% dw)	0.1	± 0.004	0.6	± 0.001	0.6	± 0.004	0.7	± 0.02	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	0.2	± 0.01	16	± 0.4	21	± 0.4	26	± 0.5	6	± 0.1
Available K (mg/g dw)	10	± 1	32	± 0.7	39	± 0.3	55	± 0.2	7	± 0.1
Available P (mg/g dw)	Not done									
EC (dS/m)	1.2	± 0.03	6.2	± 0.7	5.5	± 0.1	8.6	± 0.2	1.2	± 0.01
pH	4.7	± 0.02	8.2	± 0.01	8.1	± 0.01	8.2	± 0.01	8.4	± 0.6

Table 4. 12: Grocery waste

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	16	± 0.04	8	± 0.04	6	± 0.02	5	± 0.01	22	± 0.1
VS (% dw)	95	± 0.3	70	± 0.2	63	± 0.1	56	± 0.1	76	± 0.06
Total C (% dw)	43	± 0.7	37	± 2	34	± 0.8	34	± 0.2	40	± 0.9
Total N (% dw)	1.0	± 0.04	3.9	± 0.2	5.0	± 0.1	6.5	± 0.2	1.9	± 0.08
Total K (% dw)	1.8	± 0.01	3.8	± 0.06	4.9	± 0.04	6.9	± 0.1	1.5	± 0.02
Total P (% dw)	0.2	± 0.001	1.0	± 0.01	1.1	± 0.009	1.5	± 0.02	0.9	± 0.009
Total S (% dw)	0.1	± 0.01	0.6	± 0.01	0.7	± 0.004	0.7	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	0.03	± 0.01	6	± 0.9	20	± 0.1	28	± 0.6	5	± 0.1
Available K (mg/g dw)	17	± 0.2	33	± 1	40	± 0.1	56	± 0.3	9	± 0.02
Available P (mg/g dw)	Not done									
EC (dS/m)	1.1	± 0.03	5.0	± 0.5	6.2	± 1.0	8.2	± 0.4	1.2	± 0.01
pH	3.9	± 0.02	8.2	± 0.06	8.1	± 0.01	8.3	± 0.02	8.8	± 0.03

Table 4. 13: Thermally hydrolyzed biowaste #1

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	20	± 0.06	8	± 0.01	7	± 0.01	5	± 0.01	22	± 0.04
VS (% dw)	82	± 0.08	68	± 0.1	63	± 0.05	56	± 0.1	75	± 0.2
Total C (% dw)	42	± 3	38	± 2	34	± 1	34	± 0.4	40	± 0.7
Total N (% dw)	7.2	± 0.6	4.9	± 0.2	5.7	± 0.2	7.4	± 0.4	2.1	± 0.07
Total K (% dw)	0.4	± 0.003	3.7	± 0.06	4.5	± 0.04	6.1	± 0.09	1.3	± 0.01
Total P (% dw)	2.7	± 0.01	1.3	± 0.02	1.3	± 0.01	1.8	± 0.01	1.0	± 0.007
Total S (% dw)	0.4	± 0.01	0.6	± 0.004	0.7	± 0.01	0.8	± 0.01	0.5	± 0.005
Available NH ₄ ⁺ -N (mg/g dw)	5	± 0.03	9	± 0.6	28	± 0.1	36	± 0.5	7	± 0.1
Available K (mg/g dw)	5 ^a	± 1	33	± 0.3	38	± 0.2	51	± 2	9	± 0.5
Available P (mg/g dw)	0.6	± 0.01	1	± 0.05	1	± 0.02	2	± 0.03	0.7	± 0.01
EC (dS/m)	1.3	± 0.03	4.7	± 0.05	6.7	± 0.1	8.7	± 1.1	1.7	± 0.5
pH	5.8	± 0.03	8.3	± 0.02	8.2	± 0.01	7.9	± 0.6	7.8	± 0.01

^a Triplicate results were within 25% of the mean

Table 4. 14: Thermally hydrolyzed biowaste #2

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	29	± 0.06	8	± 0.01	7	± 0.01	5	± 0.003	21	± 0.06
VS (% dw)	79	± 1	67	± 0.08	62	± 0.2	56	± 0.2	77	± 0.1
Total C (% dw)	40	± 1	37	± 0.8	35	± 0.9	33	± 0.4	40	± 0.7
Total N (% dw)	9.4	± 0.3	4.9	± 0.1	5.7	± 0.2	7.3	± 0.3	2.2	± 0.07
Total K (% dw)	0.4	± 0.002	4.0	± 0.08	4.4	± 0.05	6.0	± 0.08	1.5	± 0.03
Total P (% dw)	2.9	± 0.02	1.3	± 0.02	1.3	± 0.02	1.9	± 0.01	0.8	± 0.01
Total S (% dw)	0.4	± 0.03	0.6	± 0.006	0.7	± 0.008	0.8	± 0.01	0.5	± 0.02
Available NH ₄ ⁺ -N (mg/g dw)	17	± 0.1	19	± 0.3	26	± 0.1	31	± 2	7	± 0.1
Available K (mg/g dw)	5	± 0.5	32	± 0.5	38	± 0.5	49	± 2	10	± 0.1
Available P (mg/g dw)	0.6	± 0.03	1.2	± 0.03	1.2	± 0.01	1.7	± 0.02	0.8	± 0.02
EC (dS/m)	3.3	± 0.5	5.1	± 0.04	6.7	± 0.3	7.0	± 0.03	1.6	± 0.06
pH	6.9	± 0.01	8.3	± 0.01	8.2	± 0.01	8.3	± 0	8.8	± 0.02

Table 4. 15: Thin stillage

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	9	± 0.03	8	± 0.05	7	± 0.02	5	± 0	22	± 0.2
VS (% dw)	93	± 0.6	68	± 0.2	63	± 0.1	56	± 0.2	78	± 0.07
Total C (% dw)	46	± 0.3	36	± 0.7	34	± 1	33	± 0.4	39	± 0.7
Total N (% dw)	7.5	± 0.1	4.8	± 0.3	5.2	± 0.1	6.4	± 0.1	2.1	± 0.09
Total K (% dw)	1.4	± 0.02	4.1	± 0.08	3.9	± 0.2	5.1	± 0.2	1.2	± 0.04
Total P (% dw)	0.9	± 0.02	1.2	± 0.01	1.3	± 0.05	1.5	± 0.1	0.8	± 0.02
Total S (% dw)	1.1	± 0.03	0.7	± 0.004	0.6	± 0.01	0.7	± 0.03	0.5	± 0.002
Available NH ₄ ⁺ -N (mg/g dw)	0.2	± 0.0003	15	± 0.3	20	± 0.5	25	± 1	6	± 0.1
Available K (mg/g dw)	13	± 0.1	34	± 0.1	38	± 0.4	51	± 0.3	9	± 0.2
Available P (mg/g dw)	Not done									
EC (dS/m)	1.7	± 0.01	4.9	± 0.04	6.6	± 0.1	8.5	± 0.1	1.6	± 0.02
pH	4.1	± 0.03	8.4	± 0.01	8.2	± 0.01	8.2	± 0.02	8.6	± 0.01

Table 4. 16: Glycerine with methanol

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	80	± 0.1	8	± 0.01	7	± 0.005	5	± 0.01	22	± 0.04
VS (% dw)	91	± 0.1	70	± 0.08	62	± 0.08	55	± 0.1	78	± 0.08
Total C (% dw)	56	± 2	39	± 0.8	35	± 0.6	34	± 0.4	42	± 0.7
Total N (% dw)	BD (<0.008)		4.3	± 0.2	4.7	± 0.2	6.8	± 0.4	2.1	± 0.08
Total K (% dw)	4	± 0.04	4.8	± 0.09	5.0	± 0.04	6.5	± 0.03	1.6	± 0.02
Total P (% dw)	0.04	± 0.0006	0.9	± 0.01	1.2	± 0.005	1.4	± 0.01	0.7	± 0.008
Total S (% dw)	0.1 - 0.2 ^a	±	0.6	± 0.01	0.7	± 0.01	0.8	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	BD (<0.001)		17	± 0.2	24	± 0.3	31	± 0.1	6	± 0.2
Available K (mg/g dw)	33	± 0.4	36	± 0.6	47	± 0.7	65	± 5	11	± 0.02
Available P (mg/g dw)	Not done									
EC (dS/m)	0.8	± 0.04	5.1	± 0.04	6.2	± 0.7	7.0	± 0.09	1.3	± 0.003
pH	10.2	± 0.03	8.5	± 0.01	8.3	± 0.02	8.3	± 0.01	9.0	± 0.01

^a Sulphur result reported as a range because 5 replicates showed variability of 50% from the mean

Table 4. 17: Cheese whey

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	6	± 0.02	7	± 0.02	5	± 0.003	4	± 0.001	23	± 0.02
VS (% dw)	92	± 0.1	73	± 0.1	62	± 0.04	55	± 0.06	76	± 0.04
Total C (% dw)	40	± 0.5	35	± 1	34	± 0.9	31	± 0.2	11	± 0.3
Total N (% dw)	2.5 ^a	± 0.4	3.6	± 0.1	5.4	± 0.2	6.9	± 0.5	0.6	± 0.03
Total K (% dw)	2.6	± 0.04	2.9	± 0.07	4.8	± 0.1	6.1	± 0.1	1.1	± 0.03
Total P (% dw)	0.7	± 0.01	1.0	± 0.02	1.5	± 0.03	1.4	± 0.01	1.2	± 0.03
Total S (% dw)	0.2	± 0.004	0.5	± 0.008	0.6	± 0.01	0.7	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	2	± 0.03	16	± 0.06	25	± 1	34	± 0.6	5	± 0.3
Available K (mg/g dw)	23	± 0.5	30	± 0.4	46	± 2	66	± 2	8	± 0.07
Available P (mg/g dw)	Not done									
EC (dS/m)	2.5	± 0.01	4.6	± 0.1	6.6	± 0.3	9.7	± 0.1	1.3	± 0.06
pH	4.9	± 0.02	8.1	± 0	8.1	± 0.01	8.2	± 0.01	8.7	± 0.03

^a Triplicate results were within 25% of the mean

Table 4. 18: Wet distillers grains

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (%ww)	23	± 0.5	7	± 0.08	5	± 0.01	4	± 0.004	21	± 0.2
VS (% dw)	97	± 0.1	76	± 0.1	67	± 0.08	58	± 0.1	81	± 0.1
Total C (% dw)	50	± 3	38	± 1	36	± 0.6	33	± 1	11	± 0.1
Total N (% dw)	4.0	± 0.3	4.2	± 0.2	5.9	± 0.2	7.8	± 0.3	0.6	± 0.02
Total K (% dw)	0.4	± 0.004	2.9	± 0.07	3.8	± 0.1	5.2	± 0.1	1.0	± 0.03
Total P (% dw)	0.5	± 0.004	1.0	± 0.02	1.3	± 0.03	1.6	± 0.02	0.8	± 0.008
Total S (% dw)	0.9	± 0.06	0.6	± 0.01	0.7	± 0.007	0.8	± 0.02	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	0.2	± 0.002	14	± 0.3	29	± 1	40	± 0.8	6	± 0.2
Available K (mg/g dw)	5	± 0.1	23	± 0.07	37	± 1	51	± 0.9	7	± 0.2
Available P (mg/g dw)	3	± 0.2	1	± 0.01	2	± 0.01	2	± 0.04	0.8	± 0.02
EC (dS/m)	0.7	± 0.002	3.9	± 0.006	7.2	± 0.05	9.7	± 0.04	1.4	± 0.06
pH	4.0	± 0.03	8.0	± 0.01	8.2	± 0.02	8.3	± 0.01	8.6	± 0.02

Table 4. 19: Cattle slaughterhouse waste - paunch

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	15	± 0.1	7	± 0.004	6	± 0.04	5	± 0.002	24	± 0.01
VS (% dw)	90	± 0.1	68	± 0.05	65	± 0.2	56	± 0.08	80	± 0.04
Total C (% dw)	46	± 1	35	± 0.8	35	± 1	32	± 0.5	39	± 0.5
Total N (% dw)	1.8	± 0.06	4.2	± 0.1	5.1	± 0.1	6.7	± 0.1	1.8	± 0.04
Total K (% dw)	0.7	± 0.03	4.1	± 0.1	3.2	± 0.1	5.0	± 0.2	1.1	± 0.02
Total P (% dw)	0.6	± 0.02	1.0	± 0.02	1.1	± 0.01	1.5	± 0.03	0.7	± 0.01
Total S (% dw)	0.2	± 0.02	0.6	± 0.02	0.6	± 0.01	0.7	± 0.01	0.5	± 0.02
Available NH ₄ ⁺ -N (mg/g dw)	1	± 0.03	15	± 0.5	24	± 0.3	34	± 0.3	5	± 0.2
Available K (mg/g dw)	6	± 0.3	33	± 0.7	36	± 0.3	50	± 0.7	8	± 0.8
Available P (mg/g dw)	Not done									
EC (dS/m)	1.7	± 0.02	4.5	± 0.2	5.8	± 0.04	7.5	± 0.09	1.3	± 0.02
pH	7.2	± 0.02	8.4	± 0.01	8.2	± 0.01	8.2	± 0	8.9	± 0.01

Table 4. 20: Solid hog manure

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	29	± 0.2	8	± 0.05	6	± 0.05	4	± 0	24	± 0.06
VS (% dw)	84	± 0.1	73	± 0.3	67	± 0.01	58	± 0.01	82	± 0.03
Total C (% dw)	44	± 1	33	± 1	34	± 2	33	± 0.4	41	± 0.6
Total N (% dw)	3.5	± 0.09	4.0	± 0.06	5.2	± 0.2	7.2	± 0.3	1.9	± 0.05
Total K (% dw)	2.0	± 0.02	3.5	± 0.1	3.4	± 0.1	5.3	± 0.1	0.9	± 0.03
Total P (% dw)	1.0	± 0.01	1.5	± 0.03	1.4	± 0.01	1.9	± 0.03	0.6	± 0.01
Total S (% dw)	0.8	± 0.03	0.6	± 0.03	0.7	± 0.02	0.8	± 0.01	0.6	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	6	± 0.4	15	± 2	23	± 0.4	34	± 0.04	5	± 0.08
Available K (mg/g dw)	16	± 1	28	± 0.3	34	± 0.6	51	± 2	7	± 0.4
Available P (mg/g dw)	Not done									
EC (dS/m)	2.5	± 0.1	3.9	± 0.3	5.2	± 0.2	7.4	± 0.2	1.1	± 0.05
pH	8.4	± 0.05	8.3	± 0.02	8.2	± 0	8.3	± 0.01	9.0	± 0.03

Table 4. 21: Sewage sludge

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	8	± 0.2	7	± 0.02	6	± 0.02	5	± 0.01	27	± 0.02
VS (% dw)	72	± 0.3	69	± 0.2	61	± 0.1	55	± 0.01	78	± 0.06
Total C (% dw)	41 ^a	± 3	35	± 1	32	± 0.6	31	± 0.5	38	± 0.4
Total N (% dw)	3.4	± 0.3	4.1	± 0.09	5.3	± 0.2	6.6	± 0.2	1.7	± 0.04
Total K (% dw)	0.3	± 0.01	3.3	± 0.07	3.1	± 0.1	3.6	± 0.1	0.8	± 0.03
Total P (% dw)	1.5	± 0.02	1.1	± 0.02	1.5	± 0.02	1.8	± 0.03	0.9	± 0.02
Total S (% dw)	0.7	± 0.03	0.6	± 0.02	0.7	± 0.01	0.8	± 0.02	0.5	± 0.02
Available NH ₄ ⁺ -N (mg/g dw)	5	± 0.3	14	± 0.5	25	± 0.4	34	± 0.2	4	± 0.2
Available K (mg/g dw)	1	± 0.02	24	± 0.6	29	± 0.4	40	± 1	5 ^a	± 0.9
Available P (mg/g dw)	Not done									
EC (dS/m)	1.3	± 0.02	3.7	± 0.2	5.3	± 0.04	6.7	± 0.06	0.9	± 0.004
pH	5.6	± 0.01	7.9	± 0.02	8.2	± 0.01	8.2	± 0.01	8.8	± 0.01

^a The triplicate values were within 20% of the mean

Table 4. 22: Liquid hog

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	2	± 0.01	5	± 0.003	5	± 0.01	4	± 0.004	24	± 0.1
VS (% dw)	73	± 0.2	68	± 0.1	63	± 0.1	56	± 0.02	79	± 0.1
Total C (% dw)	46	± 3	37	± 1	34	± 0.8	32	± 0.5	39	± 0.5
Total N (% dw)	12.1	± 0.5	5.1	± 0.2	6.4	± 0.2	7.9	± 0.2	1.8	± 0.1
Total K (% dw)	4.7	± 0.1	3.7	± 0.1	4.2	± 0.1	5.8	± 0.2	0.9	± 0.02
Total P (% dw)	2.5	± 0.1	1.2	± 0.02	1.4	± 0.02	1.8	± 0.04	0.8	± 0.01
Total S (% dw)	0.8	± 0.02	0.6	± 0.01	0.7	± 0.02	0.8	± 0.02	0.5	± 0.03
Available NH ₄ ⁺ -N (mg/g dw)	54	± 2	28	± 0.4	34	± 0.9	46	± 0.7	5	± 0.7
Available K (mg/g dw)	45	± 0.7	36	± 0.6	43	± 2	55	± 0.2	6	± 0.4
Available P (mg/g dw)	Not done									
EC (dS/m)	12.0 ^a	± 0.3	5.6	± 0.03	7.4	± 0.2	9.3	± 0.5	1.2	± 0.003
pH	7.8 ^a	± 0.02	8.4	± 0.02	8.1	± 0	8.3	± 0	8.8	± 0.01

^a Performed on undiluted substrate

Table 4. 23: Liquid sewage

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate		Culture				Post-digestion separation fractions			
			Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
TS (% ww)	0.1 ^a	± 0	5	± 0.02	4	± 0.01	3	± 0.01	23	± 0.04
VS (% dw)	45	± 2	67	± 0.1	63	± 0.05	56	± 0.1	79	± 0.1
Total C (% dw)	BD(<0.005)	±	33	± 1	33	± 1	30	± 0.6	38	± 0.5
Total N (% dw)	0-20 ^b	±	4.2	± 0.2	5.2	± 0.2	6.5	± 0.4	1.4	± 0.05
Total K (% dw)	0.9	± 0.03	3.8	± 0.07	3.7	± 0.1	5.7	± 0.2	0.9	± 0.02
Total P (% dw)	0.7	± 0.01	1.1	± 0.02	1.2	± 0.02	1.6	± 0.02	0.6	± 0.01
Total S (% dw)	3.3	± 0.1	0.6	± 0.02	0.7	± 0.02	0.8	± 0.01	0.5	± 0.03
Available NH ₄ ⁺ -N (mg/g dw)	35	± 0.7	22	± 0.4	26	± 0.6	36	± 0.7	4	± 0.3
Available K (mg/g dw)	11	± 0.4	35	± 1	39	± 0.7	55	± 0.9	6	± 0.1
Available P (mg/g dw)	Not done									
EC (dS/m)	1.3 ^c	± 0.03	4.8	± 0.06	5.9	± 0.2	8.4	± 0.03	1.0	± 0.006
pH	7.0 ^c	± 0.03	8.4	± 0.02	8.0	± 0.01	8.1	± 0	8.8	± 0.02

^a This results would be more accurate if measured using the COD method. This was not done for this study ^b The substrate was analyzed 20 times for TN. Only a range could be obtained. ^c Performed on undiluted substrate

Table 4. 24: Hog slaughterhouse waste

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Substrate only, no culture mixtures analyzed			
	Hog slaughterhouse peptone		Hog slaughterhouse sludge (DAF)	
	Mean	Std dev	Mean	Std dev
TS (% ww)	18	± 0.3	20	± 0.04
VS (% dw)	80	± 2	90	± 0.7
Total C (% dw)	44	± 0.2	60	± 1
Total N (% dw)	9.0	± 0.05	6.8	± 0.06
Total K (% dw)	1.4	± 0.03	0.03	± 0.0004
Total P (% dw)	1.0	± 0.007	0.3	± 0.002
Total S (% dw)	3.0	± 0.03	0.7	± 0.02
Available NH ₄ ⁺ -N (mg/g dw)	2	± 0.2	2	± 0.01
Available K (mg/g dw)	11	± 0.2	0.2	± 0.0006
Available P (mg/g dw)	Not done			
EC (dS/m)	3.7	± 0.04	0.6	± 0.01
pH	8.1	± 0.01	4.8	± 0.06

4.3 Chemical characteristics for substrates used in the ARC Pilot Plant

Table 4. 25: ARC Pilot Plant substrates

Parameters, refer to Table 2.1 and Section 4.2 for explanations	Culture				Post culture separation			
	Initial		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
ARC Pilot Plant (1) mixture								
TS (% ww)	14	± 0.05	12	± 0.08	4	± 0.01	29	± 0.2
VS (% dw)	88	± 1	83	± 0.3	62	± 0.4	90	± 0.2
Total C (% dw)	52	± 0.9	47	± 4	38	± 2	49	± 1
Total N (% dw)	1.6	± 0.07	2.5	± 0.2	5.5	± 0.4	1.0 ^a	± 0.2
Total K (% dw)	3.2	± 0.05	4.5	± 0.1	12	± 0.2	1.6	± 0.04
Total P (% dw)	0.3	± 0.003	0.5	± 0.003	0.9	± 0.01	0.3	± 0.004
Total S (% dw)	0.2	± 0.01	0.3	± 0.01	0.6	± 0.003	0.2	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	6	± 0.2	5	± 0.01	2	± 0.08	2	± 0.07
Available K (mg/g dw)	31	± 0.2	41	± 1	15	± 0.4	13	± 0.3
Available P (mg/g dw)	Not done							
EC (µS/cm)	2.7	± 0.08	3.0	± 0.06	8.2	± 0.05	1.0	± 0.05
pH	8.0	± 0.05	8.6	± 0.02	8.3	± 0.01	9.4	± 0.02
ARC Pilot Plant (2) mixture								
TS (% ww)	6	± 0.03	6	± 0.2	1	± 0.01	26	± 0.2
VS (% dw)	96	± 0.01	89	± 0.2	76	± 0.2	91	± 0.3
Total C (% dw)	50	± 2	36	± 5	49	± 0.9	47	± 1
Total N (% dw)	3.9	± 0.2	4.3	± 0.1	16.7	± 0.8	1.7	± 0.06
Total K (% dw)	0.7	± 0.01	0.9	± 0.01	4.0	± 0.03	0.2	± 0.002
Total P (% dw)	0.5	± 0.004	1.0	± 0.007	1.4	± 0.009	0.9	± 0.007
Total S (% dw)	0.7	± 0.06	0.6	± 0.04	1.0	± 0.03	0.4	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	0.1	± 0.007	21	± 0.3	88	± 17	4	± 0.2
Available K (mg/g dw)	6	± 0.2	8	± 0.2	36	± 1	1	± 0.01
Available P (mg/g dw)	Not done							
EC (µS/cm)	0.9	± 0.01	4.0	± 0.3	12.0 ^b	± 0.03	0.7	± 0.04
pH	3.7	± 0.02	7.9	± 0.02	8.0 ^b	± 0.02	8.4	± 0.03

^a Triplicate results were within 30% of the mean. ^b Performed on undiluted substrate

4.4 Chemical characteristics for inoculum used in this study

Table 4. 26: Inoculum

Parameters, refer to Table 2.1 and Section 4.2 for explanations	After collection and before incubation		Culture				Post-digestion separation fractions			
			Initial (after 7 d incubation) ^a		Final		Liquid		Solid	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Held for 44 days										
TS (% ww)	8	± 0.01	7	± 0.02	6	± 0.05	5	± 0.03	23	± 0.3
VS (% dw)	69	± 0.01	67	± 0.3	62	± 0.5	56	± 0.5	76	± 0.1
Total C (% dw)	36	± 0.4	36	± 0.9	37	± 1	33	± 0.6	40	± 1.1
Total N (% dw)	4.5	± 0.1	4.8	± 0.2	5.0	± 0.2	6.9	± 0.4	1.9	± 0.07
Total K (% dw)	3.4	± 0.1	3.6	± 0.1	4.6	± 0.08	6.6	± 0.07	1.5	± 0.02
Total P (% dw)	1.1	± 0.02	1.3	± 0.02	1.1	± 0.02	1.4	± 0.01	0.7	± 0.01
Total S (% dw)	0.6	± 0.01	0.6	± 0.01	0.6	± 0.04	0.7	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)	26	± 3	24	± 0.9	35	± 0.4	49	± 0.8	8	± 0.05
Available K (mg/g dw)	33	± 0.8	39	± 4	41	± 0.2	56	± 0.5	9	± 0.2
Available P (mg/g dw)	1	± 0.1	1	± 0.01	1	±	1	± 0.04	0.7	± 0.01
EC (µS/cm)	5.7	± 0.03	4.2	± 0.4	6.6	± 0.06	9.2	± 0.06	1.0	± 0.01
pH	8.0	± 0.03	8.5	± 0.02	8.5	± 0.01	8.2	± 0.01	8.9	± 0.03
Held for 30 days										
TS (% ww)	Not done				7	± 0.01	5	± 0.01	24	± 0.1
VS (% dw)					63	± 0.08	55	± 0.07	76	± 0.1
Total C (% dw)					35	± 0.9	32	± 0.5	39	± 1.0
Total N (% dw)					5.0	± 0.2	5.8	± 0.1	1.9	± 0.1
Total K (% dw)					4.8	± 0.04	5.8	± 0.1	1.5	± 0.02
Total P (% dw)					1.1	± 0.01	1.5	± 0.01	0.7	± 0.01
Total S (% dw)					0.7	± 0.01	0.7	± 0.01	0.5	± 0.01
Available NH ₄ ⁺ -N (mg/g dw)					17 ^b	± 0.7	20	± 1	4	± 0.1
Available K (mg/g dw)					42	± 0.7	55	± 1	9	± 0.2
Available P (mg/g dw)					1	± 0.1	2	± 0.03	1	± 0.03
EC (µS/cm)					6.1	± 0.06	8.3	± 0.02	1.5	± 0.02
pH					8.3	± 0.01	8.3	± 0	8.9	± 0.02

^a The 7 day incubation was done before the inoculum was used for the culture mixture ^b There appeared to be a loss of available NH₄-N with storage

4.5 *Heavy metals for biowaste substrates and batch cultures, please refer to Section 4.2 for explanations*

Table 4. 27: Heavy metals for cattle feedlot bed pack manure

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	BD	±		0.0193	± 0.00081
Al	13.7	±	0.50	2290	± 36
As	0.230	±	0.012	0.623	± 0.019
B	6.09	±	0.37	17.5	± 0.63
Ba	1.11	±	0.019	56.2	± 1.7
Be	BD	±		0.0787	± 0.0084
Bi	0.00218	±	0.00040	0.0253	± 0.00040
Ca	1090	±	27	19600	± 420
Cd	0.00463	±	0.00019	0.0858	± 0.0044
Cl	9050	±	400	21800	± 690
Co	0.754	±	0.026	2.93	± 0.065
Cr	0.497	±	0.012	28.5	± 0.45
Cu	1.32	±	0.047	26.9	± 0.82
Fe	93.3	±	1.7	2650	± 76
Hg	0.0121	±		0.163	± 0.0074
Li	1.34	±	0.054	2.43	± 0.078
Mg	483	±	35	5740	± 140
Mn	3.47	±	0.15	153	± 2.5
Mo	0.548	±	0.046	4.67	± 0.17
Na	4390	±	200	5420	± 170
Ni	1.18	±	0.028	28.1	± 0.79
Pb	0.032	±	0.0020	0.932	± 0.018
Sb	0.0127	±	0.00099	0.0191	± 0.00083
Se	0.111	±	0.0063	0.178	± 0.048
Sn	BD	±		0.0318	± 0.0012
Sr	3.40	±	0.14	32.8	± 1.4
Th	0.00492	±	0.00072	0.334	± 0.010
Ti	6.27	±	0.042	71.0	± 2.4
Tl	BD	±		0.0439	± 0.0011
U	0.00560	±	0.00025	0.214	± 0.0052
V	0.252	±	0.0038	5.38	± 0.13
Zn	7.10	±	0.31	177	± 3.8

Table 4. 28: Heavy metals for cattle feedlot bed pack manure culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0656	± 0.0028	0.111	± 0.0021	0.115	± 0.0025	0.0460	± 0.0014
Al	2560	± 15	6530	± 22	8970	± 120	3690	± 14
As	1.03	± 0.0091	1.79	± 0.011	2.22	± 0.035	0.865	± 0.0064
B	21.3	± 0.46	31.2	± 0.39	35.1	± 0.92	14.4	± 0.28
Ba	71.3	± 0.98	96.9	± 1.2	125	± 1.6	52.4	± 0.67
Be	0.129	± 0.0078	0.145	± 0.0093	0.162	± 0.0095	0.0786	± 0.0067
Bi	0.0463	± 0.0017	0.0308	± 0.00076	0.0373	± 0.0010	0.0169	± 0.00023
Ca	21100	± 590	27200	± 430	36500	± 630	18900	± 220
Cd	0.0969	± 0.0028	0.156	± 0.0031	0.193	± 0.0031	0.0877	± 0.0019
Cl	27800	± 930	14200	± 240	33000	± 1100	11800	± 200
Co	3.60	± 0.050	5.17	± 0.035	5.78	± 0.049	2.40	± 0.020
Cr	14.5	± 0.24	35.7	± 0.27	52.0	± 0.28	29.4	± 0.20
Cu	32.3	± 0.79	53.3	± 0.44	63.7	± 0.63	28.2	± 0.31
Fe	2900	± 100	4850	± 32	6250	± 25	3390	± 21
Hg	0.109	± 0.0020	0.0704	± 0.0023	0.0818	± 0.0023	0.0617	± 0.0017
Li	3.29	± 0.078	6.12	± 0.064	7.72	± 0.10	2.60	± 0.031
Mg	6870	± 75	7480	± 49	10000	± 74	5690	± 47
Mn	179	± 3.0	253	± 1.2	332	± 1.6	139	± 0.87
Mo	3.19	± 0.050	6.87	± 0.076	8.96	± 0.082	4.62	± 0.050
Na	7840	± 120	11300	± 83	14400	± 140	2360	± 25
Ni	17.8	± 0.38	28.0	± 0.25	38.1	± 0.29	20.6	± 0.19
Pb	1.69	± 0.0095	1.99	± 0.0097	2.70	± 0.016	1.21	± 0.0071
Sb	0.0368	± 0.0019	0.0548	± 0.00041	0.0494	± 0.00061	0.0282	± 0.0015
Se	0.272	± 0.082	1.33	± 0.052	1.55	± 0.099	0.823	± 0.067
Sn	0.0668	± 0.0035	0.393	± 0.0041	0.282	± 0.0020	0.0631	± 0.0014
Sr	84.7	± 2.6	136	± 1.8	172	± 3.4	69.2	± 0.65
Th	0.309	± 0.0090	0.923	± 0.0092	1.08	± 0.012	0.434	± 0.0042
Ti	129	± 14	223	± 3.1	284	± 5.9	136	± 3.1
Tl	0.0709	± 0.0016	0.0694	± 0.0010	0.0947	± 0.0010	0.0389	± 0.00059
U	0.349	± 0.0037	0.284	± 0.0033	0.369	± 0.0042	0.157	± 0.0020
V	5.47	± 0.070	13.8	± 0.13	17.6	± 0.14	7.26	± 0.077
Zn	180	± 5.1	306	± 2.0	389	± 7.6	174	± 2.1

Table 4. 29: Heavy metals for dairy manure #1

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	BD	±		0.0179	± 0.00064
Al	15.7	±	0.48	962	± 15
As	BD	±		0.244	± 0.0062
B	15.0	±	0.51	34.5	± 1.0
Ba	2.07	±	0.0084	50.0	± 0.88
Be	0.00158	±		0.0539	± 0.0070
Bi	0.00887	±	0.0066	0.0300	± 0.0032
Ca	2820	±	16	20400	± 760
Cd	0.00749	±	0.00098	0.139	± 0.0044
Cl	6160	±	46	12200	± 780
Co	0.993	±	0.011	2.76	± 0.047
Cr	0.486	±	0.0020	2.83	± 0.58
Cu	3.12	±	0.080	58.7	± 1.8
Fe	99.3	±	1.5	1190	± 47
Hg	BD	±		0.0110	± 0.00083
Li	2.12	±	0.11	2.46	± 0.065
Mg	1310	±	16	6150	± 87
Mn	8.86	±	0.19	222	± 2.5
Mo	0.498	±	0.013	1.77	± 0.043
Na	3470	±	35	4050	± 42
Ni	1.25	±	0.026	3.54	± 0.11
Pb	0.0241	±	0.0015	0.498	± 0.014
Sb	0.0131	±	0.0018	0.0492	± 0.0016
Se	BD	±		0.500	± 0.060
Sn	BD	±		0.0942	± 0.00057
Sr	17.5	±	0.18	75.0	± 2.6
Th	0.0140	±	0.0091	0.194	± 0.0037
Ti	2.82	±	0.065	41.0	± 1.6
Tl	0.00410	±	0.0013	0.111	± 0.0039
U	0.785	±	0.0078	3.21	± 0.91
V	0.566	±	0.0069	3.43	± 0.075
Zn	11.5	±	0.54	265	± 8.7

Table 4. 30: Heavy metals for dairy manure #1 culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0875	± 0.0020	0.0754	± 0.0016	0.133	± 0.0023	0.0580	± 0.00093
Al	3450	± 53	4270	± 15	8420	± 65	2500	± 11
As	1.07	± 0.029	1.35	± 0.015	2.09	± 0.023	0.726	± 0.0061
B	30.7	± 1.1	31.4	± 0.57	42.6	± 0.82	18.1	± 0.56
Ba	77.6	± 1.9	70.3	± 0.72	124	± 1.4	39.5	± 0.52
Be	0.114	± 0.010	0.108	± 0.0081	0.180	± 0.014	0.0607	± 0.0034
Bi	0.0406	± 0.0019	0.0276	± 0.00075	0.0383	± 0.00090	0.0131	± 0.00049
Ca	23600	± 710	22600	± 250	40100	± 260	13900	± 140
Cd	0.145	± 0.0031	0.159	± 0.0036	0.229	± 0.0029	0.0909	± 0.0018
Cl	34500	± 1100	18100	± 520	25800	± 480	5240	± 300
Co	4.31	± 0.092	4.03	± 0.048	5.73	± 0.061	2.26	± 0.018
Cr	5.74	± 0.092	7.17	± 0.044	13.9	± 0.053	5.04	± 0.014
Cu	54.1	± 1.3	60.5	± 0.66	93.4	± 0.66	35.5	± 0.31
Fe	2970	± 65	3180	± 25	5650	± 51	1950	± 11
Hg	0.0192	± 0.0012	0.169	± 0.00095	0.0274	± 0.0014	0.0129	± 0.0014
Li	4.10	± 0.079	4.86	± 0.096	8.05	± 0.10	2.06	± 0.030
Mg	7870	± 84	6850	± 26	11800	± 83	3950	± 21
Mn	232	± 2.1	230	± 0.83	389	± 2.7	130	± 0.77
Mo	2.21	± 0.048	2.72	± 0.038	4.24	± 0.037	1.44	± 0.017
Na	7600	± 102	8380	± 51	14000	± 67	2260	± 20
Ni	10.1	± 0.25	8.57	± 0.13	13.3	± 0.14	4.21	± 0.056
Pb	1.98	± 0.0095	1.65	± 0.0067	2.87	± 0.010	0.925	± 0.0051
Sb	0.0280	± 0.0017	0.433	± 0.0010	0.0532	± 0.0012	0.0324	± 0.0013
Se	0.461	± 0.076	1.19	± 0.048	1.65	± 0.030	0.770	± 0.039
Sn	0.0705	± 0.0027	0.0826	± 0.0019	0.305	± 0.0037	0.320	± 0.0039
Sr	128	± 3.3	136	± 1.7	227	± 3.1	73.0	± 0.88
Th	0.575	± 0.011	0.530	± 0.0064	1.08	± 0.0091	0.367	± 0.0041
Ti	145	± 7.4	162	± 2.6	301	± 3.4	116	± 1.9
Tl	0.129	± 0.0038	0.0760	± 0.0012	0.136	± 0.0018	0.0474	± 0.00091
U	2.25	± 0.051	1.29	± 0.020	2.05	± 0.026	0.713	± 0.0083
V	7.91	± 0.17	9.67	± 0.083	16.5	± 0.13	5.48	± 0.043
Zn	240	± 6.5	274	± 2.8	453	± 6.3	173	± 1.7

Table 4. 31: Heavy metals for potato tuber waste

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	BD	±		0.0103	± 0.00026
Al	0.838	±	0.079	117	± 2.7
As	BD	±		0.0247	± 0.0041
B	3.52	±	0.17	7.05	± 0.16
Ba	0.448	±	0.035	2.66	± 0.047
Be	BD	±		0.00750	± 0.0037
Bi	0.000560	±	0.00035	0.00371	± 0.00031
Ca	69.4	±	0.83	363	± 10
Cd	0.164	±	0.0025	0.452	± 0.015
Cl	2490	±	100	11700	± 290
Co	0.0650	±	0.0048	0.180	± 0.0069
Cr	0.0701	±	0.022	0.501	± 0.040
Cu	1.27	±	0.079	2.66	± 0.068
Fe	5.80	±	0.57	173	± 1.1
Hg	0.00345	±		0.00386	± 0.00079
Li	0.190	±	0.042	0.124	± 0.0044
Mg	634	±	7.9	1340	± 21
Mn	3.26	±	0.055	10.1	± 0.070
Mo	0.0594	±	0.0017	0.177	± 0.0070
Na	37.0	±	64	214	± 4.6
Ni	0.343	±	0.011	0.854	± 0.028
Pb	0.0490	±		0.0505	± 0.0012
Sb	0.00126	±	0.00059	0.00458	± 0.00070
Se	BD	±		BD	±
Sn	BD	±		0.00458	± 0.00081
Sr	0.275	±	0.0059	1.16	± 0.043
Th	0.0297	±	0.0083	0.0276	± 0.00077
Ti	3.26	±	0.20	10.9	± 0.64
Tl	0.0212	±	0.00072	0.00920	± 0.00065
U	0.000887	±	0.000090	0.0149	± 0.00071
V	0.0197	±	0.0070	0.287	± 0.012
Zn	6.97	±	0.22	11.0	± 0.29

Table 4. 32: Heavy metals for potato tuber waste culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0647	± 0.0011	0.105	± 0.00094	0.153	± 0.0032	0.0527	± 0.0012
Al	5960	± 28	5960	± 103	8280	± 160	3030	± 50
As	1.28	± 0.025	1.80	± 0.022	2.08	± 0.033	0.834	± 0.0085
B	18.6	± 0.30	24.8	± 0.61	28.5	± 1.3	11.9	± 0.37
Ba	71.9	± 0.63	97.7	± 0.98	129	± 2.8	48.4	± 0.60
Be	0.102	± 0.0083	0.113	± 0.0041	0.147	± 0.0054	0.0574	± 0.0046
Bi	0.0253	± 0.00045	0.0406	± 0.00043	0.0650	± 0.0018	0.0182	± 0.00035
Ca	19700	± 190	25300	± 750	31200	± 610	13500	± 350
Cd	0.222	± 0.0039	0.331	± 0.0046	0.389	± 0.0072	0.157	± 0.0018
Cl	15500	± 380	23400	± 380	21900	± 930	19000	± 290
Co	2.94	± 0.014	3.79	± 0.025	4.27	± 0.069	2.16	± 0.031
Cr	7.77	± 0.11	7.80	± 0.12	11.2	± 0.21	4.82	± 0.065
Cu	32.9	± 0.24	39.9	± 0.54	47.0	± 1.1	20.8	± 0.47
Fe	3940	± 12	4700	± 56	6340	± 85	2770	± 31
Hg	0.158	± 0.00086	0.0238	± 0.00070	0.0350	± 0.0017	0.0119	± 0.00094
Li	4.46	± 0.049	5.11	± 0.067	6.72	± 0.18	2.12	± 0.032
Mg	5940	± 44	4510	± 72	5850	± 190	6080	± 41
Mn	199	± 1.1	257	± 3.6	318	± 5.7	118	± 0.92
Mo	1.99	± 0.015	2.98	± 0.049	3.24	± 0.073	1.42	± 0.022
Na	6350	± 63	9360	± 190	11800	± 220	2280	± 46
Ni	7.75	± 0.074	9.87	± 0.061	12.0	± 0.23	4.78	± 0.063
Pb	1.87	± 0.012	2.51	± 0.0073	3.42	± 0.085	1.30	± 0.0063
Sb	0.0306	± 0.0011	0.0439	± 0.00071	0.0478	± 0.0020	0.0252	± 0.00075
Se	0.841	± 0.045	0.919	± 0.061	1.26	± 0.036	0.586	± 0.026
Sn	0.0894	± 0.0019	0.0814	± 0.0017	0.190	± 0.0030	BD	±
Sr	130	± 1.2	170	± 1.7	200	± 4.9	87.9	± 1.9
Th	0.717	± 0.0048	0.776	± 0.0042	1.34	± 0.039	0.210	± 0.0018
Ti	214	± 1.9	230	± 4.7	330	± 4.1	135	± 3.5
Tl	0.0651	± 0.0015	0.0809	± 0.0012	0.113	± 0.0036	0.0441	± 0.00059
U	0.263	± 0.0029	0.310	± 0.0025	0.433	± 0.013	0.154	± 0.0020
V	10.0	± 0.086	10.9	± 0.19	14.5	± 0.35	5.76	± 0.11
Zn	167	± 1.5	265	± 6.0	322	± 2.8	114	± 2.3

Table 4. 33: Heavy metals for chicken broiler manure

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	BD	±		0.00747	± 0.00063
Al	0.358	±	0.032	197	± 2.1
As	BD	±		0.0468	± 0.012
B	8.72	±	0.28	27.5	± 0.93
Ba	0.299	±	0.017	44.9	± 1.2
Be	BD	±		0.0101	± 0.0028
Bi	BD	±		0.00405	± 0.00046
Ca	892	±	49	34000	± 960
Cd	0.0130	±	0.00095	0.107	± 0.0040
Cl	8940	±	300	22500	± 700
Co	0.129	±	0.0051	0.492	± 0.011
Cr	0.147	±		10.9	± 1.2
Cu	127	±	5.4	322	± 9.3
Fe	29.5	±	1.8	579	± 20
Hg	BD	±		0.0617	± 0.0051
Li	0.789	±	0.030	1.02	± 0.030
Mg	762	±	62	5850	± 130
Mn	3.48	±	0.33	366	± 1.8
Mo	0.480	±	0.051	4.21	± 0.089
Na	3500	±	110	4410	± 110
Ni	2.76	±	0.070	14.7	± 0.39
Pb	0.00341	±	0.0050	0.184	± 0.0035
Sb	0.00572	±	0.0015	0.121	± 0.0028
Se	0.660	±	0.021	1.20	± 0.14
Sn	BD	±		0.0304	± 0.0010
Sr	1.26	±	0.059	38.7	± 0.92
Th	BD	±		0.0532	± 0.0014
Ti	2.70	±	0.27	68.4	± 3.1
Tl	0.00808	±	0.00048	0.122	± 0.0069
U	0.000230	±	0.000096	0.0591	± 0.0013
V	0.0120	±	0.0020	0.729	± 0.025
Zn	23.5	±	0.77	397	± 11

Table 4. 34: Heavy metals for chicken broiler manure culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0743	± 0.0017	0.0665	± 0.0016	0.0724	± 0.0025	0.0381	± 0.0014
Al	5030	± 23	5450	± 110	5740	± 88	2640	± 34
As	1.20	± 0.012	1.45	± 0.019	1.82	± 0.021	0.734	± 0.016
B	25.0	± 0.53	25.4	± 0.87	27.9	± 0.81	17.9	± 0.34
Ba	75.6	± 0.82	96.9	± 1.3	113	± 1.1	62.7	± 0.59
Be	0.0902	± 0.0041	0.0927	± 0.0034	0.0985	± 0.0060	0.0496	± 0.0037
Bi	0.0220	± 0.00060	0.0331	± 0.00081	0.0367	± 0.0032	0.0179	± 0.00033
Ca	27900	± 200	37500	± 720	38400	± 700	37900	± 680
Cd	0.136	± 0.0027	0.170	± 0.0017	0.217	± 0.0033	0.0953	± 0.0030
Cl	6410	± 210	32600	± 690	41900	± 1300	22000	± 260
Co	2.87	± 0.023	2.94	± 0.020	3.52	± 0.031	2.12	± 0.024
Cr	18.6	± 0.062	16.1	± 0.19	17.1	± 0.19	9.84	± 0.11
Cu	139	± 1.2	141	± 2.0	162	± 1.6	103	± 1.2
Fe	3470	± 14	4380	± 33	3800	± 50	2400	± 20
Hg	0.0236	± 0.00091	0.0367	± 0.0019	0.0344	± 0.0017	0.0218	± 0.00046
Li	4.11	± 0.041	4.91	± 0.088	4.99	± 0.092	2.18	± 0.043
Mg	6990	± 24	6550	± 120	7010	± 120	6070	± 88
Mn	293	± 1.4	422	± 2.0	440	± 3.3	246	± 1.3
Mo	4.35	± 0.042	4.77	± 0.073	4.84	± 0.061	2.80	± 0.028
Na	7670	± 64	9810	± 250	12300	± 300	2540	± 43
Ni	16.7	± 0.16	15.1	± 0.16	16.7	± 0.20	9.31	± 0.14
Pb	1.64	± 0.0064	2.19	± 0.018	2.55	± 0.015	1.21	± 0.0056
Sb	0.0603	± 0.0010	0.0503	± 0.00051	0.0517	± 0.0013	0.0341	± 0.0010
Se	1.24	± 0.089	1.46	± 0.060	1.83	± 0.052	1.46	± 0.026
Sn	0.267	± 0.0021	BD	±	BD	±	BD	±
Sr	126	± 1.5	155	± 2.9	184	± 2.6	99.4	± 0.96
Th	0.758	± 0.0083	0.108	± 0.0016	0.0373	± 0.00070	0.0156	± 0.00038
Ti	220	± 1.9	133	± 2.7	83.7	± 1.3	90.9	± 1.9
Tl	0.0854	± 0.0018	0.107	± 0.0013	0.107	± 0.0021	0.0923	± 0.0013
U	0.226	± 0.0013	0.262	± 0.0034	0.296	± 0.0017	0.135	± 0.0021
V	9.04	± 0.059	9.25	± 0.13	10.1	± 0.15	5.08	± 0.052
Zn	289	± 2.4	404	± 10	488	± 8.8	181	± 1.9

Table 4. 35: Heavy metals for chicken laying hen manure

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	BD	±		0.0190	± 0.0010
Al	1.82	±	0.60	323	± 19
As	0.0146	±		0.331	± 0.020
B	13.3	±	0.91	28.6	± 0.51
Ba	0.527	±	0.096	30.2	± 0.39
Be	BD	±		0.0643	± 0.012
Bi	0.00242	±	0.00079	0.00731	± 0.00088
Ca	245	±	42	230000	± 5500
Cd	0.0197	±	0.012	0.251	± 0.0069
Cl	4060	±	200	15100	± 160
Co	0.380	±	0.031	1.65	± 0.046
Cr	0.386	±	0.067	6.60	± 0.13
Cu	4.14	±	0.37	43.8	± 0.67
Fe	85.9	±	11	1110	± 98
Hg	0.00641	±		0.00662	± 0.00091
Li	0.508	±	0.060	0.807	± 0.014
Mg	24.7	±	5.5	7640	± 130
Mn	2.08	±	0.38	481	± 7.9
Mo	0.846	±	0.048	3.14	± 0.070
Na	BD	±		2080	± 30
Ni	2.34	±	0.30	11.9	± 0.38
Pb	0.0634	±	0.043	0.633	± 0.016
Sb	0.0109	±	0.0022	0.123	± 0.0034
Se	0.560	±	0.11	2.10	± 0.16
Sn	BD	±		0.0799	± 0.0024
Sr	0.881	±	0.18	132	± 3.5
Th	0.00241	±	0.0012	0.193	± 0.0071
Ti	12.9	±	0.80	71.7	± 3.3
Tl	0.00333	±	0.00022	0.0559	± 0.0011
U	0.0200	±	0.0012	0.376	± 0.017
V	0.0762	±	0.0084	2.04	± 0.031
Zn	35.7	±	5.5	485	± 12

Table 4. 36: Heavy metals for chicken laying hen manure culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0481	± 0.00057	0.0626	± 0.00055	0.0774	± 0.0026	0.0184	± 0.00041
Al	3640	± 26	3960	± 88	3850	± 62	886	± 16
As	1.05	± 0.012	1.24	± 0.018	1.57	± 0.028	0.390	± 0.033
B	27.5	± 0.55	28.3	± 0.30	33.9	± 0.91	8.43	± 0.22
Ba	63.3	± 0.57	81.4	± 0.90	94.5	± 0.74	39.4	± 0.36
Be	0.0732	± 0.0057	0.0780	± 0.0053	0.0967	± 0.0073	0.0486	± 0.0041
Bi	0.0198	± 0.00062	0.0271	± 0.00014	0.0361	± 0.00082	0.00828	± 0.00034
Ca	83700	± 1300	75900	± 870	55800	± 760	281000	± 4800
Cd	0.178	± 0.0040	0.250	± 0.0030	0.293	± 0.0050	0.143	± 0.0017
Cl	10600	± 130	36400	± 520	70400	± 2100	14500	± 330
Co	2.81	± 0.016	2.95	± 0.041	3.65	± 0.034	0.830	± 0.019
Cr	6.80	± 0.017	7.13	± 0.043	7.74	± 0.10	3.71	± 0.077
Cu	50.4	± 0.32	57.6	± 0.71	73.2	± 0.74	10.7	± 0.18
Fe	2730	± 9.2	3450	± 48	3570	± 60	1430	± 48
Hg	0.0129	± 0.00088	0.0171	± 0.00053	0.0249	± 0.0021	0.00964	± 0.00049
Li	3.24	± 0.020	4.04	± 0.059	4.05	± 0.048	1.03	± 0.019
Mg	6670	± 24	6850	± 73	7800	± 94	4410	± 110
Mn	382	± 1.3	513	± 5.1	578	± 4.6	303	± 2.9
Mo	3.46	± 0.029	4.14	± 0.042	5.03	± 0.052	0.923	± 0.012
Na	6070	± 44	6240	± 120	9310	± 150	986	± 23
Ni	7.50	± 0.072	8.18	± 0.15	11.0	± 0.17	3.34	± 0.43
Pb	1.54	± 0.0031	2.00	± 0.0050	2.59	± 0.010	0.968	± 0.018
Sb	0.0623	± 0.0010	0.0652	± 0.0010	0.0700	± 0.00082	0.0492	± 0.0015
Se	1.82	± 0.061	2.14	± 0.046	2.47	± 0.033	1.40	± 0.065
Sn	0.0789	± 0.0011	BD	±	BD	±	BD	±
Sr	126	± 1.2	149	± 1.8	158	± 1.7	144	± 0.86
Th	0.443	± 0.0037	0.0836	± 0.0010	0.0437	± 0.0011	0.0379	± 0.0014
Ti	181	± 1.1	128	± 1.8	114	± 2.5	70.9	± 2.1
Tl	0.0694	± 0.00073	0.0911	± 0.00075	0.104	± 0.0016	0.0357	± 0.00080
U	0.352	± 0.0024	0.374	± 0.0058	0.435	± 0.0024	0.488	± 0.0070
V	7.28	± 0.039	7.49	± 0.069	7.56	± 0.14	2.66	± 0.042
Zn	354	± 1.9	576	± 7.3	694	± 11	94.1	± 0.78

Table 4. 37: Heavy metals for dairy manure #2

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000911	±	0.00035	0.0307	±	0.0013
Al	6.90	±	0.17	394	±	8.0
As	0.0911	±	0.0090	0.246	±	0.0067
B	9.10	±	0.32	18.4	±	0.72
Ba	5.75	±	0.059	56.1	±	0.44
Be	0.000959	±	0.000018	0.0425	±	0.0042
Bi	0.000792	±	0.0064	0.0102	±	0.00024
Ca	2940	±	1.4	18000	±	300
Cd	0.0109	±	0.00055	0.293	±	0.0038
Cl	10400	±	190	22800	±	570
Co	0.410	±	0.00019	1.23	±	0.013
Cr	0.666	±	0.024	6.83	±	0.088
Cu	3.01	±	0.055	87.4	±	1.3
Fe	30.2	±	0.55	923	±	28
Hg	0.0109	±	0.000037	0.0224	±	0.00091
Li	0.738	±	0.075	1.07	±	0.020
Mg	3210	±	69	5220	±	90
Mn	6.56	±	0.011	144	±	1.2
Mo	0.182	±	0.0012	1.82	±	0.023
Na	8910	±	200	9370	±	240
Ni	0.884	±	0.0018	4.15	±	0.039
Pb	0.0136	±	0.0023	0.285	±	0.0011
Sb	0.00590	±	0.00032	0.0337	±	0.0010
Se	0.168	±	0.0026	0.614	±	0.020
Sn	0.00558	±	0.0038	0.0329	±	0.0011
Sr	19.8	±	0.22	56.5	±	0.63
Th	0.00212	±	0.00036	0.0756	±	0.00068
Ti	2.51	±	0.044	25.6	±	0.40
Tl	0.00233	±	0.00047	0.0390	±	0.0012
U	0.216	±	0.0086	0.778	±	0.0072
V	0.273	±	0.0084	1.81	±	0.028
Zn	5.93	±	0.11	121	±	1.8

Table 4. 38: Heavy metals for dairy manure #2 culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0483	± 0.00079	0.142	± 0.0027	0.197	± 0.0046	0.0767	± 0.0030
Al	3780	± 29	6560	± 81	8970	± 100	3600	± 55
As	1.20	± 0.019	1.37	± 0.030	1.88	± 0.020	0.647	± 0.0072
B	19.8	± 0.59	26.3	± 0.59	31.5	± 0.54	14.4	± 0.24
Ba	75.4	± 1.2	114	± 1.1	157	± 1.6	62.8	± 0.90
Be	0.0779	± 0.012	0.110	± 0.0057	0.140	± 0.010	0.0669	± 0.011
Bi	0.0364	± 0.0012	0.0291	± 0.00093	0.0383	± 0.0011	0.155	± 0.00024
Ca	23100	± 230	20000	± 260	26700	± 320	11100	± 140
Cd	0.199	± 0.0035	0.245	± 0.0053	0.307	± 0.0030	0.121	± 0.0047
Cl	22100	± 700	29300	± 550	36000	± 550	20800	± 87
Co	2.90	± 0.030	3.26	± 0.022	3.99	± 0.027	1.79	± 0.019
Cr	8.27	± 0.066	10.8	± 0.097	15.4	± 0.14	5.87	± 0.046
Cu	63.2	± 0.60	78.4	± 0.82	90.7	± 0.90	36.6	± 0.46
Fe	3100	± 25	3560	± 49	4720	± 73	2150	± 21
Hg	0.0223	± 0.0016	0.0239	± 0.0020	0.0329	± 0.0025	0.0125	± 0.0012
Li	3.48	± 0.067	3.80	± 0.032	5.38	± 0.095	1.53	± 0.045
Mg	8000	± 93	7670	± 92	10400	± 140	4090	± 45
Mn	213	± 1.6	222	± 0.92	333	± 1.6	115	± 0.50
Mo	2.39	± 0.028	2.95	± 0.028	3.86	± 0.048	1.35	± 0.017
Na	10700	± 180	11900	± 190	18500	± 250	2790	± 41
Ni	7.87	± 0.14	9.36	± 0.097	12.6	± 0.14	4.20	± 0.044
Pb	1.65	± 0.012	2.17	± 0.0097	3.13	± 0.026	1.20	± 0.0026
Sb	0.0282	± 0.00070	0.139	± 0.0040	0.167	± 0.0013	0.0665	± 0.0023
Se	1.07	± 0.043	1.18	± 0.089	1.81	± 0.087	0.781	± 0.089
Sn	0.0750	± 0.0067	0.373	± 0.0041	0.478	± 0.0068	0.205	± 0.0033
Sr	125	± 1.6	158	± 1.4	219	± 1.5	73.6	± 0.66
Th	0.523	± 0.0056	0.981	± 0.0051	1.31	± 0.018	0.483	± 0.0065
Ti	165	± 1.6	317	± 3.4	422	± 3.8	190	± 2.8
Tl	0.0670	± 0.00024	0.0915	± 0.0018	0.118	± 0.0025	0.0498	± 0.0014
U	0.652	± 0.0096	0.718	± 0.0092	0.960	± 0.019	0.365	± 0.0038
V	7.85	± 0.062	9.71	± 0.082	12.9	± 0.15	5.20	± 0.021
Zn	166	± 1.5	187	± 1.3	343	± 4.0	98.9	± 0.87

Table 4. 39: Heavy metals for potato process waste

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000565	±	0.00010	0.00378	±	0.00055
Al	1.55	±	0.16	47.6	±	0.49
As	BD	±		0.0605	±	0.024
B	2.38	±	0.081	3.38	±	0.15
Ba	0.913	±	0.013	1.95	±	0.025
Be	BD	±		BD	±	
Bi	0.000578	±	0.00034	0.00185	±	0.00019
Ca	158	±	1.6	264	±	2.9
Cd	0.0479	±	0.0015	0.0654	±	0.0035
Cl	49700	±	760	72600	±	1500
Co	0.0368	±	0.00077	0.0408	±	0.0015
Cr	0.551	±	0.22	1.39	±	0.15
Cu	0.467	±	0.034	1.32	±	0.022
Fe	3.85	±	0.17	24.0	±	0.81
Hg	0.00464	±	0.0014	0.0110	±	0.00090
Li	BD	±		0.0654	±	0.0047
Mg	567	±	6.5	612	±	3.9
Mn	2.98	±	0.13	3.10	±	0.016
Mo	0.0793	±	0.0014	0.122	±	0.0066
Na	27600	±	1800	28300	±	310
Ni	0.0804	±	0.018	0.180	±	0.0068
Pb	0.0533	±	0.083	0.0246	±	0.00034
Sb	0.000446	±	0.000075	0.00282	±	0.00067
Se	0.0274	±	0.014	0.238	±	0.12
Sn	0.00221	±	0.00034	0.00767	±	0.00079
Sr	0.612	±	0.0077	1.19	±	0.0097
Th	0.0134	±	0.0028	0.0137	±	0.0021
Ti	2.74	±	0.31	4.57	±	0.065
Tl	0.00115	±	0.000031	0.00322	±	0.00041
U	0.00278	±	0.0020	0.00313	±	0.00014
V	0.113	±	0.062	0.364	±	0.036
Zn	4.52	±	0.062	4.91	±	0.034

Table 4. 40: Heavy metals for potato process waste culture
 Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.151	± 0.0021	0.0660	± 0.0016	0.113	± 0.0020	0.0455	± 0.00061
Al	7410	± 110	5920	± 65	9190	± 23	4020	± 220
As	1.48	± 0.018	1.85	± 0.029	2.39	± 0.0097	0.838	± 0.0089
B	21.1	± 0.44	24.6	± 0.25	30.6	± 0.33	12.2	± 0.22
Ba	106	± 1.5	85.6	± 0.38	110	± 0.58	56.2	± 0.18
Be	0.131	± 0.0047	0.115	± 0.018	0.155	± 0.0092	0.0451	± 0.0052
Bi	0.0329	± 0.00045	0.0321	± 0.00029	0.0426	± 0.00051	0.0181	± 0.00014
Ca	17500	± 300	24600	± 79	29500	± 160	16700	± 64
Cd	0.141	± 0.0042	0.167	± 0.0017	0.210	± 0.0020	0.0840	± 0.0025
Cl	47600	± 1100	40400	± 1310	24100	± 290	12300	± 220
Co	3.08	± 0.032	3.96	± 0.023	4.86	± 0.032	1.93	± 0.016
Cr	11.9	± 0.12	12.9	± 0.11	14.2	± 0.054	4.84	± 0.035
Cu	33.3	± 0.43	41.2	± 0.22	49.7	± 0.13	21.9	± 0.069
Fe	3760	± 11	4300	± 42	5670	± 48	2970	± 20
Hg	0.0210	± 0.00037	0.0203	± 0.0013	0.0283	± 0.0019	0.0120	± 0.0012
Li	4.43	± 0.084	4.95	± 0.039	7.01	± 0.077	2.36	± 0.020
Mg	6540	± 85	8040	± 55	8170	± 29	10400	± 22
Mn	197	± 1.3	292	± 1.5	357	± 1.6	150	± 0.66
Mo	2.26	± 0.035	2.70	± 0.017	3.47	± 0.019	1.38	± 0.0060
Na	19300	± 350	20700	± 220	29500	± 190	4610	± 13
Ni	9.83	± 0.14	11.8	± 0.13	14.0	± 0.093	5.00	± 0.021
Pb	2.40	± 0.015	2.50	± 0.015	3.04	± 0.0073	1.43	± 0.0039
Sb	0.119	± 0.0032	0.0382	± 0.0014	0.0489	± 0.0020	0.0277	± 0.00079
Se	1.08	± 0.040	0.826	± 0.057	1.23	± 0.040	0.582	± 0.068
Sn	0.376	± 0.0031	0.104	± 0.0025	0.249	± 0.0035	0.0857	± 0.0010
Sr	151	± 2.0	196	± 1.3	227	± 0.97	150	± 0.35
Th	1.13	± 0.010	0.542	± 0.0051	1.22	± 0.0033	0.350	± 0.0035
Ti	359	± 12	223	± 0.85	347	± 0.87	180	± 0.53
Tl	0.0879	± 0.0017	0.0726	± 0.00083	0.0979	± 0.00086	0.0478	± 0.00077
U	0.359	± 0.0050	0.278	± 0.0027	0.366	± 0.0045	0.156	± 0.0015
V	11.4	± 0.15	10.6	± 0.047	15.7	± 0.025	6.27	± 0.027
Zn	169	± 1.7	219	± 0.95	251	± 1.3	119	± 0.29

Table 4. 41: Heavy metals for potato stems and leaves

Heavy metal	Results given as $\mu\text{g/g dw}$				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	0.00215	\pm	0.00016	0.119	\pm 0.0037
Al	7.75	\pm	0.45	14600	\pm 190
As	0.349	\pm	0.0050	2.49	\pm 0.025
B	10.5	\pm	0.081	35.5	\pm 0.96
Ba	20.7	\pm	0.12	315	\pm 2.5
Be	0.000797	\pm	0.00033	0.373	\pm 0.025
Bi	0.00354	\pm	0.0015	0.0503	\pm 0.00055
Ca	1520	\pm	8.1	24300	\pm 160
Cd	0.119	\pm	0.00087	1.14	\pm 0.0097
Cl	5520	\pm	31	30100	\pm 120
Co	0.410	\pm	0.0060	2.95	\pm 0.0076
Cr	1.11	\pm	0.012	147	\pm 0.89
Cu	1.21	\pm	0.0090	12.6	\pm 0.082
Fe	71.2	\pm	0.20	7300	\pm 73
Hg	0.0252	\pm	0.0012	1.94	\pm 0.026
Li	0.411	\pm	0.0099	7.18	\pm 0.13
Mg	6.41	\pm	0.050	13700	\pm 81
Mn	11.8	\pm	0.13	199	\pm 0.72
Mo	0.231	\pm	0.0076	16.4	\pm 0.13
Na	170	\pm	2.0	2650	\pm 27
Ni	2.62	\pm	0.023	97.9	\pm 0.52
Pb	0.130	\pm	0.0012	4.57	\pm 0.034
Sb	0.0251	\pm	0.00060	0.240	\pm 0.0032
Se	0.0419	\pm	0.0042	0.567	\pm 0.053
Sn	0.00401	\pm	0.00031	0.431	\pm 0.0030
Sr	14.0	\pm	0.076	97.2	\pm 0.64
Th	0.0668	\pm	0.024	2.14	\pm 0.042
Ti	2.17	\pm	0.083	439	\pm 18
Tl	0.00824	\pm	0.00028	0.183	\pm 0.0026
U	0.0390	\pm	0.00053	0.590	\pm 0.0082
V	0.321	\pm	0.0040	21.3	\pm 0.41
Zn	1.60	\pm	0.037	30.3	\pm 0.16

Table 4. 42: Heavy metals for potato stems and leaves culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.159	± 0.0015	0.0753	± 0.0029	0.0950	± 0.0023	0.0705	± 0.0016
Al	11800	± 110	7580	± 20	8750	± 450	5620	± 17
As	2.19	± 0.023	2.24	± 0.029	2.70	± 0.031	1.28	± 0.028
B	29.9	± 0.63	29.2	± 0.31	36.7	± 0.61	19.8	± 0.63
Ba	204	± 2.0	150	± 0.43	180	± 0.72	95.2	± 0.56
Be	0.260	± 0.0090	0.176	± 0.015	0.201	± 0.014	0.114	± 0.0091
Bi	0.0409	± 0.00082	0.0417	± 0.00075	0.0481	± 0.00082	0.0329	± 0.0016
Ca	23500	± 300	29800	± 37	36900	± 260	18800	± 54
Cd	0.535	± 0.0078	0.608	± 0.0041	0.712	± 0.010	0.347	± 0.0056
Cl	30600	± 1200	17500	± 560	38200	± 1400	2920	± 29
Co	3.55	± 0.048	3.90	± 0.012	4.56	± 0.020	2.38	± 0.0050
Cr	59.0	± 0.26	61.4	± 0.27	49.2	± 0.21	32.6	± 0.12
Cu	30.4	± 0.23	34.9	± 0.11	42.3	± 0.38	20.8	± 0.081
Fe	5800	± 25	5970	± 24	6910	± 27	4260	± 53
Hg	0.358	± 0.0061	0.211	± 0.0026	0.249	± 0.0051	0.226	± 0.0051
Li	5.93	± 0.096	5.97	± 0.037	7.68	± 0.087	3.26	± 0.024
Mg	10600	± 65	12800	± 27	17400	± 84	7090	± 9.9
Mn	232	± 1.8	324	± 1.2	382	± 1.7	156	± 0.97
Mo	7.58	± 0.047	5.19	± 0.018	5.81	± 0.061	4.28	± 0.037
Na	7580	± 62	5690	± 25	8650	± 64	1360	± 5.3
Ni	40.3	± 0.41	33.9	± 0.15	35.5	± 0.17	23.3	± 0.17
Pb	3.45	± 0.038	3.08	± 0.0074	3.90	± 0.027	1.98	± 0.0060
Sb	0.181	± 0.00083	0.0493	± 0.00091	0.101	± 0.0011	0.0435	± 0.0015
Se	1.18	± 0.065	0.755	± 0.12	0.923	± 0.074	0.566	± 0.056
Sn	0.438	± 0.0043	0.0536	± 0.0013	0.0894	± 0.0020	0.127	± 0.0019
Sr	162	± 1.2	181	± 0.65	216	± 1.3	93.6	± 0.62
Th	1.49	± 0.028	0.643	± 0.0031	0.799	± 0.0047	0.759	± 0.0049
Ti	426	± 11	198	± 0.59	209	± 1.5	176	± 0.92
Tl	0.131	± 0.0015	0.0994	± 0.0015	0.123	± 0.0018	0.0686	± 0.00038
U	0.448	± 0.0073	0.296	± 0.0017	0.371	± 0.0032	0.189	± 0.0014
V	17.7	± 0.18	14.4	± 0.13	16.5	± 0.15	10.5	± 0.077
Zn	141	± 0.94	158	± 0.49	183	± 1.3	107	± 0.26

Table 4. 43: Heavy metals for poultry slaughter waste

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000684	±	0.00016	0.00930	±	0.00040
Al	0.573	±	0.074	175	±	1.7
As	0.00957	±	0.00026	0.0231	±	0.0012
B	0.321	±	0.058	0.900	±	0.051
Ba	0.0345	±	0.0074	1.12	±	0.015
Be	0.000290	±	0.00050	0.00110	±	0.00070
Bi	0.000498	±	0.00011	0.00410	±	0.00030
Ca	62.8	±	11	603	±	7.0
Cd	0.0233	±	0.0049	0.0522	±	0.0014
Cl	1950	±	198	5410	±	180
Co	0.0316	±	0.0052	0.0834	±	0.0012
Cr	0.102	±	0.0062	2.42	±	0.037
Cu	6.82	±	1.2	12.4	±	0.20
Fe	15.2	±	2.2	236	±	8.6
Hg	BD	±		0.00540	±	0.00070
Li	0.174	±	0.065	0.703	±	0.019
Mg	119	±	21	475	±	4.2
Mn	0.817	±	0.16	6.24	±	0.042
Mo	0.0986	±	0.019	0.314	±	0.0059
Na	1570	±	200	2870	±	41
Ni	0.0560	±	0.017	1.11	±	0.012
Pb	0.00567	±	0.0016	0.949	±	0.00090
Sb	0.000325	±	0.00012	0.00220	±	0.00020
Se	0.456	±	0.076	1.11	±	0.032
Sn	0.00390	±	0.00091	0.0185	±	0.00040
Sr	0.636	±	0.13	5.50	±	0.092
Th	0.0289	±	0.010	0.0427	±	0.0015
Ti	8.27	±	1.3	22.9	±	0.50
Tl	0.00910	±	0.0022	0.0161	±	0.00040
U	0.0000619	±	0.000052	0.00890	±	0.00020
V	0.0226	±	0.0024	0.159	±	0.0030
Zn	15.5	±	2.7	62.2	±	0.82

Table 4. 44: Heavy metals for poultry slaughter waste culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0901	± 0.0030	0.110	± 0.0026	0.175	± 0.0019	0.0737	± 0.0014
Al	6520	± 35	7070	± 240	11800	± 140	4330	± 41
As	1.62	± 0.014	1.97	± 0.017	2.66	± 0.036	0.984	± 0.017
B	21.1	± 0.61	25.6	± 0.63	32.2	± 0.57	16.4	± 0.32
Ba	82.1	± 1.0	102	± 1.7	147	± 1.0	56.5	± 0.62
Be	0.101	± 0.011	0.123	± 0.012	0.199	± 0.0086	0.0875	± 0.0058
Bi	0.0309	± 0.00040	0.0428	± 0.0013	0.0638	± 0.00090	0.0220	± 0.00030
Ca	23700	± 350	29100	± 360	40200	± 680	17200	± 370
Cd	0.134	± 0.0024	0.155	± 0.0037	0.216	± 0.0032	0.0839	± 0.0015
Cl	15000	± 260	21100	± 310	17800	± 400	2390	± 38
Co	3.49	± 0.043	4.20	± 0.068	5.40	± 0.054	2.60	± 0.020
Cr	12.8	± 0.24	14.0	± 0.065	15.7	± 0.17	6.30	± 0.072
Cu	39.3	± 0.44	45.9	± 0.75	62.4	± 0.42	26.6	± 0.43
Fe	4410	± 37	4960	± 48	6490	± 51	3310	± 30
Hg	0.0183	± 0.0019	0.0232	± 0.0016	0.0295	± 0.00090	0.0105	± 0.00030
Li	5.00	± 0.075	5.23	± 0.13	8.22	± 0.26	2.97	± 0.043
Mg	8310	± 110	8420	± 49	11500	± 140	5150	± 61
Mn	273	± 2.8	277	± 1.3	375	± 4.3	159	± 1.0
Mo	2.49	± 0.024	2.95	± 0.058	4.02	± 0.027	1.74	± 0.025
Na	8030	± 110	9340	± 79	13000	± 310	1810	± 34
Ni	11.1	± 0.15	13.0	± 0.27	16.0	± 0.13	6.74	± 0.060
Pb	2.51	± 0.033	2.79	± 0.014	3.65	± 0.030	2.16	± 0.022
Sb	0.0375	± 0.0011	0.0305	± 0.0013	0.0368	± 0.0013	0.0247	± 0.00060
Se	0.905	± 0.073	1.06	± 0.060	1.45	± 0.064	0.831	± 0.041
Sn	0.0917	± 0.0027	0.186	± 0.0047	0.261	± 0.0018	0.167	± 0.0015
Sr	186	± 2.1	188	± 2.8	252	± 4.7	102	± 1.7
Th	0.550	± 0.0071	0.842	± 0.012	1.32	± 0.0095	0.680	± 0.0034
Ti	245	± 3.8	268	± 11	354	± 7.8	194	± 2.6
Tl	0.0728	± 0.0015	0.0914	± 0.0016	0.140	± 0.0018	0.0584	± 0.0011
U	0.259	± 0.0029	0.303	± 0.0066	0.434	± 0.0040	0.165	± 0.0013
V	10.9	± 0.13	12.0	± 0.14	17.0	± 0.24	7.33	± 0.14
Zn	188	± 2.4	250	± 2.2	327	± 8.1	135	± 3.1

Table 4. 45: Heavy metals for cattle carcass

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000176	±	0.00013	0.0106	±	0.00030
Al	0.131	±	0.034	612	±	4.4
As	0.0104	±	0.0039	0.0936	±	0.0036
B	0.204	±	0.043	1.69	±	0.089
Ba	0.0402	±	0.0042	5.83	±	0.099
Be	BD	±		0.0127	±	0.0024
Bi	0.000208	±	0.000087	0.00300	±	0.00030
Ca	105	±	12	3680	±	62
Cd	0.000546	±	0.00013	0.00910	±	0.00090
Cl	1610	±	96	2450	±	42
Co	0.00768	±	0.0015	0.165	±	0.0021
Cr	0.0700	±	0.00070	2.85	±	0.042
Cu	0.327	±	0.064	6.19	±	0.12
Fe	11.8	±	2.3	496	±	14
Hg	BD	±		0.00410	±	0.00070
Li	0.234	±	0.011	0.522	±	0.024
Mg	175	±	21	955	±	14
Mn	0.677	±	0.096	11.0	±	0.093
Mo	0.0277	±	0.0063	0.224	±	0.0036
Na	1240	±	110	2980	±	60
Ni	BD	±		1.08	±	0.026
Pb	0.00194	±	0.00042	0.534	±	0.0026
Sb	0.00103	±	0.00038	0.0112	±	0.00050
Se	0.0550	±	0.017	0.467	±	0.047
Sn	0.00114	±	0.00017	0.0340	±	0.00090
Sr	1.40	±	0.16	13.3	±	0.20
Th	0.0146	±	0.00084	0.118	±	0.0052
Ti	6.69	±	0.64	45.4	±	1.2
Tl	0.000163	±	0.000077	0.00570	±	0.00020
U	BD	±		0.0230	±	0.00050
V	0.0217	±	0.0016	0.710	±	0.014
Zn	5.89	±	1.1	127	±	1.7

Table 4. 46: Heavy metals for cattle carcass culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0816	± 0.0031	0.111	± 0.0044	0.127	± 0.0065	0.0779	± 0.0026
Al	5050	± 42	7310	± 110	10200	± 54	4580	± 44
As	1.54	± 0.025	1.90	± 0.021	2.64	± 0.034	0.911	± 0.0063
B	20.1	± 0.34	26.7	± 0.61	29.4	± 0.74	15.0	± 0.29
Ba	73.4	± 0.61	109	± 1.3	136	± 1.2	59.4	± 0.63
Be	0.109	± 0.0076	0.129	± 0.0016	0.181	± 0.011	0.0980	± 0.0038
Bi	0.0297	± 0.00070	0.0439	± 0.00060	0.0539	± 0.0010	0.0259	± 0.00050
Ca	22500	± 230	29400	± 350	39300	± 470	17900	± 290
Cd	0.123	± 0.0017	0.158	± 0.0044	0.206	± 0.0047	0.103	± 0.0028
Cl	22100	± 430	15900	± 240	52500	± 1400	3080	± 45
Co	3.33	± 0.038	4.46	± 0.041	4.95	± 0.039	2.59	± 0.035
Cr	14.5	± 0.20	15.1	± 0.18	12.7	± 0.15	6.20	± 0.068
Cu	35.5	± 0.40	47.6	± 0.51	59.0	± 0.57	27.5	± 0.33
Fe	3950	± 24	5120	± 39	6070	± 82	3860	± 88
Hg	0.0185	± 0.0015	0.0207	± 0.0010	0.0264	± 0.00050	0.0146	± 0.0015
Li	4.41	± 0.045	5.77	± 0.12	7.17	± 0.12	2.91	± 0.029
Mg	7600	± 51	8340	± 82	11900	± 80	4880	± 41
Mn	257	± 2.0	273	± 1.3	377	± 3.1	165	± 1.0
Mo	2.39	± 0.035	3.13	± 0.049	3.70	± 0.038	1.73	± 0.024
Na	7620	± 71	9560	± 170	13000	± 190	2360	± 43
Ni	11.7	± 0.13	14.0	± 0.15	14.3	± 0.087	6.45	± 0.095
Pb	18.5	± 0.12	2.76	± 0.016	3.55	± 0.024	1.57	± 0.0043
Sb	0.0839	± 0.00070	0.0326	± 0.00050	0.0296	± 0.0011	0.0250	± 0.00090
Se	0.814	± 0.061	1.14	± 0.075	0.933	± 0.072	0.678	± 0.039
Sn	0.0768	± 0.0037	0.250	± 0.0027	0.176	± 0.0042	0.193	± 0.0018
Sr	174	± 1.4	186	± 3.2	248	± 4.9	111	± 2.5
Th	0.369	± 0.0052	1.21	± 0.014	1.03	± 0.0062	0.573	± 0.0057
Ti	185	± 1.9	294	± 4.9	283	± 6.3	212	± 13
Tl	0.0642	± 0.00070	0.0977	± 0.00070	0.125	± 0.0029	0.0594	± 0.00090
U	0.245	± 0.0024	0.338	± 0.0040	0.415	± 0.0043	0.182	± 0.0027
V	8.87	± 0.081	12.7	± 0.13	15.0	± 0.17	7.54	± 0.063
Zn	187	± 1.7	255	± 4.6	341	± 7.2	172	± 4.1

Table 4. 47: Heavy metals for sugar beet tailings

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000738	±	0.000072	0.0237	±	0.0013
Al	9.37	±	1.6	2710	±	18
As	0.0845	±	0.018	1.25	±	0.032
B	2.27	±	0.29	23.3	±	0.31
Ba	5.69	±	0.81	62.2	±	1.0
Be	0.00348	±	0.00052	0.124	±	0.0093
Bi	0.000509	±	0.00010	0.0174	±	0.00050
Ca	2080	±	330	9590	±	89
Cd	0.0254	±	0.0049	0.218	±	0.0027
Cl	1690	±	270	6060	±	170
Co	0.408	±	0.065	1.79	±	0.032
Cr	0.388	±	0.070	22.4	±	0.49
Cu	1.17	±	0.14	10.2	±	0.12
Fe	4.06	±	0.59	3180	±	54
Hg	BD	±		0.00680	±	0.00070
Li	0.450	±	0.082	2.88	±	0.064
Mg	1310	±	340	3680	±	43
Mn	29.8	±	4.6	111	±	1.3
Mo	0.0417	±	0.0070	0.667	±	0.0093
Na	3230	±	510	5080	±	73
Ni	0.710	±	0.110	10.8	±	0.18
Pb	0.0153	±	0.0025	1.14	±	0.018
Sb	0.0107	±	0.0017	0.0345	±	0.0011
Se	0.0371	±	0.0082	0.203	±	0.024
Sn	0.00313	±	0.00044	0.0688	±	0.00090
Sr	3.55	±	0.52	28.6	±	0.27
Th	0.0327	±	0.0068	0.737	±	0.039
Ti	4.57	±	0.83	53.6	±	0.37
Tl	0.0169	±	0.0025	0.0563	±	0.0012
U	0.0183	±	0.0021	0.167	±	0.0042
V	0.197	±	0.012	8.19	±	0.080
Zn	6.54	±	1.1	24.3	±	0.29

Table 4. 48: Heavy metals for sugar beet tailings culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.104	± 0.0023	0.138	± 0.0047	0.162	± 0.0029	0.0670	± 0.0013
Al	7070	± 47	9670	± 420	12600	± 70	4390	± 28
As	1.74	± 0.023	2.16	± 0.040	2.63	± 0.025	1.04	± 0.026
B	24.8	± 0.34	30.0	± 0.42	33.0	± 1.0	16.1	± 0.20
Ba	90.2	± 0.51	122	± 1.4	149	± 2.1	59.9	± 0.58
Be	0.121	± 0.0086	0.141	± 0.0076	0.202	± 0.011	0.0876	± 0.0049
Bi	0.0319	± 0.00030	0.0466	± 0.0013	0.0590	± 0.0012	0.0227	± 0.00050
Ca	24800	± 110	30700	± 550	39500	± 600	17200	± 210
Cd	0.161	± 0.0019	0.205	± 0.0031	0.247	± 0.0033	0.104	± 0.0010
Cl	14500	± 110	5310	± 86	13400	± 170	2060	± 46
Co	3.67	± 0.023	4.73	± 0.035	5.32	± 0.055	2.50	± 0.020
Cr	13.6	± 0.12	22.4	± 0.19	17.6	± 0.17	8.09	± 0.16
Cu	37.6	± 0.21	46.8	± 0.45	57.7	± 0.72	25.0	± 0.26
Fe	4890	± 38	6140	± 62	6860	± 130	3580	± 63
Hg	0.0169	± 0.0012	0.0247	± 0.0018	0.0269	± 0.0014	0.00830	± 0.00040
Li	5.33	± 0.062	6.32	± 0.16	7.70	± 0.25	2.89	± 0.033
Mg	8760	± 10	8490	± 100	11100	± 130	5940	± 62
Mn	287	± 1.2	300	± 1.8	384	± 5.1	169	± 1.8
Mo	2.46	± 0.018	3.23	± 0.036	3.88	± 0.063	1.70	± 0.023
Na	8300	± 28	11100	± 230	13500	± 60	2470	± 210
Ni	11.4	± 0.071	17.1	± 0.16	16.1	± 0.23	6.96	± 0.066
Pb	2.40	± 0.0062	2.86	± 0.022	3.71	± 0.035	1.78	± 0.011
Sb	0.0418	± 0.0012	0.0429	± 0.0011	0.043	± 0.0020	0.0229	± 0.00060
Se	1.01	± 0.044	1.27	± 0.045	1.37	± 0.041	0.749	± 0.021
Sn	0.134	± 0.0025	0.406	± 0.0056	0.319	± 0.0034	0.152	± 0.0029
Sr	186	± 0.93	200	± 4.0	250	± 3.4	98.9	± 1.2
Th	0.884	± 0.0048	1.35	± 0.012	1.49	± 0.015	0.603	± 0.0088
Ti	260	± 0.57	332	± 6.1	382	± 13	197	± 7.2
Tl	0.0822	± 0.0017	0.117	± 0.0017	0.146	± 0.0022	0.0606	± 0.00040
U	0.281	± 0.0027	0.360	± 0.0049	0.443	± 0.0062	0.175	± 0.0030
V	12.2	± 0.049	14.9	± 0.25	17.8	± 0.24	7.81	± 0.10
Zn	186	± 0.46	254	± 4.9	323	± 2.8	143	± 2.4

Table 4. 49: Heavy metals for grocery waste

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000271	±	0.00016	0.00920	±	0.00025
Al	1.18	±	0.17	21.5	±	0.25
As	0.0159	±	0.0013	0.0262	±	0.0032
B	18.0	±	0.20	22.1	±	0.41
Ba	1.26	±	0.014	4.34	±	0.045
Be	BD	±		BD	±	
Bi	0.000183	±		0.00302	±	0.00015
Ca	726	±	2.3	1650	±	13
Cd	0.0225	±	0.00081	0.0631	±	0.0010
Cl	1750	±	19	8120	±	320
Co	0.0347	±	0.0015	0.0787	±	0.0022
Cr	0.220	±	0.0064	2.64	±	0.037
Cu	2.54	±	0.020	6.86	±	0.044
Fe	1.08	±	0.60	45.0	±	0.26
Hg	BD	±		BD	±	
Li	0.167	±	0.011	0.313	±	0.011
Mg	516	±	5.8	818	±	4.2
Mn	5.78	±	0.012	11.4	±	0.090
Mo	0.138	±	0.0027	0.294	±	0.0037
Na	460	±	13	660	±	3.5
Ni	0.872	±	0.0076	1.80	±	0.034
Pb	0.00324	±	0.00041	0.0365	±	0.00048
Sb	0.000953	±	0.00028	0.00359	±	0.00043
Se	0.0650	±	0.0031	BD	±	
Sn	BD	±		BD	±	
Sr	4.38	±	0.012	12.1	±	0.065
Th	0.000189	±		0.00483	±	0.00010
Ti	4.82	±	0.16	7.08	±	0.12
Tl	0.00405	±	0.000042	0.00641	±	0.00041
U	0.000273	±	0.000025	0.00653	±	0.00012
V	0.0180	±	0.00073	0.0787	±	0.0057
Zn	7.27	±	0.16	10.3	±	0.025

Table 4. 50: Heavy metals for grocery waste culture
 Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0901	± 0.0047	0.100	± 0.0039	0.154	± 0.0043	0.0823	± 0.0014
Al	6530	± 26	6400	± 37	11200	± 79	4780	± 54
As	1.69	± 0.044	1.98	± 0.026	2.67	± 0.021	1.02	± 0.013
B	24.6	± 0.96	28.8	± 0.52	34.5	± 0.70	16.5	± 0.40
Ba	97.8	± 1.5	92.8	± 0.88	133	± 1.9	61.1	± 0.64
Be	0.126	± 0.0037	0.136	± 0.0067	0.168	± 0.0066	0.0856	± 0.0044
Bi	0.0607	± 0.0035	0.0404	± 0.00054	0.0508	± 0.00073	0.0243	± 0.00046
Ca	25300	± 261	30100	± 320	35300	± 360	17400	± 220
Cd	0.164	± 0.0027	0.168	± 0.0032	0.216	± 0.0010	0.0929	± 0.0014
Cl	7860	± 350	39100	± 950	24500	± 470	12500	± 310
Co	3.88	± 0.065	4.30	± 0.029	5.38	± 0.044	2.56	± 0.019
Cr	11.4	± 0.12	13.5	± 0.18	19.9	± 0.13	7.28	± 0.099
Cu	43.6	± 0.58	46.6	± 0.35	57.9	± 0.54	28.1	± 0.41
Fe	4400	± 50	4920	± 47	6850	± 71	3720	± 32
Hg	0.0175	± 0.0010	0.0228	± 0.00074	0.0281	± 0.0017	0.0142	± 0.0011
Li	4.90	± 0.13	5.37	± 0.035	6.53	± 0.14	2.68	± 0.065
Mg	6930	± 89	8640	± 54	9870	± 88	7300	± 85
Mn	235	± 1.9	277	± 2.1	371	± 2.8	172	± 0.95
Mo	2.80	± 0.033	2.88	± 0.038	4.00	± 0.029	1.72	± 0.017
Na	8600	± 98	11100	± 99	15000	± 210	2350	± 35
Ni	11.7	± 0.16	13.3	± 0.071	17.6	± 0.19	6.30	± 0.11
Pb	2.30	± 0.014	2.75	± 0.020	3.53	± 0.018	2.07	± 0.011
Sb	0.0355	± 0.0011	0.0331	± 0.00080	0.0396	± 0.0013	0.0258	± 0.00042
Se	0.944	± 0.065	0.976	± 0.083	1.60	± 0.061	0.740	± 0.043
Sn	0.219	± 0.0025	0.110	± 0.0024	0.192	± 0.0029	0.0721	± 0.0018
Sr	165	± 2.1	191	± 2.0	253	± 3.9	116	± 2.0
Th	1.02	± 0.014	0.778	± 0.0069	1.22	± 0.016	0.505	± 0.0040
Ti	260	± 5.4	247	± 2.3	359	± 4.1	210	± 4.0
Tl	0.0964	± 0.00086	0.0834	± 0.0016	0.118	± 0.0018	0.0624	± 0.00071
U	0.298	± 0.0040	0.346	± 0.0036	0.445	± 0.0069	0.191	± 0.0022
V	11.2	± 0.14	12.2	± 0.12	17.4	± 0.20	8.63	± 0.13
Zn	217	± 2.0	269	± 4.0	371	± 3.0	188	± 2.6

Table 4. 51: Heavy metals for thermally hydrolyzed biowaste (#1)

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	±	Std dev	Mean	± Std dev
Ag	0.000212	±	0.000037	0.0111	± 0.00040
Al	0.117	±	0.0034	251	± 79
As	0.262	±	0.0063	0.555	± 0.0061
B	2.34	±	0.012	3.75	± 0.13
Ba	0.351	±	0.012	57.2	± 0.24
Be	BD	±		0.00454	± 0.0013
Bi	0.000200	±	0.000033	0.0561	± 0.0010
Ca	219	±	0.30	52300	± 220
Cd	0.000573	±	0.00013	0.0772	± 0.0016
Cl	3650	±	51	13700	± 250
Co	0.0768	±	0.0018	0.353	± 0.0057
Cr	0.319	±	0.012	4.48	± 0.015
Cu	0.160	±	0.0084	24.5	± 0.19
Fe	210	±	6.1	5720	± 22
Hg	0.000671	±	0.00011	0.0027	± 0.00075
Li	0.156	±	0.0057	0.644	± 0.011
Mg	548	±	5.1	1280	± 2.5
Mn	0.483	±	0.0014	75.4	± 0.15
Mo	0.188	±	0.011	1.03	± 0.0065
Na	4130	±	41	4740	± 13
Ni	0.470	±	0.0090	2.39	± 0.034
Pb	0.00108	±	0.00012	2.23	± 0.0076
Sb	0.00667	±	0.00021	0.288	± 0.019
Se	0.255	±	0.0062	0.265	± 0.030
Sn	0.00507	±	0.00066	0.207	± 0.0074
Sr	0.199	±	0.00059	49.5	± 0.27
Th	0.000121	±	0.000090	0.0282	± 0.00024
Ti	1.83	±	0.036	101	± 0.99
Tl	0.000963	±	0.000026	0.00420	± 0.00035
U	0.000196	±	0.000015	0.0385	± 0.00025
V	0.0604	±	0.0029	0.576	± 0.0048
Zn	1.29	±	0.030	60.8	± 0.27

Table 4. 52: Heavy metals for thermally hydrolyzed biowaste (#1) culture
 Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0994	± 0.0053	0.0908	± 0.0041	0.180	± 0.0033	0.0575	± 0.0024
Al	6720	± 47	5740	± 31	11000	± 110	3300	± 28
As	1.68	± 0.023	1.90	± 0.026	2.56	± 0.026	0.955	± 0.0081
B	23.5	± 0.43	24.7	± 0.64	29.2	± 0.78	13.9	± 0.32
Ba	99.2	± 2.2	93.6	± 0.77	129	± 2.2	60.6	± 0.72
Be	0.137	± 0.016	0.123	± 0.013	0.152	± 0.0049	0.0625	± 0.0044
Bi	0.0473	± 0.0012	0.0432	± 0.00067	0.0588	± 0.0013	0.0263	± 0.00066
Ca	30600	± 330	33400	± 430	37700	± 430	21500	± 160
Cd	0.153	± 0.0017	0.152	± 0.0048	0.205	± 0.0014	0.0807	± 0.0020
Cl	13000	± 180	39300	± 770	22800	± 180	16600	± 210
Co	3.72	± 0.079	4.19	± 0.056	5.38	± 0.056	2.36	± 0.022
Cr	11.3	± 0.098	12.2	± 0.076	17.0	± 0.19	5.89	± 0.070
Cu	43.9	± 0.97	46.7	± 0.49	58.2	± 0.62	27.6	± 0.23
Fe	5230	± 76	5020	± 36	7190	± 66	3680	± 18
Hg	0.0184	± 0.00095	0.0212	± 0.0014	0.0260	± 0.0010	0.0125	± 0.00082
Li	5.03	± 0.085	5.19	± 0.11	6.02	± 0.22	2.13	± 0.039
Mg	7290	± 65	8270	± 70	9550	± 110	5270	± 30
Mn	249	± 3.1	266	± 1.2	366	± 1.3	161	± 0.75
Mo	2.77	± 0.067	2.88	± 0.045	3.97	± 0.040	1.66	± 0.015
Na	9350	± 83	11300	± 140	15100	± 270	2400	± 25
Ni	11.4	± 0.27	12.4	± 0.15	16.0	± 0.24	5.85	± 0.073
Pb	2.54	± 0.018	2.73	± 0.022	3.52	± 0.018	1.72	± 0.012
Sb	0.0423	± 0.00047	0.0404	± 0.0044	0.0552	± 0.0013	0.0253	± 0.00049
Se	1.01	± 0.028	1.03	± 0.035	1.51	± 0.053	0.654	± 0.057
Sn	0.258	± 0.0069	0.108	± 0.0040	0.294	± 0.0028	0.0179	± 0.00035
Sr	168	± 3.2	181	± 2.5	242	± 4.0	116	± 1.5
Th	1.01	± 0.022	0.755	± 0.0097	1.35	± 0.0083	0.313	± 0.0037
Ti	295	± 34	238	± 3.5	364	± 7.3	152	± 2.8
Tl	0.0855	± 0.0020	0.0757	± 0.00069	0.108	± 0.0022	0.0504	± 0.0010
U	0.281	± 0.0078	0.321	± 0.0036	0.415	± 0.0055	0.170	± 0.0021
V	11.0	± 0.082	11.3	± 0.15	16.7	± 0.20	6.52	± 0.043
Zn	238	± 3.4	262	± 2.8	383	± 7.0	165	± 2.0

Table 4. 53: Heavy metals for thermally hydrolyzed biowaste (#2)

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000137	±	0.000047	0.00923	±	0.00080
Al	0.379	±	0.032	146	±	0.64
As	0.457	±	0.0052	0.677	±	0.011
B	2.35	±	0.017	3.27	±	0.18
Ba	0.311	±	0.036	29.4	±	0.20
Be	0.000172	±		0.00566	±	0.0040
Bi	0.000895	±	0.00025	0.834	±	0.0065
Ca	565	±	75	57900	±	210
Cd	0.00105	±	0.000043	0.0292	±	0.0014
Cl	4640	±	52	23300	±	520
Co	0.170	±	0.00071	0.520	±	0.0046
Cr	1.84	±	0.012	6.01	±	0.0032
Cu	0.0368	±	0.0014	14.3	±	0.097
Fe	929	±	42	7750	±	39
Hg	0.00116	±	0.00021	0.00989	±	0.0011
Li	0.0319	±	0.0024	0.270	±	0.0088
Mg	431	±	3.4	1470	±	4.7
Mn	1.82	±	0.13	82.1	±	0.30
Mo	0.489	±	0.0090	1.11	±	0.0061
Na	6200	±	190	6750	±	24
Ni	1.15	±	0.0039	2.94	±	0.044
Pb	0.00305	±	0.000085	2.83	±	0.0096
Sb	0.147	±	0.0011	0.725	±	0.0042
Se	0.287	±	0.0049	0.180	±	0.049
Sn	0.00400	±	0.000039	0.272	±	0.0027
Sr	0.535	±	0.067	44.9	±	0.27
Th	0.000998	±	0.00016	0.0296	±	0.00057
Ti	1.52	±	0.17	105	±	1.3
Tl	0.0000710	±	0.000034	0.00359	±	0.00022
U	0.000741	±	0.000095	0.0409	±	0.00043
V	0.229	±	0.0033	0.625	±	0.0041
Zn	9.60	±	0.20	86.0	±	0.48

Table 4. 54: Heavy metals for thermally hydrolyzed biowaste (#2) culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.111	± 0.0029	0.125	± 0.0049	0.136	± 0.0041	0.0539	± 0.0013
Al	6830	± 76	7170	± 43	10100	± 86	2860	± 12
As	1.72	± 0.024	2.02	± 0.023	2.56	± 0.019	1.05	± 0.0094
B	23.8	± 0.75	25.6	± 0.39	30.4	± 0.84	15.0	± 0.29
Ba	96.9	± 2.5	97.2	± 0.85	125	± 1.7	51.1	± 0.65
Be	0.125	± 0.0099	0.118	± 0.0062	0.165	± 0.0048	0.0707	± 0.0068
Bi	0.123	± 0.0035	0.125	± 0.0011	0.160	± 0.0022	0.0602	± 0.0011
Ca	29500	± 550	35400	± 440	40600	± 470	18100	± 230
Cd	0.144	± 0.0034	0.155	± 0.0022	0.189	± 0.0047	0.0857	± 0.0022
Cl	6380	± 200	17900	± 180	33100	± 660	19200	± 200
Co	4.02	± 0.094	4.22	± 0.032	5.31	± 0.027	2.54	± 0.025
Cr	13.0	± 0.18	14.6	± 0.057	23.9	± 0.29	5.49	± 0.055
Cu	42.9	± 1.0	46.1	± 0.45	55.8	± 0.60	27.0	± 0.35
Fe	5290	± 62	5770	± 34	7580	± 91	3340	± 43
Hg	0.0159	± 0.0014	0.0223	± 0.0016	0.0271	± 0.0011	0.0138	± 0.0010
Li	5.31	± 0.18	5.33	± 0.056	6.31	± 0.17	2.37	± 0.061
Mg	6900	± 84	8720	± 61	9890	± 110	5270	± 49
Mn	250	± 2.8	275	± 1.6	373	± 1.2	156	± 1.2
Mo	2.86	± 0.065	2.99	± 0.026	3.64	± 0.040	1.69	± 0.018
Na	9660	± 130	11100	± 95	14800	± 180	2800	± 31
Ni	12.6	± 0.33	13.0	± 0.11	18.4	± 0.13	6.30	± 0.072
Pb	4.38	± 0.031	2.93	± 0.022	3.69	± 0.035	1.71	± 0.012
Sb	0.0531	± 0.00051	0.0605	± 0.0013	0.0581	± 0.00076	0.0292	± 0.00056
Se	0.932	± 0.56	1.19	± 0.026	1.35	± 0.085	0.757	± 0.037
Sn	0.265	± 0.0044	0.218	± 0.0038	0.124	± 0.0016	0.0110	± 0.0012
Sr	173	± 3.1	191	± 1.6	248	± 3.1	106	± 1.5
Th	1.02	± 0.030	1.04	± 0.0083	1.01	± 0.014	0.300	± 0.0027
Ti	277	± 3.3	312	± 2.9	327	± 5.5	128	± 2.9
Tl	0.0896	± 0.0023	0.0880	± 0.0010	0.108	± 0.0026	0.0479	± 0.0013
U	0.284	± 0.0077	0.343	± 0.0030	0.402	± 0.0070	0.167	± 0.0021
V	11.1	± 0.17	13.4	± 0.13	15.6	± 0.17	6.18	± 0.080
Zn	234	± 3.4	271	± 3.6	371	± 4.5	170	± 2.6

Table 4. 55: Heavy metals for thin stillage

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000233	±	0.000021	0.00546	±	0.00019
Al	0.821	±	0.024	35.6	±	0.41
As	0.138	±	0.0027	0.133	±	0.0088
B	7.35	±	0.053	8.58	±	0.17
Ba	4.30	±	0.023	5.08	±	0.025
Be	BD	±		0.00204	±	0.0014
Bi	0.0000729	±		0.000810	±	0.00010
Ca	1300	±	6.0	2460	±	14
Cd	0.0353	±	0.00070	0.0480	±	0.00076
Cl	2190	±	12	12900	±	320
Co	0.0230	±	0.00053	0.0438	±	0.0021
Cr	0.167	±	0.0044	1.20	±	0.029
Cu	3.06	±	0.035	10.1	±	0.097
Fe	18.5	±	0.10	210	±	0.82
Hg	0.00141	±	0.0012	0.00372	±	0.0015
Li	1.08	±	0.036	1.14	±	0.014
Mg	4880	±	12	4480	±	30
Mn	96.5	±	0.64	101	±	0.32
Mo	1.04	±	0.0040	1.95	±	0.022
Na	3280	±	20	3240	±	28
Ni	0.263	±	0.0050	0.668	±	0.010
Pb	0.00455	±	0.00043	0.0274	±	0.00028
Sb	0.00111	±	0.00017	0.00178	±	0.00023
Se	0.141	±	0.010	0.298	±	0.027
Sn	BD	±		0.00500	±	0.00045
Sr	13.2	±	0.072	15.4	±	0.089
Th	0.00102	±	0.00042	0.00780	±	0.00019
Ti	25.8	±	0.50	29.6	±	0.25
Tl	0.000457	±	0.00010	0.00151	±	0.00010
U	0.000275	±	0.000054	0.0120	±	0.00010
V	0.0203	±	0.00081	0.130	±	0.0065
Zn	78.5	±	0.31	74.6	±	0.37

Table 4. 56: Heavy metals for thin stillage culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy Metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0830	± 0.0037	0.0567	± 0.00044	0.0694	± 0.00033	0.0350	± 0.00064
Al	5590	± 36	6410	± 200	7070	± 230	2910	± 43
As	1.85	± 0.016	1.80	± 0.015	2.20	± 0.055	1.08	± 0.015
B	23.3	± 0.41	23.1	± 0.58	26.0	± 0.84	17.0	± 0.71
Ba	87.2	± 1.2	84.3	± 1.3	101	± 1.5	53.6	± 0.63
Be	0.116	± 0.012	0.114	± 0.0059	0.138	± 0.0069	0.0768	± 0.0040
Bi	0.0484	± 0.0016	0.0340	± 0.00074	0.0414	± 0.00077	0.0226	± 0.00055
Ca	25900	± 210	25600	± 450	30700	± 710	17500	± 290
Cd	0.146	± 0.0022	0.145	± 0.0033	0.177	± 0.0020	0.0861	± 0.0023
Cl	26300	± 840	19600	± 360	37100	± 820	10100	± 450
Co	4.08	± 0.034	3.74	± 0.034	4.27	± 0.045	2.51	± 0.018
Cr	12.2	± 0.11	12.1	± 0.088	14.8	± 0.26	7.71	± 0.087
Cu	42.6	± 0.39	40.8	± 0.68	48.4	± 1.3	27.4	± 0.41
Fe	4480	± 61	4460	± 150	4930	± 220	2660	± 76
Hg	0.0293	± 0.0024	0.0168	± 0.00060	0.0235	± 0.0019	0.0142	± 0.00095
Li	5.14	± 0.16	5.25	± 0.081	6.43	± 0.13	2.78	± 0.082
Mg	7860	± 65	7210	± 89	8430	± 150	6300	± 73
Mn	267	± 1.4	298	± 8.3	354	± 14	164	± 2.9
Mo	2.73	± 0.015	2.54	± 0.020	2.93	± 0.067	1.72	± 0.023
Na	9550	± 110	9310	± 380	12000	± 400	2300	± 51
Ni	12.0	± 0.11	11.3	± 0.19	13.7	± 0.16	5.80	± 0.083
Pb	2.49	± 0.014	2.50	± 0.016	3.05	± 0.045	1.62	± 0.0078
Sb	0.0261	± 0.00044	0.0242	± 0.00054	0.0235	± 0.00084	0.0272	± 0.0010
Se	1.23	± 0.049	1.25	± 0.040	1.34	± 0.052	0.959	± 0.035
Sn	0.0287	± 0.0012	0.0599	± 0.00081	0.0713	± 0.0017	0.0469	± 0.00053
Sr	179	± 3.8	179	± 2.5	216	± 4.8	118	± 2.0
Th	0.488	± 0.0029	0.700	± 0.0065	0.814	± 0.014	0.337	± 0.0029
Ti	192	± 3.3	204	± 3.1	201	± 5.3	149	± 12
Tl	0.0752	± 0.0010	0.0729	± 0.0020	0.0872	± 0.0018	0.0488	± 0.0014
U	0.287	± 0.0022	0.303	± 0.0034	0.365	± 0.0061	0.188	± 0.0022
V	10.9	± 0.10	10.6	± 0.18	11.8	± 0.27	6.43	± 0.075
Zn	267	± 3.4	277	± 12	309	± 11	149	± 1.9

Table 4. 57: Heavy metals for glycerine with methanol

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	0.000420	±	0.00010	0.00517	± 0.00067
Al	0.265	±	0.052	2.86	± 0.052
As	0.00487	±		0.0118	± 0.0024
B	BD	±		0.559	± 0.083
Ba	0.138	±	0.0022	0.446	± 0.017
Be	BD	±		BD	±
Bi	0.00288	±	0.0018	0.00464	± 0.00041
Ca	39.2	±	6.2	189	± 3.6
Cd	0.00251	±	0.00015	0.00540	± 0.0012
Cl	BD	±		2430	± 280
Co	0.00127	±	0.00024	0.00304	± 0.00077
Cr	0.458	±	0.023	0.371	± 0.013
Cu	0.0612	±	0.012	0.172	± 0.0031
Fe	6.52	±	0.39	11.8	± 0.55
Hg	BD	±		BD	±
Li	BD	±		0.0138	± 0.0031
Mg	126	±	1.3	138	± 1.5
Mn	1.65	±	0.032	2.20	± 0.023
Mo	0.00595	±	0.0017	0.0187	± 0.00039
Na	213	±	2.0	213	± 3.1
Ni	0.0454	±	0.0015	0.125	± 0.0056
Pb	0.0192	±	0.0023	0.0449	± 0.0011
Sb	0.00119	±	0.00014	0.00352	± 0.00037
Se	0.0130	±	0.0031	0.0406	± 0.030
Sn	0.00110	±	0.00012	0.0118	± 0.00023
Sr	0.268	±	0.0052	0.438	± 0.0097
Th	0.104	±	0.042	0.00686	± 0.00048
Ti	1.72	±	0.011	1.40	± 0.032
Tl	0.00447	±	0.00078	0.00451	± 0.00058
U	0.00250	±	0.00036	0.00438	± 0.00047
V	0.0660	±	0.00074	0.0780	± 0.0031
Zn	2.78	±	0.48	3.63	± 0.041

Table 4. 58: Heavy metals for glycerine with methanol culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0436	± 0.0011	0.0871	± 0.00064	0.0953	± 0.0038	0.0505	± 0.0013
Al	6450	± 120	8030	± 31	8790	± 16	4620	± 49
As	1.67	± 0.027	1.99	± 0.038	2.44	± 0.026	1.07	± 0.0093
B	24.0	± 0.73	26.8	± 0.68	30.8	± 0.20	14.8	± 0.27
Ba	88.1	± 0.87	101	± 0.88	121	± 0.25	56.5	± 0.17
Be	0.125	± 0.011	0.135	± 0.011	0.137	± 0.0065	0.0852	± 0.0055
Bi	0.0372	± 0.00052	0.0375	± 0.00076	0.0439	± 0.00094	0.0222	± 0.00056
Ca	21900	± 380	28300	± 220	33800	± 85	16700	± 64
Cd	0.129	± 0.0025	0.158	± 0.0033	0.179	± 0.0031	0.0898	± 0.0026
Cl	16400	± 240	13700	± 120	29600	± 730	8260	± 220
Co	3.86	± 0.038	4.32	± 0.036	5.09	± 0.041	2.60	± 0.0070
Cr	14.1	± 0.16	14.0	± 0.17	14.1	± 0.059	5.96	± 0.061
Cu	43.6	± 0.39	47.5	± 0.36	51.9	± 0.15	26.4	± 0.075
Fe	4660	± 55	5370	± 35	5810	± 40	3390	± 67
Hg	0.0255	± 0.0011	0.0219	± 0.0011	0.0280	± 0.0013	0.0126	± 0.00022
Li	5.72	± 0.21	5.86	± 0.097	7.55	± 0.050	2.85	± 0.038
Mg	8020	± 140	9770	± 70	11800	± 46	6220	± 16
Mn	244	± 2.7	330	± 3.0	395	± 2.8	155	± 0.61
Mo	2.57	± 0.019	2.93	± 0.041	3.44	± 0.021	1.67	± 0.015
Na	10500	± 210	9480	± 80	13400	± 50	2700	± 3.0
Ni	12.5	± 0.15	12.9	± 0.094	15.1	± 0.091	6.30	± 0.059
Pb	2.73	± 0.036	2.95	± 0.021	3.30	± 0.014	1.65	± 0.0016
Sb	0.0305	± 0.0013	0.0455	± 0.0015	0.0431	± 0.0021	0.0305	± 0.0015
Se	1.02	± 0.060	1.23	± 0.065	1.31	± 0.064	0.818	± 0.053
Sn	0.101	± 0.0022	0.141	± 0.0057	0.114	± 0.0022	0.0931	± 0.0024
Sr	172	± 1.1	225	± 1.7	274	± 0.73	128	± 0.51
Th	0.872	± 0.011	0.926	± 0.013	0.889	± 0.0058	0.507	± 0.0027
Ti	234	± 6.2	305	± 2.6	340	± 1.5	206	± 0.89
Tl	0.0697	± 0.00062	0.0903	± 0.0015	0.105	± 0.0010	0.0565	± 0.0013
U	0.290	± 0.0037	0.324	± 0.0035	0.391	± 0.0011	0.185	± 0.0014
V	10.8	± 0.13	13.6	± 0.12	15.6	± 0.045	7.95	± 0.031
Zn	238	± 3.5	231	± 2.6	264	± 0.92	147	± 0.55

Table 4. 59: Heavy metals for hog slaughterhouse sludge (DAF)

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000275	±	0.00018	0.0278	±	0.0013
Al	0.810	±	0.038	308	±	1.6
As	0.0262	±	0.0014	0.237	±	0.010
B	0.530	±	0.36	1.80	±	0.079
Ba	0.285	±	0.0011	14.7	±	0.12
Be	BD	±		0.0121	±	0.0032
Bi	0.000559	±	0.00013	0.0223	±	0.00038
Ca	1990	±	15	5470	±	78
Cd	0.000208	±	0.000056	0.0666	±	0.0018
Cl	1570	±	370	7470	±	660
Co	0.0485	±	0.0015	0.433	±	0.0055
Cr	0.103	±	0.021	25.8	±	0.18
Cu	0.124	±	0.00045	78.1	±	0.90
Fe	7.35	±	0.19	1570	±	9.8
Hg	BD	±		0.0201	±	0.0011
Li	0.0211	±		0.400	±	0.0073
Mg	165	±	1.3	443	±	4.2
Mn	3.38	±	0.032	32.0	±	0.22
Mo	0.0118	±	0.00010	7.99	±	0.077
Na	627	±	19	938	±	8.1
Ni	0.311	±	0.0039	10.1	±	0.075
Pb	0.00393	±	0.00089	1.10	±	0.0094
Sb	0.00430	±	0.00046	0.286	±	0.0030
Se	0.0364	±	0.0096	1.38	±	0.024
Sn	0.000685	±	0.000010	1.34	±	0.0097
Sr	3.47	±	0.046	9.28	±	0.074
Th	0.000312	±	0.00011	0.0837	±	0.0015
Ti	3.58	±	0.060	20.7	±	0.33
Tl	0.000745	±	0.000081	0.0231	±	0.00050
U	0.00149	±	0.000018	0.851	±	0.012
V	0.0525	±	0.0045	0.656	±	0.078
Zn	9.11	±	0.11	177	±	2.3

Table 4. 60: Heavy metals for hog slaughterhouse peptone

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	BD	±		0.000626	± 0.00018
Al	3.73	±	0.33	362	± 1.6
As	0.0300	±	0.0029	0.00774	± 0.0036
B	0.173	±	0.072	0.569	± 0.060
Ba	0.159	±	0.0039	0.624	± 0.011
Be	0.000713	±	0.00011	0.0173	± 0.0017
Bi	0.000910	±	0.00014	0.0605	± 0.0011
Ca	20.2	±	0.18	865	± 16
Cd	0.0700	±	0.00058	0.0921	± 0.0030
Cl	5000	±	200	11100	± 140
Co	0.00860	±	0.00029	0.0180	± 0.00050
Cr	0.336	±	0.028	0.540	± 0.018
Cu	6.12	±	0.065	7.95	± 0.11
Fe	36.8	±	4.9	115	± 1.1
Hg	0.00881	±	0.0033	BD	±
Li	0.241	±	0.041	1.04	± 0.020
Mg	154	±	6.7	982	± 9.8
Mn	1.17	±	0.0012	3.45	± 0.0016
Mo	0.164	±	0.015	0.213	± 0.0040
Na	38700	±	480	60500	± 1000
Ni	0.0162	±	0.00046	0.185	± 0.0056
Pb	0.00786	±	0.00032	0.0291	± 0.00028
Sb	0.00134	±	0.00013	0.00303	± 0.00041
Se	1.29	±	0.036	1.55	± 0.060
Sn	0.00121	±	0.00021	BD	±
Sr	0.472	±	0.0042	1.66	± 0.023
Th	0.000830	±	0.00022	0.0285	± 0.00062
Ti	17.4	±	0.59	34.5	± 0.73
Tl	0.00791	±	0.00019	0.0181	± 0.0011
U	0.00171	±	0.00052	0.0318	± 0.0011
V	0.0558	±	0.0066	0.00892	± 0.0036
Zn	51.2	±	0.59	73.3	± 0.98

Table 4. 61: Heavy metals for wet distillers grains

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000581	±	0.00011	0.00564	±	0.00044
Al	0.635	±	0.061	13.7	±	0.10
As	0.0445	±	0.0023	0.0584	±	0.0041
B	2.69	±	0.090	3.96	±	0.075
Ba	0.910	±	0.024	3.42	±	0.025
Be	BD	±		0.00262	±	0.0016
Bi	0.000375	±	0.000076	0.000422	±	0.00010
Ca	589	±	13	1300	±	12
Cd	0.0168	±	0.00033	0.0410	±	0.0014
Cl	945	±	27	4880	±	200
Co	0.00959	±	0.00050	0.0159	±	0.00098
Cr	0.0758	±	0.0039	0.162	±	0.023
Cu	1.52	±	0.041	10.5	±	0.12
Fe	5.34	±	0.18	107	±	0.79
Hg	0.00204	±	0.00016	0.00177	±	0.00050
Li	0.387	±	0.027	0.494	±	0.011
Mg	1880	±	51	2060	±	14
Mn	37.1	±	0.76	53.0	±	0.37
Mo	0.477	±	0.015	1.41	±	0.013
Na	1320	±	29	1380	±	12
Ni	0.114	±	0.0032	0.194	±	0.0076
Pb	0.00222	±	0.00063	0.0186	±	0.00038
Sb	0.000366	±	0.00011	0.00168	±	0.00029
Se	0.0472	±	0.0092	0.0918	±	0.032
Sn	BD	±		0.00314	±	0.00040
Sr	4.65	±	0.11	8.88	±	0.075
Th	0.00129	±	0.00033	0.00354	±	0.00016
Ti	10.0	±	0.27	14.4	±	0.22
Tl	0.000399	±	0.00012	0.000738	±	0.00013
U	0.0000331	±	0.000023	0.00784	±	0.00016
V	0.0141	±	0.0013	0.0492	±	0.0029
Zn	33.1	±	0.83	42.8	±	0.33

Table 4. 62: Heavy metals for wet distillers grains culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0795	± 0.0016	0.121	± 0.0045	0.125	± 0.0064	0.0748	± 0.0023
Al	3580	± 43	7110	± 66	7390	± 94	3570	± 29
As	1.39	± 0.026	1.67	± 0.017	2.02	± 0.022	0.720	± 0.016
B	18.9	± 0.56	31.4	± 1.2	35.6	± 1.1	14.8	± 0.56
Ba	63.8	± 0.81	91.1	± 1.5	108	± 1.9	44.2	± 0.74
Be	0.0813	± 0.0084	0.148	± 0.0072	0.171	± 0.012	0.0705	± 0.0035
Bi	0.0288	± 0.00067	0.0347	± 0.00049	0.446	± 0.00053	0.0184	± 0.00057
Ca	20200	± 440	24800	± 510	31400	± 650	13700	± 370
Cd	0.127	± 0.0028	0.152	± 0.0045	0.191	± 0.0052	0.0707	± 0.0037
Cl	23400	± 490	3810	± 110	40800	± 2000	5070	± 610
Co	2.91	± 0.020	3.99	± 0.043	4.67	± 0.060	1.91	± 0.0023
Cr	7.86	± 0.16	15.9	± 0.064	15.5	± 0.17	5.92	± 0.10
Cu	34.9	± 0.56	44.9	± 0.76	53.7	± 1.1	22.1	± 0.46
Fe	2820	± 91	4390	± 54	5590	± 39	2650	± 24
Hg	0.0131	± 0.0015	0.0204	± 0.0014	0.0248	± 0.0011	0.0104	± 0.0012
Li	3.99	± 0.065	6.29	± 0.15	7.59	± 0.15	2.61	± 0.069
Mg	6080	± 140	6910	± 49	8650	± 130	4400	± 83
Mn	217	± 4.6	272	± 3.1	346	± 1.9	136	± 1.3
Mo	2.30	± 0.037	3.35	± 0.035	3.88	± 0.073	1.74	± 0.043
Na	5720	± 110	9940	± 120	13900	± 270	2270	± 57
Ni	8.52	± 0.082	12.6	± 0.23	14.6	± 0.24	4.88	± 0.11
Pb	1.89	± 0.025	2.60	± 0.014	3.22	± 0.038	1.27	± 0.013
Sb	0.0187	± 0.00055	0.0449	± 0.00053	0.0352	± 0.0015	0.0261	± 0.00098
Se	0.969	± 0.034	1.12	± 0.045	1.08	± 0.059	0.620	± 0.028
Sn	0.0439	± 0.00099	0.345	± 0.0052	0.146	± 0.0033	0.174	± 0.00081
Sr	137	± 2.4	178	± 5.2	224	± 4.2	85.7	± 2.0
Th	0.415	± 0.0068	1.17	± 0.019	0.925	± 0.015	0.553	± 0.0072
Ti	125	± 3.6	308	± 11	292	± 6.9	177	± 7.2
Tl	0.0525	± 0.0014	0.0905	± 0.0014	0.0993	± 0.0020	0.0460	± 0.0011
U	0.224	± 0.0032	0.320	± 0.0091	0.392	± 0.0073	0.153	± 0.0025
V	6.96	± 0.17	12.7	± 0.13	13.6	± 0.32	6.47	± 0.17
Zn	181	± 3.7	264	± 8.4	322	± 7.3	124	± 3.2

Table 4. 63: Heavy metals for cheese whey

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	BD	±		0.000476	± 0.00033
Al	0.101	±	0.0059	2.80	± 0.21
As	0.0574	±	0.00077	0.00507	± 0.0045
B	3.68	±	0.067	4.48	± 0.073
Ba	0.101	±	0.00051	0.129	± 0.0056
Be	BD	±		0.00118	± 0.00079
Bi	0.000553	±	0.000070	0.00133	± 0.00017
Ca	5620	±	17	6300	± 57
Cd	0.00101	±	0.00016	0.000768	± 0.00029
Cl	14200	±	54	5920	± 58
Co	0.000868	±	0.00044	0.00119	± 0.00065
Cr	0.485	±	0.031	0.0362	± 0.014
Cu	0.0963	±	0.0032	0.237	± 0.0086
Fe	BD	±		0.742	± 0.43
Hg	0.177	±		0.00336	± 0.00061
Li	0.0617	±	0.018	0.279	± 0.011
Mg	1230	±	2.6	1100	± 4.7
Mn	0.0106	±	0.0016	0.0849	± 0.0062
Mo	0.388	±	0.0039	0.500	± 0.013
Na	6310	±	50	5090	± 50
Ni	BD	±		0.00395	± 0.0024
Pb	0.00232	±	0.00023	0.184	± 0.0015
Sb	0.000397	±	0.000016	0.000233	± 0.00010
Se	0.195	±	0.049	0.0380	± 0.017
Sn	BD	±		0.00364	± 0.00047
Sr	1.86	±	0.0097	2.09	± 0.033
Th	0.000639	±	0.00016	0.000753	± 0.00015
Ti	21.7	±	0.097	23.3	± 0.58
Tl	0.00172	±	0.000051	0.00309	± 0.00027
U	0.000128	±	0.000032	0.000450	± 0.00010
V	0.0899	±	0.0053	0.0287	± 0.0015
Zn	1.14	±	0.0026	1.79	± 0.017

Table 4. 64: Heavy metals for cheese whey culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0795	± 0.0016	0.130	± 0.0037	0.164	± 0.0056	0.0710	± 0.0019
Al	3580	± 43	7430	± 130	9040	± 260	4280	± 94
As	1.39	± 0.026	1.71	± 0.026	2.05	± 0.037	0.793	± 0.014
B	18.9	± 0.56	30.0	± 0.82	35.1	± 0.91	15.0	± 0.24
Ba	63.8	± 0.81	91.1	± 1.5	112	± 1.3	50.0	± 0.99
Be	0.0813	± 0.0084	0.144	± 0.0095	0.170	± 0.0082	0.0716	± 0.0049
Bi	0.0288	± 0.00067	0.0406	± 0.00067	0.0472	± 0.00048	0.0218	± 0.00078
Ca	20200	± 440	27400	± 580	32500	± 660	15800	± 430
Cd	0.127	± 0.0028	0.144	± 0.0011	0.163	± 0.0015	0.0682	± 0.0022
Cl	23400	± 490	8200	± 260	8550	± 180	1080	± 71
Co	2.91	± 0.020	4.02	± 0.076	4.50	± 0.063	2.08	± 0.037
Cr	7.86	± 0.16	12.4	± 0.16	14.0	± 0.14	7.47	± 0.70
Cu	34.9	± 0.56	41.4	± 0.73	48.4	± 0.96	22.0	± 0.60
Fe	2820	± 91	4320	± 63	5340	± 160	2890	± 30
Hg	0.0131	± 0.0015	0.0211	± 0.0010	0.0251	± 0.00093	0.0118	± 0.00082
Li	3.99	± 0.065	6.11	± 0.15	7.47	± 0.14	2.72	± 0.060
Mg	6080	± 140	6530	± 89	6350	± 68	6960	± 98
Mn	217	± 4.6	272	± 2.6	309	± 2.5	146	± 2.7
Mo	2.30	± 0.037	2.94	± 0.049	3.49	± 0.057	1.51	± 0.032
Na	5720	± 110	12100	± 240	17000	± 290	2560	± 44
Ni	8.52	± 0.082	11.7	± 0.21	13.3	± 0.21	5.08	± 0.10
Pb	1.89	± 0.025	2.70	± 0.015	3.23	± 0.020	1.44	± 0.010
Sb	0.0187	± 0.00055	0.0426	± 0.00090	0.0496	± 0.00074	0.0309	± 0.0010
Se	0.969	± 0.034	1.06	± 0.052	1.24	± 0.059	0.613	± 0.022
Sn	0.0439	± 0.00099	0.254	± 0.0018	0.412	± 0.0054	0.195	± 0.0038
Sr	137	± 2.4	191	± 4.6	217	± 4.1	97.7	± 1.9
Th	0.145	± 0.0068	1.10	± 0.015	1.47	± 0.021	0.630	± 0.0077
Ti	125	± 3.6	347	± 16	359	± 11	232	± 18
Tl	0.0525	± 0.0014	0.0911	± 0.0015	0.111	± 0.0019	0.0535	± 0.0013
U	0.224	± 0.0032	0.335	± 0.0027	0.400	± 0.0055	0.168	± 0.0033
V	6.96	± 0.17	12.6	± 0.25	15.4	± 0.26	7.24	± 0.13
Zn	181	± 3.7	264	± 6.8	301	± 5.7	143	± 2.3

Table 4. 65: Heavy metals for cattle slaughterhouse waste - paunch

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag		±		0.0528	± 0.0078
Al	1.61	±	0.89	628	± 3.1
As	0.0639	±	0.0020	0.143	± 0.0075
B	2.50	±	0.29	8.19	± 0.13
Ba		±		35.0	± 0.51
Be		±		0.0135	± 0.0022
Bi		±		0.0663	± 0.020
Ca	331	±	45	4410	± 69
Cd		±		0.0448	± 0.0017
Cl	5240	±	750	6910	± 280
Co	0.0183	±	0.0038	0.187	± 0.0018
Cr		±		1.92	± 0.025
Cu	0.420	±	0.085	5.37	± 0.082
Fe	9.97	±	3.6	434	± 7.2
Hg		±		0.00469	± 0.0010
Li	0.808	±	0.12	1.21	± 0.033
Mg	240	±	34	633	± 5.6
Mn	4.00	±	0.58	51.8	± 0.72
Mo	0.0689	±	0.060	0.824	± 0.013
Na	16900	±	360	19700	± 550
Ni	0.389	±	0.049	1.51	± 0.030
Pb	0.00931	±	0.00018	0.222	± 0.0018
Sb	0.000869	±	0.0011	0.00651	± 0.00043
Se	0.0104	±	0.0024	0.107	± 0.011
Sn		±		0.0211	± 0.0011
Sr	2.52	±	0.35	23.8	± 1.0
Th	0.00120	±	0.00063	0.0812	± 0.0012
Ti	11.1	±	1.6	30.3	± 0.92
Tl	0.00592	±	0.00063	0.0458	± 0.0010
U	0.000834	±	0.00078	0.0247	± 0.0011
V	0.0569	±	0.0061	1.18	± 0.012
Zn	1.84	±	0.19	30.6	± 0.49

Table 4. 66: Heavy metals for cattle slaughterhouse waste - paunch culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.904	± 0.0055	0.0591	± 0.0020	0.0627	± 0.0013	0.0332	± 0.0017
Al	6680	± 92	4380	± 64	5150	± 70	2830	± 54
As	1.45	± 0.033	1.69	± 0.023	2.29	± 0.027	0.854	± 0.015
B	23.0	± 0.97	24.4	± 0.64	27.8	± 0.93	15.0	± 0.41
Ba	91.8	± 1.7	77.2	± 1.1	109	± 2.6	46.5	± 0.96
Be	0.113	± 0.0035	0.107	± 0.0027	0.138	± 0.0083	0.0625	± 0.0025
Bi	0.0355	± 0.0012	0.0273	± 0.00059	0.0396	± 0.0012	0.0138	± 0.00079
Ca	27900	± 490	24600	± 660	31900	± 740	15000	± 460
Cd	0.135	± 0.0038	0.138	± 0.0034	0.200	± 0.0020	0.0724	± 0.0017
Cl	22400	± 760	39900	± 820	86400	± 4200	22700	± 450
Co	3.43	± 0.066	3.87	± 0.080	4.96	± 0.055	2.27	± 0.044
Cr	16.2	± 0.26	11.1	± 0.098	12.0	± 0.19	6.70	± 0.13
Cu	36.3	± 1.0	40.9	± 0.85	54.7	± 0.93	22.9	± 0.59
Fe	5390	± 120	3770	± 58	4640	± 79	2810	± 24
Hg	0.0250	± 0.00047	0.0297	± 0.0012	0.0308	± 0.00094	0.0161	± 0.0010
Li	5.31	± 0.12	4.76	± 0.15	6.37	± 0.14	2.39	± 0.067
Mg	6860	± 67	6500	± 120	8480	± 140	4530	± 72
Mn	264	± 1.7	220	± 3.8	319	± 2.5	152	± 0.93
Mo	2.38	± 0.065	2.52	± 0.041	3.31	± 0.058	1.49	± 0.042
Na	9060	± 620	9310	± 150	13500	± 370	2850	± 53
Ni	11.8	± 0.24	11.7	± 0.25	14.8	± 0.18	5.83	± 0.17
Pb	2.46	± 0.059	2.52	± 0.014	3.37	± 0.037	1.46	± 0.011
Sb	0.0369	± 0.0016	0.0173	± 0.00068	0.0184	± 0.00082	0.0154	± 0.0010
Se	0.842	± 0.064	0.965	± 0.022	1.02	± 0.048	0.589	± 0.036
Sn	0.194	± 0.0039	0.0544	± 0.0010	0.0690	± 0.0030	0.0338	± 0.0014
Sr	176	± 4.3	150	± 2.4	215	± 5.3	95.7	± 1.9
Th	0.955	± 0.020	0.766	± 0.0098	0.889	± 0.0097	0.405	± 0.0072
Ti	278	± 7.0	159	± 2.5	148	± 4.3	128	± 3.6
Tl	0.0864	± 0.0029	0.0741	± 0.00091	0.0915	± 0.0020	0.0461	± 0.0010
U	0.295	± 0.0056	0.289	± 0.0058	0.390	± 0.0069	0.156	± 0.0028
V	10.9	± 0.30	9.45	± 0.14	10.5	± 0.25	5.85	± 0.12
Zn	210	± 4.2	226	± 0.39	323	± 8.8	139	± 2.9

Table 4. 67: Heavy metals for solid hog manure

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	Std dev		Mean	Std dev
Ag	0.000614	±	0.000029	0.0300	± 0.0025
Al	2.21	±	0.057	256	± 4.9
As	0.0952	±	0.0057	0.163	± 0.0086
B	3.31	±	0.052	9.91	± 0.22
Ba	0.952	±	0.020	45.2	± 0.29
Be		±		0.00769	± 0.0016
Bi	0.000720	±	0.000040	0.0211	± 0.0036
Ca	255	±	1.3	18000	± 110
Cd	0.0212	±	0.00081	0.254	± 0.0027
Cl	13700	±	420	4710	± 32
Co	0.153	±	0.0035	0.902	± 0.0048
Cr	0.448	±	0.011	2.96	± 0.050
Cu	22.2	±	0.73	277	± 3.3
Fe	50.6	±	1.6	771	± 5.8
Hg		±		0.0200	± 0.0020
Li	0.343	±	0.026	0.638	± 0.013
Mg	130	±	4.0	7390	± 26
Mn	4.74	±	0.064	361	± 2.5
Mo	0.213	±	0.0086	1.82	± 0.014
Na	5300	±	190	6380	± 27
Ni	0.438	±	0.015	2.47	± 0.026
Pb	0.0185	±	0.00035	0.325	± 0.0021
Sb	0.00960	±	0.00065	0.0547	± 0.0012
Se	0.230	±	0.0089	2.22	± 0.066
Sn	0.0103	±	0.00041	0.168	± 0.0025
Sr	0.754	±	0.011	27.5	± 0.28
Th	0.000907	±	0.000010	0.0513	± 0.0015
Ti	4.67	±	0.014	46.7	± 0.29
Tl	0.00482	±	0.00010	0.0830	± 0.0013
U	0.0152	±	0.00034	0.0590	± 0.0011
V	0.0590	±	0.0072	0.972	± 0.13
Zn	36.4	±	0.70	566	± 6.6

Table 4. 68: Heavy metals for solid hog manure culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0721	± 0.0016	0.243	± 0.042	0.0796	± 0.0043	0.0896	± 0.0052
Al	4550	± 65	4470	± 75	6380	± 110	2220	± 27
As	1.14	± 0.015	1.42	± 0.042	1.97	± 0.032	0.657	± 0.012
B	20.6	± 0.70	26.3	± 1.2	30.3	± 0.73	14.7	± 0.34
Ba	78.8	± 1.7	78.0	± 1.0	114	± 2.2	38.4	± 0.63
Be	0.106	± 0.0052	0.0983	± 0.0060	0.140	± 0.0080	0.0504	± 0.0020
Bi	0.0287	± 0.00066	0.0479	± 0.0052	0.0336	± 0.00051	0.0148	± 0.0014
Ca	28300	± 670	27000	± 620	32500	± 560	15100	± 240
Cd	0.193	± 0.0026	0.220	± 0.0031	0.296	± 0.0037	0.123	± 0.0036
Cl	23200	± 1100	55800	± 5800	43900	± 1200	11800	± 1300
Co	2.87	± 0.070	3.43	± 0.036	4.56	± 0.072	2.04	± 0.049
Cr	8.99	± 0.14	10.9	± 0.13	13.9	± 0.13	5.95	± 0.037
Cu	151	± 3.3	154	± 3.6	171	± 2.2	134	± 2.1
Fe	3760	± 72	3740	± 22	4850	± 77	2100	± 33
Hg	0.0289	± 0.0026	0.0312	± 0.0023	0.0350	± 0.0012	0.0202	± 0.0011
Li	4.34	± 0.12	4.33	± 0.070	6.11	± 0.085	1.74	± 0.024
Mg	7720	± 98	7970	± 77	10000	± 98	4530	± 61
Mn	337	± 3.2	343	± 2.0	464	± 6.2	170	± 2.0
Mo	2.57	± 0.049	2.91	± 0.052	4.00	± 0.052	1.66	± 0.031
Na	31100	± 930	8740	± 100	12500	± 250	2350	± 28
Ni	8.40	± 0.25	10.1	± 0.16	14.1	± 0.24	4.27	± 0.091
Pb	1.78	± 0.025	2.09	± 0.0078	2.74	± 0.015	1.12	± 0.0085
Sb	0.0390	± 0.0013	0.0286	± 0.00073	0.0369	± 0.00079	0.0340	± 0.00031
Se	1.27	± 0.071	1.84	± 0.11	1.94	± 0.056	1.24	± 0.028
Sn	0.169	± 0.0046	0.102	± 0.0030	0.168	± 0.0045	0.190	± 0.0037
Sr	140	± 3.0	132	± 2.0	188	± 3.9	78.1	± 1.5
Th	0.669	± 0.018	0.677	± 0.0092	0.939	± 0.0082	0.352	± 0.0034
Ti	212	± 4.7	169	± 6.4	224	± 8.0	122	± 1.9
Tl	0.0873	± 0.0023	0.100	± 0.0018	0.111	± 0.0021	0.0811	± 0.0020
U	0.232	± 0.0058	0.268	± 0.0032	0.354	± 0.0065	0.134	± 0.0020
V	8.40	± 0.13	8.80	± 0.18	12.0	± 0.16	4.91	± 0.062
Zn	356	± 8.2	422	± 8.3	595	± 12	267	± 3.5

Table 4. 69: Heavy metals for sewage sludge

Heavy metal	Results given as µg/g dw				
	Aqueous substrate			Dried substrate	
	Mean	±	Std dev	Mean	± Std dev
Ag	0.00420	±	0.00016	5.72	± 0.075
Al	26.7	±	0.48	35000	± 270
As	0.127	±	0.0022	2.82	± 0.038
B	3.88	±	0.21	16.7	± 0.47
Ba	8.56	±	0.24	588	± 6.2
Be		±		0.296	± 0.010
Bi	0.00826	±	0.000099	20.9	± 0.19
Ca	4540	±	150	15400	± 180
Cd	0.000734	±	0.000042	0.977	± 0.0085
Cl	569	±	21	685	± 35
Co	0.153	±	0.0020	3.24	± 0.034
Cr	0.0342	±	0.0063	30.4	± 0.47
Cu	0.240	±	0.0084	326	± 7.5
Fe	293	±	5.0	7050	± 78
Hg		±		0.691	± 0.0080
Li	0.704	±	0.088	9.61	± 0.13
Mg	1110	±	17	2840	± 37
Mn	20.6	±	0.40	121	± 0.60
Mo	0.0471	±	0.00025	4.13	± 0.029
Na	965	±	22	8440	± 180
Ni	0.348	±	0.016	17.8	± 0.24
Pb	0.00956	±	0.00017	16.3	± 0.057
Sb	0.00494	±	0.00021	0.810	± 0.0063
Se	0.105	±	0.0062	2.60	± 0.068
Sn	0.00828	±	0.00013	6.24	± 0.046
Sr	36.5	±	0.70	198	± 5.1
Th	0.000303	±	0.000036	0.651	± 0.0072
Ti	0.306	±	0.012	200	± 9.0
Tl	0.0000930	±	0.000088	0.107	± 0.0017
U	0.00607	±	0.00023	2.30	± 0.024
V	0.00651	±	0.0014	14.6	± 0.21
Zn	0.615	±	0.022	725	± 13

Table 4. 70: Heavy metals for sewage sludge culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	1.64	± 0.095	1.93	± 0.051	2.53	± 0.036	1.43	± 0.017
Al	15800	± 300	16400	± 150	21000	± 380	10400	± 99
As	2.00	± 0.066	2.57	± 0.030	3.06	± 0.057	1.36	± 0.016
B	23.1	± 1.1	26.1	± 0.41	29.0	± 0.67	15.4	± 0.62
Ba	227	± 3.5	287	± 5.8	359	± 6.0	181	± 3.9
Be	0.193	± 0.014	0.187	± 0.0034	0.247	± 0.010	0.120	± 0.010
Bi	6.64	± 0.16	7.97	± 0.096	10.1	± 0.040	5.59	± 0.050
Ca	22700	± 250	25700	± 550	32100	± 670	14900	± 420
Cd	0.352	± 0.010	0.441	± 0.0051	0.555	± 0.0051	0.269	± 0.0036
Cl	6040	± 290	23000	± 1400	10100	± 190	7300	± 690
Co	3.69	± 0.12	4.47	± 0.074	5.06	± 0.081	2.79	± 0.049
Cr	17.4	± 0.55	19.8	± 0.23	22.9	± 0.23	10.4	± 0.12
Cu	83.2	± 1.8	100	± 2.0	128	± 2.8	69.0	± 1.5
Fe	6890	± 140	6270	± 39	7800	± 110	4050	± 76
Hg	0.490	± 0.018	0.208	± 0.0065	0.243	± 0.0034	0.145	± 0.0037
Li	6.74	± 0.24	7.28	± 0.11	9.37	± 0.10	3.71	± 0.086
Mg	5310	± 57	6590	± 74	8000	± 54	3910	± 76
Mn	199	± 1.3	239	± 1.4	302	± 2.0	143	± 0.53
Mo	3.06	± 0.11	3.78	± 0.059	4.78	± 0.067	2.27	± 0.057
Na	6020	± 340	7200	± 110	8000	± 97	1700	± 42
Ni	12.8	± 0.47	16.5	± 0.34	17.9	± 0.22	7.58	± 0.20
Pb	6.48	± 0.14	7.93	± 0.050	9.72	± 0.064	5.39	± 0.042
Sb	0.269	± 0.0083	0.165	± 0.0032	0.225	± 0.0035	0.181	± 0.0049
Se	1.39	± 0.040	1.92	± 0.070	2.27	± 0.015	1.10	± 0.063
Sn	3.02	± 0.077	1.31	± 0.0080	3.78	± 0.027	1.09	± 0.013
Sr	171	± 3.4	208	± 5.2	253	± 6.3	118	± 2.5
Th	1.22	± 0.032	0.952	± 0.019	1.43	± 0.0093	0.582	± 0.011
Ti	267	± 5.7	218	± 7.1	316	± 6.5	162	± 4.8
Tl	0.101	± 0.0028	0.101	± 0.0021	0.133	± 0.0027	0.0642	± 0.0020
U	1.02	± 0.035	1.18	± 0.021	1.47	± 0.016	0.690	± 0.013
V	13.5	± 0.54	14.5	± 0.26	19.3	± 0.39	9.22	± 0.19
Zn	257	± 5.6	386	± 8.6	468	± 11	237	± 6.5

Table 4. 71: Heavy metals for liquid hog manure

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.000564	±	0.00011	0.0223	±	0.00052
Al	2.72	±	0.057	477	±	9.0
As	0.464	±	0.0094	0.708	±	0.016
B	36.2	±	0.74	52.1	±	1.4
Ba	1.90	±	0.020	36.0	±	0.30
Be	0.0200	±		0.173	±	0.0060
Bi	0.00148	±	0.00013	0.0167	±	0.00031
Ca	5140	±	150	26500	±	490
Cd	0.00473	±	0.00011	0.523	±	0.0026
Cl	23900	±	430	73900	±	1700
Co	0.519	±	0.0098	2.34	±	0.0065
Cr	1.82	±	0.044	9.84	±	0.28
Cu	3.83	±	0.14	587	±	13
Fe	73.1	±	1.2	1590	±	63
Hg	0.00431	±	0.00030	0.0192	±	0.0013
Li	1.16	±	0.035	1.56	±	0.029
Mg	525	±	13	12900	±	220
Mn	9.57	±	0.23	594	±	14
Mo	0.815	±	0.016	9.11	±	0.11
Na	13100	±	430	13600	±	160
Ni	2.81	±	0.059	9.58	±	0.13
Pb	0.00688	±	0.00066	0.781	±	0.0050
Sb	0.0152	±	0.00036	0.0971	±	0.0019
Se	0.906	±	0.022	3.80	±	0.057
Sn	BD	±		0.0563	±	0.0020
Sr	20.9	±	0.55	68.6	±	1.1
Th	0.000582	±	0.000017	0.0455	±	0.00055
Ti	13.9	±	0.26	94.7	±	2.3
Tl	0.00113	±	0.00014	0.173	±	0.0023
U	0.532	±	0.0087	2.82	±	0.028
V	0.684	±	0.017	5.67	±	0.089
Zn	6.22	±	0.29	1770	±	33

Table 4. 72: Heavy metals for liquid hog culture

Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.0869	± 0.0021	0.134	± 0.0021	0.134	± 0.0026	0.0550	± 0.0016
Al	4520	± 54	6840	± 180	8320	± 120	3160	± 45
As	1.63	± 0.010	1.85	± 0.026	2.40	± 0.020	0.793	± 0.013
B	25.2	± 1.0	26.2	± 0.72	31.9	± 0.95	14.5	± 0.35
Ba	75.6	± 1.3	95.2	± 1.3	114	± 1.4	48.5	± 0.96
Be	0.111	± 0.0055	0.140	± 0.0064	0.162	± 0.0052	0.0772	± 0.0055
Bi	0.0315	± 0.00057	0.0362	± 0.0010	0.0443	± 0.00039	0.0187	± 0.00042
Ca	25000	± 630	28500	± 690	34700	± 510	16800	± 380
Cd	0.181	± 0.0020	0.210	± 0.0031	0.263	± 0.0034	0.0987	± 0.0026
Cl	50000	± 2000	11500	± 290	39300	± 1100	14000	± 480
Co	3.63	± 0.051	3.93	± 0.054	4.91	± 0.061	2.09	± 0.035
Cr	9.71	± 0.090	13.0	± 0.27	14.1	± 0.20	6.73	± 0.15
Cu	103	± 2.2	114	± 2.6	139	± 2.5	58.3	± 1.2
Fe	3860	± 63	4820	± 62	5730	± 110	2930	± 38
Hg	0.0153	± 0.0012	0.0212	± 0.0013	0.0232	± 0.00086	0.0108	± 0.0016
Li	4.56	± 0.10	5.15	± 0.10	6.53	± 0.15	2.32	± 0.055
Mg	6960	± 85	8060	± 69	9300	± 130	4850	± 79
Mn	279	± 1.6	324	± 2.0	403	± 4.3	182	± 1.2
Mo	3.48	± 0.058	4.05	± 0.064	4.99	± 0.074	1.90	± 0.039
Na	8920	± 210	9600	± 180	13700	± 190	1910	± 45
Ni	10.4	± 0.17	11.7	± 0.22	14.6	± 0.22	5.21	± 0.10
Pb	2.27	± 0.010	2.58	± 0.013	3.25	± 0.20	1.41	± 0.0060
Sb	0.0258	± 0.0010	0.0455	± 0.0013	0.0445	± 0.00033	0.0236	± 0.00031
Se	1.20	± 0.031	1.61	± 0.036	1.98	± 0.11	0.924	± 0.050
Sn	0.0392	± 0.00071	0.264	± 0.0029	0.134	± 0.0016	0.0372	± 0.0016
Sr	160	± 3.1	182	± 2.6	224	± 2.9	104	± 1.8
Th	0.481	± 0.0067	0.977	± 0.015	0.916	± 0.012	0.301	± 0.0061
Ti	167	± 4.1	295	± 7.6	305	± 6.0	156	± 0.83
Tl	0.0854	± 0.0025	0.108	± 0.0022	0.125	± 0.0031	0.0550	± 0.0014
U	0.619	± 0.011	0.682	± 0.010	0.888	± 0.017	0.341	± 0.0064
V	9.31	± 0.19	12.4	± 0.24	13.9	± 0.23	6.57	± 0.14
Zn	436	± 11	486	± 9.3	625	± 10	257	± 5.2

Table 4. 73: Heavy metals for liquid sewage

Heavy metal	Results given as µg/g dw					
	Aqueous substrate			Dried substrate		
	Mean	±	Std dev	Mean	±	Std dev
Ag	1.18	±	0.016	10.1	±	0.73
Al	41.3	±	0.62	3490	±	43
As	0.514	±	0.048	0.956	±	0.034
B	421	±	1.2	290	±	13
Ba	34.4	±	0.57	118	±	2.2
Be	BD	±		0.0313	±	0.0023
Bi	0.0760	±	0.00047	6.42	±	0.11
Ca	68900	±	210	63400	±	1700
Cd	0.0201	±	0.0060	0.223	±	0.0039
Cl	151000	±	600	212000	±	9300
Co	0.270	±	0.015	0.803	±	0.0087
Cr	1.16	±	0.047	10.4	±	0.22
Cu	9.12	±	0.28	46.0	±	0.97
Fe	80.5	±	6.7	1320	±	44
Hg	BD	±		0.127	±	0.0027
Li	10.3	±	0.76	8.11	±	0.15
Mg	19600	±	0	13800	±	230
Mn	69.3	±	0.59	103	±	1.5
Mo	1.14	±	0.012	1.62	±	0.018
Na	132000	±	4300	97500	±	1800
Ni	2.06	±	0.26	3.50	±	0.072
Pb	0.229	±	0.023	2.52	±	0.041
Sb	0.313	±	0.0082	0.419	±	0.0072
Se	0.880	±	0.067	1.23	±	0.16
Sn	0.311	±	0.0080	1.58	±	0.024
Sr	536	±	2.7	470	±	12
Th	BD	±		0.0491	±	0.0011
Ti	10.4	±	0.10	37.9	±	1.8
Tl	0.00978	±	0.0042	0.0153	±	0.00056
U	0.710	±	0.0021	0.814	±	0.019
V	0.125	±	0.043	1.93	±	0.055
Zn	10.9	±	0.060	114	±	2.1

Table 4. 74: Heavy metals for liquid sewage culture
Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.231	± 0.0069	0.230	± 0.0036	0.309	± 0.0054	0.166	± 0.0029
Al	7000	± 210	5900	± 66	9680	± 98	4360	± 50
As	1.91	± 0.014	1.95	± 0.011	2.61	± 0.029	0.876	± 0.019
B	25.6	± 0.71	23.9	± 0.54	31.5	± 0.68	12.8	± 0.44
Ba	92.7	± 2.1	93.0	± 1.7	122	± 1.5	57.9	± 0.79
Be	0.130	± 0.011	0.114	± 0.0045	0.161	± 0.0050	0.0786	± 0.0024
Bi	0.0844	± 0.00069	0.115	± 0.0010	0.113	± 0.0011	0.0472	± 0.00075
Ca	26000	± 670	26400	± 660	33400	± 610	15800	± 270
Cd	0.143	± 0.0029	0.153	± 0.0024	0.189	± 0.0031	0.0736	± 0.0029
Cl	20200	± 390	32200	± 1000	33000	± 480	5470	± 680
Co	4.00	± 0.049	4.15	± 0.056	5.16	± 0.040	2.06	± 0.032
Cr	11.0	± 0.099	10.2	± 0.14	14.0	± 0.13	7.13	± 0.098
Cu	42.9	± 0.48	45.5	± 0.60	54.7	± 0.53	23.1	± 0.36
Fe	4810	± 73	4530	± 44	6300	± 100	3310	± 33
Hg	0.0194	± 0.0011	0.0214	± 0.00056	0.0267	± 0.0014	0.0120	± 0.0010
Li	5.64	± 0.10	5.11	± 0.12	7.13	± 0.14	2.68	± 0.060
Mg	6610	± 71	6930	± 93	8620	± 130	4260	± 57
Mn	254	± 1.9	262	± 1.6	346	± 1.7	146	± 1.4
Mo	2.89	± 0.043	2.82	± 0.038	3.77	± 0.057	1.40	± 0.023
Na	9560	± 140	9420	± 220	13800	± 300	1870	± 55
Ni	11.2	± 0.16	11.4	± 0.22	14.8	± 0.17	5.14	± 0.086
Pb	2.70	± 0.018	2.72	± 0.025	3.42	± 0.028	1.50	± 0.0085
Sb	0.0405	± 0.0011	0.0272	± 0.00059	0.0390	± 0.0012	0.0264	± 0.00075
Se	1.14	± 0.055	1.15	± 0.056	1.60	± 0.063	0.637	± 0.024
Sn	0.153	± 0.0024	0.0635	± 0.0015	0.199	± 0.0026	0.212	± 0.0037
Sr	185	± 3.6	191	± 4.4	240	± 2.9	103	± 1.5
Th	0.867	± 0.0061	0.643	± 0.0071	1.14	± 0.0068	0.677	± 0.0070
Ti	279	± 5.9	216	± 4.9	342	± 5.3	189	± 2.6
Tl	0.0834	± 0.0014	0.0801	± 0.0012	0.106	± 0.0017	0.0547	± 0.00083
U	0.310	± 0.0055	0.318	± 0.0039	0.421	± 0.0049	0.181	± 0.0026
V	12.3	± 0.18	11.3	± 0.17	15.2	± 0.19	7.38	± 0.15
Zn	250	± 4.7	255	± 5.0	325	± 3.6	138	± 2.3

4.6 Heavy metals for substrates used in the ARC Pilot Plant, please refer to Section 4.2 for explanations

Table 4. 75: Heavy metals for ARC Pilot Plant (1) mixture
Dairy manure and glycerine

Heavy metal	Results given as $\mu\text{g/g dw}$		
	Aqueous mixture		
	Mean		Std dev
Ag	0.00103	±	0.000087
Al	4.95	±	0.10
As	0.0877	±	0.00041
B	8.70	±	0.11
Ba	0.693	±	0.0040
Be	0.000554	±	0.000054
Bi	0.00351	±	0.00019
Ca	73.0	±	2.0
Cd	0.00297	±	0.000068
Cl	3240	±	36
Co	0.419	±	0.0052
Cr	0.560	±	0.0057
Cu	1.26	±	0.017
Fe	51.7	±	0.53
Hg	0.00204	±	0.00017
Li	0.211	±	0.015
Mg	189	±	1.8
Mn	3.84	±	0.017
Mo	0.169	±	0.0034
Na	2580	±	62
Ni	0.652	±	0.0058
Pb	0.0138	±	0.00025
Sb	0.00561	±	0.00013
Se	0.304	±	0.0035
Sn	0.00500	±	0.00032
Sr	2.03	±	0.0041
Th	0.0141	±	0.0067
Ti	2.78	±	0.052
Tl	0.00172	±	0.000090
U	0.598	±	0.013
V	0.304	±	0.0025
Zn	4.31	±	0.039

Table 4. 76: Heavy metals for ARC Pilot Plant (1) culture

Dairy manure and glycerine

Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures					Post-digestion separation fractions						
	Initial, µg/g dw			Final, µg/g dw			Liquid, µg/g dw			Solid, µg/g dw		
	Mean	Std dev		Mean	Std dev		Mean	Std dev		Mean	Std dev	
Ag	0.0168	±	0.00052	0.0283	±	0.0010	0.0607	±	0.0040	0.0180	±	0.0010
Al	731	±	6.3	1650	±	13	3200	±	56	808	±	5.5
As	0.167	±	0.0047	0.360	±	0.010	0.722	±	0.0078	0.301	±	0.0047
B	21.4	±	0.32	23.0	±	0.90	45.4	±	1.9	19.1	±	0.67
Ba	24.5	±	0.32	47.7	±	0.32	79.6	±	0.64	29.3	±	0.37
Be	0.0401	±	0.0047	0.0480	±	0.0095	0.106	±	0.0082	0.0300	±	0.0028
Bi	0.0761	±	0.0012	0.0574	±	0.00063	0.119	±	0.0015	0.0379	±	0.00090
Ca	9740	±	150	14800	±	270	19800	±	450	14100	±	260
Cd	0.0960	±	0.0023	0.165	±	0.0031	0.318	±	0.0041	0.110	±	0.0020
Cl	13400	±	420	18600	±	77	37500	±	760	8390	±	250
Co	1.07	±	0.0074	1.70	±	0.011	3.96	±	0.055	1.04	±	0.018
Cr	3.64	±	0.27	9.60	±	0.11	23.2	±	1.4	6.31	±	0.17
Cu	33.1	±	0.51	54.8	±	0.47	112	±	1.5	32.9	±	0.45
Fe	462	±	12	1720	±	34	2490	±	15	1010	±	19
Hg	0.00505	±	0.0014	0.0118	±	0.00089	0.0207	±	0.0013	0.00821	±	0.00070
Li	1.38	±	0.022	1.46	±	0.035	4.66	±	0.19	0.752	±	0.015
Mg	3080	±	29	3860	±	55	8700	±	130	2790	±	79
Mn	101	±	0.15	151	±	1.2	292	±	4.9	118	±	2.9
Mo	0.856	±	0.011	1.78	±	0.017	3.78	±	0.032	1.03	±	0.0082
Na	3670	±	47	3890	±	67	12300	±	200	1680	±	23
Ni	1.79	±	0.025	4.15	±	0.032	7.26	±	0.13	2.42	±	0.049
Pb	0.296	±	0.0035	0.982	±	0.014	1.69	±	0.0081	0.513	±	0.0026
Sb	0.0313	±	0.0010	0.0733	±	0.0033	0.130	±	0.0026	0.0513	±	0.0014
Se	0.313	±	0.069	0.986	±	0.025	2.24	±	0.057	0.504	±	0.064
Sn	0.0745	±	0.00074	0.221	±	0.0029	0.365	±	0.0025	0.147	±	0.0031
Sr	36.7	±	0.36	64.9	±	0.54	110	±	0.89	50.7	±	0.45
Th	0.131	±	0.0021	0.287	±	0.0026	0.470	±	0.0046	0.144	±	0.0017
Ti	40.4	±	0.98	69.1	±	2.0	119	±	1.1	46.6	±	0.75
Tl	0.0414	±	0.00068	0.0759	±	0.0024	0.133	±	0.0036	0.0426	±	0.0013
U	0.684	±	0.0055	1.29	±	0.017	2.39	±	0.041	0.802	±	0.020
V	1.97	±	0.025	3.31	±	0.16	7.38	±	0.076	2.35	±	0.023
Zn	117	±	1.2	183	±	2.1	482	±	5.1	132	±	1.5

Table 4. 77: Heavy metals for ARC Pilot Plant (2) mixture
Wet distillers grains from ethanol production

Heavy metal	Results given as µg/g dw	
	Aqueous mixture	
	Mean	Std dev
Ag		±
Al	1.34	± 0.039
As	0.0662	± 0.00078
B	3.67	± 0.045
Ba	1.16	± 0.0039
Be		±
Bi		±
Ca	805	± 12
Cd	0.0262	± 0.000090
Cl	867	± 29
Co	0.0125	± 0.00018
Cr	0.0844	± 0.0016
Cu	1.96	± 0.018
Fe	7.33	± 0.048
Hg	0.00167	± 0.00045
Li	0.772	± 0.045
Mg	2590	± 50
Mn	56.8	± 1.1
Mo	0.636	± 0.0078
Na	1950	± 31
Ni	0.140	± 0.0039
Pb	0.000406	± 0.000095
Sb	0.000739	± 0.000031
Se	0.0687	± 0.017
Sn		±
Sr	6.21	± 0.036
Th	0.0000442	± 0.0000066
Ti	12.2	± 0.34
Tl	0.000375	± 0.000039
U	0.0000530	± 0.000011
V	0.00751	± 0.00025
Zn	47.0	± 1.3

Table 4. 78: Heavy metals for ARC Pilot Plant (2) culture

Wet distillers grains from ethanol production

Includes initial culture, final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures						Post-digestion separation fractions					
	Initial, µg/g dw			Final, µg/g dw			Liquid, µg/g dw			Solid, µg/g dw		
	Mean	±	Std dev	Mean	±	Std dev	Mean	±	Std dev	Mean	±	Std dev
Ag	0.00609	±	0.00057	0.0165	±	0.00024	0.0265	±	0.00042	0.0126	±	0.00044
Al	15.2	±	0.11	210	±	0.97	275	±	1.6	151	±	0.94
As	0.0627	±	0.0024	0.241	±	0.0079	0.480	±	0.010	0.188	±	0.0041
B	4.65	±	0.087	20.1	±	0.21	57.9	±	1.2	12.7	±	0.15
Ba	3.69	±	0.032	19.8	±	0.23	29.0	±	0.25	18.1	±	0.18
Be	0.00150	±	0.00070	0.0132	±	0.0028	0.0181	±	0.0014	0.0112	±	0.0037
Bi	0.00210	±	0.00023	0.204	±	0.0010	0.236	±	0.0020	0.132	±	0.0010
Ca	1530	±	12	10000	±	100	16200	±	160	9520	±	60
Cd	0.0431	±	0.0013	0.0954	±	0.0027	0.219	±	0.0026	0.0766	±	0.0019
Cl	4700	±	60	5120	±	510	34100	±	770	4830	±	85
Co	0.0195	±	0.00045	0.814	±	0.0068	2.42	±	0.016	0.506	±	0.0078
Cr	0.179	±	0.011	6.79	±	0.094	16.1	±	0.23	5.50	±	0.072
Cu	10.8	±	0.080	30.7	±	0.29	80.0	±	0.68	23.2	±	0.17
Fe	106	±	0.55	5200	±	51	9800	±	44	4350	±	26
Hg	0.00215	±	0.00036	0.00882	±	0.0011	0.0181	±	0.00052	0.00881	±	0.0012
Li	0.578	±	0.0097	1.73	±	0.027	8.48	±	0.16	0.465	±	0.018
Mg	2090	±	9.6	6710	±	33	6720	±	57	6660	±	46
Mn	59.3	±	0.16	214	±	0.66	296	±	1.6	195	±	0.59
Mo	1.52	±	0.011	6.28	±	0.080	19.7	±	0.16	3.87	±	0.043
Na	1790	±	12	6750	±	47	32300	±	260	1540	±	13
Ni	0.230	±	0.0018	6.18	±	0.067	17.0	±	0.10	4.33	±	0.028
Pb	0.0289	±	0.00034	0.478	±	0.0016	1.50	±	0.0098	1.04	±	0.0031
Sb	0.00230	±	0.00018	0.103	±	0.0020	0.209	±	0.0012	0.0798	±	0.0012
Se	0.248	±	0.023	0.516	±	0.034	1.38	±	0.056	0.238	±	0.033
Sn	0.00648	±	0.00057	0.233	±	0.0041	0.600	±	0.0049	0.111	±	0.0026
Sr	9.76	±	0.082	39.1	±	0.49	67.2	±	0.68	35.0	±	0.47
Th	0.00379	±	0.00020	0.0755	±	0.00047	0.0844	±	0.0010	0.0489	±	0.0010
Ti	16.7	±	0.27	42.3	±	0.76	61.0	±	0.42	37.0	±	0.24
Tl	0.000740	±	0.00012	0.0157	±	0.00041	0.0287	±	0.00051	0.0121	±	0.00043
U	0.00796	±	0.00019	0.273	±	0.0019	0.422	±	0.0043	0.253	±	0.0022
V	0.0342	±	0.0017	1.17	±	0.0099	1.84	±	0.019	0.932	±	0.022
Zn	50.6	±	0.35	131	±	1.3	259	±	1.2	113	±	0.83

4.7 Heavy metals for inoculum used in this study, please refer to Section 4.2 for explanations

Table 4. 79: Heavy metals for inoculum
Inoculum after collection and before use in cultures

Heavy metal	Results given as µg/g dw					
	Aqueous inoculum			Dried inoculum		
	Mean	±	Std dev	Mean	±	Std dev
Ag	0.00114	±	0.00099	0.109	±	0.0043
Al	2.88	±	0.68	4640	±	97
As	BD	±		1.37	±	0.033
B	8.32	±	0.17	26.3	±	0.88
Ba	2.26	±	0.0034	90.1	±	1.4
Be	BD	±		0.150	±	0.012
Bi	BD	±		0.0619	±	0.0020
Ca	735	±	20	23100	±	660
Cd	0.00615	±	0.00071	0.122	±	0.0037
Cl	16600	±	150	35000	±	1600
Co	0.567	±	0.027	4.32	±	0.11
Cr	BD	±		7.16	±	0.19
Cu	0.626	±	0.16	33.6	±	0.63
Fe	265	±	26	3860	±	120
Hg	0.0213	±	0.0042	0.0223	±	0.0022
Li	1.12	±	0.034	4.76	±	0.10
Mg	214	±	3.3	8060	±	140
Mn	3.97	±	0.066	209	±	3.4
Mo	0.253	±	0.028	2.17	±	0.054
Na	7880	±	61	10500	±	190
Ni	2.24	±	0.052	12.1	±	0.41
Pb	0.123	±	0.0057	2.45	±	0.042
Sb	0.00839	±	0.0018	0.0287	±	0.00058
Se	0.155	±	0.050	0.334	±	0.12
Sn	BD	±		0.0706	±	0.0019
Sr	15.7	±	0.51	150	±	4.9
Th	0.0384	±	0.0097	0.774	±	0.022
Ti	7.58	±	0.00028	183	±	5.3
Tl	0.00107	±	0.53	0.109	±	0.0033
U		±		0.494	±	0.012
V	0.369	±	0.036	8.86	±	0.24
Zn	8.88	±	1.3	201	±	5.5

Table 4. 80: Inoculum mixture and post digestion separation fractions, 44-day
Initial culture was inoculum after 7-day pre-culture incubation
Final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw		Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw	
	Mean	Std dev	Mean	Std dev	Mean	Std dev	Mean	Std dev
Ag	0.133	± 0.0036	0.160	± 0.0022	0.225	± 0.0037	0.115	± 0.0030
Al	6050	± 93	7600	± 52	9670	± 49	5290	± 24
As	1.65	± 0.047	2.24	± 0.022	2.73	± 0.030	1.15	± 0.015
B	27.8	± 1.3	29.1	± 0.46	35.6	± 0.77	16.3	± 0.14
Ba	97.6	± 2.1	103	± 1.0	127	± 1.4	61.9	± 0.77
Be	0.146	± 0.0089	0.148	± 0.016	0.180	± 0.0073	0.115	± 0.0082
Bi	0.0601	± 0.0015	0.0483	± 0.0012	0.0501	± 0.0012	0.0246	± 0.00062
Ca	23900	± 470	26100	± 430	33000	± 250	18300	± 230
Cd	0.126	± 0.0084	0.156	± 0.0031	0.187	± 0.0023	0.0975	± 0.0019
Cl	25500	± 980	22800	± 650	17900	± 140	7660	± 93
Co	4.98	± 0.085	5.07	± 0.040	5.91	± 0.032	2.93	± 0.016
Cr	8.15	± 0.16	11.1	± 0.062	15.5	± 0.13	8.99	± 0.060
Cu	39.3	± 1.0	50.7	± 0.41	61.1	± 0.47	32.0	± 0.34
Fe	4430	± 48	5430	± 29	6430	± 33	3910	± 30
Hg	0.0276	± 0.0022	0.0301	± 0.0021	0.0302	± 0.0016	0.0191	± 0.00067
Li	5.52	± 0.16	6.93	± 0.12	8.58	± 0.12	3.58	± 0.027
Mg	8910	± 120	8300	± 64	10000	± 49	5600	± 45
Mn	233	± 1.3	259	± 1.9	324	± 1.4	161	± 1.2
Mo	2.43	± 0.063	3.21	± 0.030	3.84	± 0.050	1.82	± 0.035
Na	11200	± 240	11600	± 93	16100	± 130	3130	± 37
Ni	13.9	± 0.18	12.3	± 0.11	15.8	± 0.14	6.92	± 0.098
Pb	2.82	± 0.033	2.69	± 0.022	3.26	± 0.0088	1.71	± 0.011
Sb	0.0313	± 0.0016	0.0438	± 0.00069	0.0511	± 0.00087	0.0297	± 0.00087
Se	0.900	± 0.053	1.44	± 0.092	1.59	± 0.071	0.920	± 0.034
Sn	0.0858	± 0.0040	0.297	± 0.0017	0.500	± 0.0024	0.195	± 0.0041
Sr	160	± 3.0	191	± 3.2	241	± 3.2	118	± 2.2
Th	1.13	± 0.018	1.14	± 0.013	1.50	± 0.016	0.713	± 0.0057
Ti	239	± 5.4	301	± 4.0	358	± 6.6	227	± 2.8
Tl	0.129	± 0.0032	0.0889	± 0.0010	0.106	± 0.00086	0.0592	± 0.0013
U	0.580	± 0.012	0.362	± 0.0029	0.440	± 0.0050	0.215	± 0.0014
V	11.5	± 0.27	15.3	± 0.12	18.9	± 0.10	9.52	± 0.091
Zn	220	± 3.7	283	± 4.2	350	± 5.3	193	± 4.3

Table 4. 81: Inoculum mixture and post digestion separation fractions, 30-day
Includes final culture and post culture separation fractions using dried sample.

Heavy metal	Cultures				Post-digestion separation fractions			
	Initial, µg/g dw	Final, µg/g dw		Liquid, µg/g dw		Solid, µg/g dw		
		Mean	Std dev	Mean	Std dev	Mean	Std dev	
Ag	Not done	0.0998	± 0.0025	0.130	± 0.0028	0.0671	± 0.0020	
Al		6630	± 29	7850	± 270	4090	± 34	
As		2.05	± 0.016	2.50	± 0.030	1.07	± 0.012	
B		26.4	± 0.62	30.5	± 0.82	14.5	± 0.58	
Ba		96.7	± 0.60	124	± 1.3	56.7	± 0.80	
Be		0.136	± 0.0067	0.171	± 0.011	0.0704	± 0.0057	
Bi		0.0385	± 0.0010	0.0533	± 0.0010	0.0262	± 0.00055	
Ca		29900	± 270	35000	± 410	16900	± 230	
Cd		0.168	± 0.0039	0.203	± 0.0028	0.0851	± 0.0011	
Cl		29300	± 300	71400	± 1700	18300	± 530	
Co		4.40	± 0.029	5.39	± 0.055	2.82	± 0.023	
Cr		14.0	± 0.14	15.0	± 0.11	6.57	± 0.044	
Cu		46.9	± 0.27	58.0	± 0.64	27.6	± 0.40	
Fe		4980	± 20	5780	± 58	3790	± 34	
Hg		0.0231	± 0.00066	0.0309	± 0.0017	0.0130	± 0.00070	
Li		5.53	± 0.072	6.77	± 0.31	2.62	± 0.076	
Mg		8700	± 53	10300	± 68	5420	± 67	
Mn		275	± 0.83	348	± 0.80	179	± 0.96	
Mo		2.95	± 0.015	3.62	± 0.038	1.67	± 0.038	
Na		11500	± 95	13500	± 120	2580	± 36	
Ni		13.8	± 0.069	16.0	± 0.21	6.35	± 0.073	
Pb		2.88	± 0.022	3.72	± 0.012	1.85	± 0.014	
Sb		0.0297	± 0.0019	0.0313	± 0.00073	0.0212	± 0.00090	
Se		1.16	± 0.042	1.18	± 0.063	0.788	± 0.041	
Sn		0.111	± 0.0016	0.0484	± 0.0016	0.0252	± 0.0015	
Sr		193	± 1.8	236	± 3.9	120	± 1.3	
Th		0.850	± 0.0070	0.734	± 0.0053	0.353	± 0.0030	
Ti		266	± 1.6	226	± 3.2	177	± 3.1	
Tl		0.0837	± 0.0020	0.105	± 0.0014	0.0562	± 0.0015	
U		0.348	± 0.0032	0.413	± 0.0034	0.180	± 0.0027	
V	12.8	± 0.075	14.7	± 0.14	7.56	± 0.11		
Zn	269	± 1.8	346	± 6.4	187	± 1.7		

5.0 Heavy Metal Results Compared to CCME Soil and Water Guidelines

Generally, for the batch cultures, it was expected that the heavy metals would be increased in the digestate due to loss of solids during digestion. This expectation was not seen in all the heavy metal results. However, the general trend for the results showed that there was an increase in the heavy metal concentrations during digestion. There is no obvious reason for the discrepancies. However, one reason could be that the substrate matrix was such that the heavy metals may not have been evenly distributed throughout the initial culture mixture during drying and regrinding even though great care was taken to mix samples. This is a consistent problem with biowaste substrates. As well, the initial and final culture samples for the substrates were not always analyzed for heavy metals at the same time, since they were produced 30 days apart. For each substrate, the final culture digestate, separated liquid and separated solid samples were analyzed for heavy metals at the same time. Since these samples were produced during digestion, they were likely more homogenous than the initial culture mixtures due to the break down of components during the digestion process. Comparison of these samples showed that there was an increase in the heavy metal concentration in the liquid fraction compared to the digestate and a decrease in the solid fraction compared to digestate and separated liquid. These results showed that the heavy metals generally remained in the liquid fraction of the digestate and were not largely associated with the solid fraction.

The CCME soil and water guidelines, shown in Tables 2.2 and 2.3, were used to compare the heavy metal results for the batch culture and the ARC Pilot Plant substrates and samples. The CCME guidelines for soils are given in mg/kg or $\mu\text{g/g}$ and for water, $\mu\text{g/L}$. The heavy metal results in the tables for this report are given in $\mu\text{g/g}$ and the $\mu\text{g/L}$ results were calculated from these results in order to match the units from the guidelines.

The comparison results are shown in Tables 5.1 to 5.5 for soil and Tables 5.6 to 5.10 for water. The comparison to CCME guidelines was done to evaluate the digestion products for land application and agricultural water use. The comparison for soil included substrate and all post-digestion samples. The comparison for water was only for the separated liquid fraction.

The soil comparisons were for substrate alone, digestate (culture mixture after incubation) and separated liquid and solid fractions. Results for aqueous available heavy metals were also compared and not entered in the tables, since only chicken broiler manure had a Cu result that was higher than the CCME guideline (Table 5.1).

For Tables 5.1 to 5.5, columns labeled “Heavy metals related to: unmixed substrate, digestate (substrate plus inoculum), separated liquid, separated solid”, the results were taken from the reports obtained from the Analytical Chemistry Laboratory, Environmental Management Group at ARC. Many of the results for Cu, Se and Zn were above the CCME guidelines, especially for manures. The inoculum had fairly high values for Cu, Se and Zn (Tables 4.79 to 4.81), so that adding substrate to batch culture mixtures did not always dilute these heavy metals to values below the guideline limit. This finding indicated that for anaerobic digestion, the inoculum will have an influence on heavy metal concentrations.

In order to get a clearer indication as to whether the substrates influenced the heavy metal results, calculations were done to subtract the inoculum heavy metal results from the culture mixture heavy metal results for each of the substrates. In order to do this, the following assumptions were made. The first assumption was that during anaerobic digestion, the inoculum in the culture mixture behaved in the same manner as when the inoculum was incubated alone. The second assumption was that the proportion of substrate in the separated liquid and solid fractions was the same as the proportion of substrate in the initial culture mixture. Given that the cultures contained biological activity, these assumptions were loosely applied. Therefore, the results from the subtraction of heavy metals in the inoculum were interpreted in a general sense and not to specific soil and water applications. For the tables, the heavy metals were entered in the columns, labeled “Heavy metals related to substrate in culture with values for inoculum subtracted”, only if it was clear that the subtraction of heavy metals from the substrate left a value that was higher than the lowest CCME guideline value.

This approach clearly showed that heavy metals from most vegetable/fruit/grain waste (Table 5.2) and waste associated with animals (Table 5.3) did not have elevated heavy metal levels for soil application. For the substrates used in this study, the potato plant vine substrate had elevated Cr and Ni (Table 5.2). These heavy metals were diluted

when inoculum was added to the culture mixture. There was elevated heavy metals in the separated liquid for grocery waste (Se, Table 5.2), poultry slaughterhouse waste (Se, Table 5.3), thermally hydrolyzed biowaste #1 (Se and Zn, Table 5.3) and thermally hydrolyzed biowaste #2 (Zn, Table 5.3). However, for agricultural water applications, the non-manure substrates had many heavy metal values that were above the CCME guidelines (Tables 5.7 and 5.8).

The manure substrates (Tables 5.1 and 5.6) and the sewage sludge (Tables 5.3 and 5.8) contained elevated heavy metal concentrations. Even the liquid sewage with low solids had elevated soil application values for Se (Table 5.3) and water application values for Al and Fe (Table 5.8). Clearly, these substrates would create a problem for disposal of post-digestion products.

For the ARC Pilot Plant substrates, no inoculum was added. Therefore, it was expected that the heavy metals in the digestion samples would be similar to the substrates that were used. For the ARC Pilot Plant 1, Cu, Se and Zn levels were above CCME soil guidelines for the separated liquid (Table 5.4). The Zn was elevated for the dairy manure #1 substrate (Table 4.29) and the digestate from the batch culture had elevated Cu and Zn in the separated liquid after the inoculum heavy metals were subtracted (Table 5.1). For the water CCME guidelines, shown in Table 5.9, ARC Pilot Plant 1 did not have any elevated heavy metals that were different from the dairy manure #1 substrate or the glycerine (data for substrates relating to CCME water guidelines not shown).

ARC Pilot Plant 2 had elevated soil Cu and Zn values in the separated liquid (Table 5.4) and these heavy metals were not seen for the wet distillers grain substrate or the batch culture digestate samples after the inoculum heavy metals were subtracted (Table 5.2). For the water CCME guidelines, shown in Table 5.9, the ARC Pilot Plant 2 liquid sample had elevated Ni that was not seen for the wet distillers grain substrate (data for substrates relating to CCME water guidelines not shown). The CCME guideline for Ni in irrigation water is 200 µg/L and the ARC Pilot Plant value was calculated to be 230 µg/L. The wet distillers grain substrate only had 44 µg/L of Ni, so this heavy metal must have come from another source.

It is likely that the results for the wet distillers grain for the ARC Pilot Plant 2 sample could be related to the continuous process that occurs in the ARC Pilot Plant

system. Dairy manure (#1) had been used in the ARC Pilot Plant from April, 2008 to November, 2008. The wet distillers grain was gradually added to the digester in November, 2008. By January, 2009 the digester was fed 100% wet distillers grain and the ARC Pilot Plant 2 digestate sample was taken in February, 2009. It is possible that residual dairy manure remained in the digester since only two hydraulic retention times (14 day each) with 100% wet distillers grain passed through the system before collection of the digestate sample. As well, heavy metals can combine with sulfur and form insoluble precipitates that could remain in a digester.

The overall conclusion for the heavy metals is that non-manure substrates do not appear to give elevated heavy metal values for soil application, but do have elevated values for agricultural water use. The fact that most digestion systems use manures as an inoculum for methanogenic bacteria indicates that levels of heavy metals in digestion products may need to be considered before these products are disposed in an environment. The continuously stirred reactors are designed to have a stream of substrates moving through their systems, and heavy metals may be retained or precipitated in a digester for a period of time after the substrate is used. This observation should be investigated further.

An examination of minimum acceptable ratios of nitrogen and phosphorus to metals set out in the Guidelines for the Application of Municipal Wastewater Sludge to Agricultural Lands by Alberta Environment may give more information as to whether the manure and sewage sludge post-digestion products could be used for soil and agricultural application.

Table 5. 1: CCME soil guideline comparison for manures.

Please see explanation in Section 5.0

Biowaste substrates ^a	Heavy metals above the CCME soil quality guideline concentrations for given soil types ^b				
	A	P/R	I	C	
Manures (see footnote for sample types)	Heavy metals related to: unmixed substrate, digestate (substrate plus inoculum), separated liquid, separated solid				Heavy metals related to substrate in culture with values for inoculum subtracted
<i>Cattle bed pack</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Se,Zn	Se,Zn	Zn	Zn	Se,Zn
Solid ^a					
<i>Dairy manure #1</i>					
Substrate ^a	Zn	Zn			Not applicable
Digestate ^a	Zn	Zn			Zn
Liquid ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Zn
Solid ^a					
<i>Dairy manure #2</i>					
Substrate ^a	Cu	Cu			Not applicable
Digestate ^a	Cu	Cu			Cu
Liquid ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Se,Zn
Solid ^a					
<i>Chicken laying hen</i>					
Substrate ^a	Se,Zn	Se,Zn	Zn	Zn	Not applicable
Digestate ^a	Se,Zn	Se,Zn	Zn	Zn	Se,Zn
Liquid ^a	Cu,Se,Zn	Cu,Se,Zn	Zn	Zn	Cu,Se,Zn
Solid ^a					
<i>Chicken broiler</i>					
Aqueous substrate ^c	Cu	Cu	Cu	Cu	Not applicable
Substrate ^a	Cu,Zn	Cu,Zn	Cu,Zn	Cu,Zn	Not applicable
Digestate ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Zn

Liquid ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Se,Zn
Solid ^a	Cu,Se	Cu,Se	Cu	Cu	Cu,Se
<i>Solid hog</i>					
Substrate ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Not applicable
Digestate ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Se,Zn
Liquid ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Se,Zn
Solid ^a	Cu,Zn	Cu,Zn	Cu	Cu	Cu,Zn
<i>Liquid hog</i>					
Substrate ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Se,Zn	Cu,Se,Zn	Not applicable
Digestate ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	
Liquid ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Se,Zn
Solid ^a	Zn	Zn			Zn

^a Substrates from Section 3.0; Substrate refers to substrate only; Digestate refers to mixture with inoculum after incubation; Liquid refers to liquid portion of digestate after separation; Solid refers to solid portion of digestate after separation.

^b A – Agricultural; P/R – Parks and Residential; I – Industrial; C – Commercial

^c Aqueous substrate is wet substrate diluted with water to 2 g TS. See section 2.3

Table 5. 2: CCME soil guideline comparison for vegetable/fruit/grain.
Please see explanation in Section 5.0

Biowaste substrates ^a	Heavy metals above the CCME soil quality guideline concentrations for given soil types ^b				
	A	P/R	I	C	
Vegetable/fruit/grain (see footnote for sample types)	Heavy metals related to: unmixed substrate, digestate (substrate plus inoculum), separated liquid, separated solid				Heavy metals related to substrate in culture with values for inoculum subtracted
<i>Potato tuber</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Zn	Zn			
Solid ^a					
<i>Potato process</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Zn	Zn			
Solid ^a					
<i>Potato plant vines</i>					
Substrate ^a	Cr,Ni	Cr,Ni	Cr,Ni	Cr,Ni	Not applicable
Digestate ^a					
Liquid ^a					
Solid ^a					
<i>Sugar beet tailings</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Zn	Zn			
Solid ^a					
<i>Grocery</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Se,Zn	Se,Zn	Zn	Zn	Se

	Solid ^a					
<i>Glycerine</i>						
	Substrate ^a					Not applicable
	Digestate ^a	Zn	Zn			
	Liquid ^a	Zn	Zn			
	Solid ^a					
<i>Thin stillage</i>						
	Substrate ^a					Not applicable
	Digestate ^a	Zn	Zn			
	Liquid ^a	Zn	Zn			
	Solid ^a					
<i>Wet distillers grains</i>						
	Substrate ^a					Not applicable
	Digestate ^a	Zn	Zn			
	Liquid ^a	Zn	Zn			
	Solid ^a					

^a Substrates from Section 3.0; Substrate refers to substrate only; Digestate refers to mixture with inoculum after incubation; Liquid refers to liquid portion of digestate after separation; Solid refers to solid portion of digestate after separation.

^b A – Agricultural; P/R – Parks and Residential; I – Industrial; C – Commercial

Table 5. 3: CCME soil guideline comparison for human and other animal waste.
 Human waste and non-manure animal waste. Please see explanation in Section 5.0

Biowaste substrates ^a	Heavy metals above the CCME soil quality guideline concentrations for given soil types ^b				
	A	P/R	I	C	Heavy metals related to substrate in culture with values for inoculum subtracted
	Heavy metals related to: unmixed substrate, digestate (substrate plus inoculum), separated liquid, separated solid				
Waste associated with animals (see footnote for sample types)					
<i>Beef slaughterhouse paunch</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Zn	Zn			
Solid ^a					
<i>Hog slaughterhouse sludge</i>					
Substrate ^a	Cu	Cu			Not applicable
Digestate ^a	Not done				
Liquid ^a					
Solid ^a					
<i>Hog slaughterhouse peptone</i>					
Substrate ^a	Se	Se			Not applicable
Digestate ^a	Not done				
Liquid ^a					
Solid ^a					
<i>Poultry slaughterhouse waste</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Se,Zn	Se,Zn			Se
Solid ^a					
<i>Thermally hydrolyzed biowaste #1</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			

Liquid ^a	Se,Zn	Se,Zn	Zn	Zn	Se,Zn
Solid ^a					
<i>Thermally hydrolyzed biowaste #2</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Zn	Zn	Zn	Zn	Zn
Solid ^a					
<i>Cheese whey</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Zn	Zn			
Solid ^a					
<i>Cattle carcass</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Zn	Zn			
Solid ^a					
Waste associated with humans					
<i>Sewage sludge</i>					
Substrate ^a	Cu,Se,Zn	Ba,Cu,Se,Zn	Cu,Zn	Cu,Zn	Not applicable
Digestate ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Se,Zn
Liquid ^a	Cu,Se,Zn	Cu,Se,Zn	Cu,Zn	Cu,Zn	Cu,Se,Zn
Solid ^a	Cu,Zn	Cu,Zn			Cu,Se,Zn
<i>Liquid sewage</i>					
Substrate ^a					Not applicable
Digestate ^a	Zn	Zn			
Liquid ^a	Se,Zn	Se,Zn			Se
Solid ^a					

^a Substrates from Section 3.0; Substrate refers to substrate only; Digestate refers to mixture with inoculum after incubation; Liquid refers to liquid portion of digestate after separation; Solid refers to solid portion of digestate after separation.

^b A – Agricultural; P/R – Parks and Residential; I – Industrial; C – Commercial

Table 5. 4: CCME soil guideline comparison for ARC Pilot Plant samples.
Please see explanation in Section 5.0

Substrates ^a	CCME soil quality guidelines exceeded for heavy metal and soil types ^b			
	A	P/R	I	C
ARC Pilot Plant 1; Heavy metals related to dairy manure #1 and glycerine				
Mixture for digestion				
Digestate (after incubation)				
Separated liquid	Cu,Zn,Se	Cu,Zn,Se	Cu,Zn	Cu,Zn
Separated solid				
ARC Pilot Plant 2; Heavy metals related to wet distillers grain				
Mixture for digestion				
Digestate (after incubation)				
Separated liquid	Cu,Zn	Cu,Zn		
Separated solid				

^a ARC Pilot Plant 1 - substrate mixture of dairy #1 and glycerine; ARC Pilot Plant 2 - substrate was wet distillers grain, Digestate refers to mixture after incubation; Liquid refers to liquid portion of digestate after separation; Solid refers to solid portion of digestate after separation.

^b A – Agricultural; P/R – Parks and Residential; I – Industrial; C – Commercial

Table 5. 5: CCME soil guideline comparison for inoculum.
Please see explanation in Section 5.0

Inoculum^a used in batch cultures and incubated for 30 and 45 days (see footnote for sample type)^a	CCME soil quality guidelines exceeded for heavy metal and soil types^b			
	A	P/R	I	C
Inoculum (after collection)	Zn	Zn		
Inoculum (before use in cultures)	Zn	Zn		
Inoculum (after 30 d incubation)	Zn	Zn		
Separated liquid	Zn	Zn		
Separated solid				
Inoculum (after 45 d incubation)	Zn	Zn		
Separated liquid	Zn	Zn		
Separated solid				

^a Inoculum (after collection) - Inoculum from collection pails; Inoculum (before use in cultures) - Inoculum after 7 day incubation and before mixture with substrate; Inoculum (after 30 day/45 day incubation) - refers to inoculum after incubation; Liquid refers to liquid portion of inoculum after separation; Solid refers to solid portion of inoculum after separation.

^b A – Agricultural; P/R – Parks and Residential; I – Industrial; C – Commercial

Table 5. 6: CCME water guideline comparison for manures
Separated liquid from digestate after digestion. Please see explanation in Section 5.0

Biowaste substrates ^a	Heavy metals above the CCME water quality guideline concentrations for agricultural use		
	Irrigation water	Livestock water	
Manures	Heavy metals related to separated liquid containing substrate and inoculum		Heavy metals related to separated liquid with values for inoculum subtracted
<i>Cattle bed pack</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Ni,Pb,Se,V	Al,As,B,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,V,Zn
<i>Dairy manure #1</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Pb,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,B,Cl,Co,Cu,Fe,Mn,Mo,Se,U,V,Zn
<i>Dairy manure #2</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,B,Cd,Cu,Fe,Mn,Mo,Se,U,V,Zn
<i>Chicken laying hen</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Se,V	Ca,Cd,Cl,Cu,Mn,Mo,Se,Zn
<i>Chicken broiler</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Cu,Mn,Mo,Se,Zn
<i>Solid hog</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu, Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Cd,Cu,Fe,Mn,Mo,Se,Zn
<i>Liquid hog</i>			
Separated liquid	Al,BCd,Cl,Co,Cu,Fe,M n,Mo,Ni,Se, U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Cu,Fe,Mn,Mo,Se,

Table 5. 7: CCME water guideline comparison for vegetable/fruit/grain
 Separated liquid from digestate after digestion. Please see explanation in Section 5.0

Biowaste substrates ^a	Heavy metals above the CCME water quality guideline concentrations for agricultural use		
	Irrigation water	Livestock water	
Vegetable/fruit/grain	Heavy metals related to separated liquid containing substrate and inoculum		Heavy metals related to separated liquid with values for inoculum subtracted
<i>Potato tuber</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Cd,Fe
<i>Potato process</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu, Fe,Mn,Mo,Ni,Pb,Se,U, V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Fe,Mn
<i>Potato plant vines</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu, Fe,Mn,Mo,Ni,Se,U,V, Zn	Al,As,Ca,Cu,Hg,Ni,V,Zn	Al,As,B,Cd,Fe,Hg,Mn,Mo,Ni,V
<i>Sugar beet tailings</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu, Fe,Mn,Mo,Ni,Se,U,V, Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Fe,Mn,Mo,V
<i>Grocery</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu, Fe,Mn,Mo,Ni,Se,U,V, Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Fe,Mn,Mo,V
<i>Glycerine</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu, Fe,Mn,Mo,Ni,Se,U,V, Zn	Al,As,Ca,Cu,Pb,Se,V	Cl,Mn
<i>Thin stillage</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu,	Al,As,Ca,Cu,Fe,Pb,Se,V	Mn

	Fe,Mn,Mo,Ni,Se,U,V, Zn		
<i>Wet distillers grains</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,V	Al,Fe,Mn,Mo,

Table 5. 8: CCME water guideline comparison for human and other animal

Human waste and non-manure animal waste. Separated liquid from digestate after digestion. Please see explanation in Section 5.0

Biowaste substrates ^a	Heavy metals above the CCME water quality guideline concentrations for agricultural use		
	Irrigation water	Livestock water	Heavy metals related to separated liquid with values for inoculum subtracted
	Heavy metals related to separated liquid containing substrate and inoculum		
Waste associated with animals			
<i>Beef slaughterhouse paunch</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,V	Cl
<i>Poultry slaughterhouse waste</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Pb,Se,V	Al,Fe,Mn,Mo
<i>Thermally hydrolyzed biowaste #1</i>			
Separated liquid	Al,As,B,Cd,Cl,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Fe,Mn,Mo,V,Zn
<i>Thermally hydrolyzed biowaste #2</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Fe,Pb,Se,V	Al,Fe,Mn,Zn
<i>Cheese whey</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,V	Al
<i>Cattle carcass</i>			
Separated liquid	Al,As,B,Cd,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,V	Al,Fe,Mn
Waste associated with humans			
<i>Sewage sludge</i>			

Separated liquid	Al,As,B,Cd,Cl,Co,Cu, Fe,Mn,Mo,Ni,Pb,Se,U, V,Zn	Al,As,Ca,Cu,Hg,Pb,Se,V	Al,As,Cd,Cu,Fe,Hg,Mn,Mo,Ni,Pb,Se,U,V,Zn
<i>Liquid sewage</i>			
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe, Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V	Al,Fe

Table 5. 9: CCME water guideline comparison for ARC Pilot Plant samples
 Separated liquid from digestate after digestion. Please see explanation in Section 5.0

Biowaste substrates ^a	Heavy metals above the CCME water quality guideline concentrations for agricultural use	
	Irrigation water	Livestock water
ARC Pilot Plant 1 (dairy #1 and glycerine); Heavy metals for separated liquid		
Separated liquid	Al,B,Cd,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Cu,Se,U,V
ARC Pilot Plant 2 (wet distillers grains); Heavy metals for separated liquid		
Separated liquid	B,Cu,Fe,Mn,Mo,Ni,Zn	Cu

Table 5. 10: CCME water guideline comparison for inoculum

Separated liquid from digestate after digestion. Please see explanation in Section 5.0

Inoculum used in study and incubated for 30 and 45 days	Heavy metals above the CCME water quality guideline concentrations for agricultural use	
	Irrigation water	Livestock water
Separated liquid	Al,As,B,Cd,Cl,Co,Cu,Fe,Mn,Mo,Ni,Se,U,V,Zn	Al,As,Ca,Cu,Pb,Se,V

6.0 Summary

The following summary points have been taken from this study:

- Manure and non-manure substrates contained sufficient amounts of carbon, nitrogen, phosphorous and potassium for anaerobic digestion.
- Carbon, nitrogen, phosphorous and potassium remained in post-digestion samples.
- Some substrates contained high values of nitrogen and sulphur that can cause toxicity in a digester.
- Substrates with low VS may have low organic carbon concentrations.
- Manure substrates used in this study had elevated heavy metal concentrations that exceeded the CCME guidelines for specified soil and CCME water guidelines for agricultural use (irrigation and livestock applications).
- Non-manure substrates used in this study did not have elevated heavy metal concentrations that exceeded the CCME specified soil guidelines except for grocery waste, poultry slaughterhouse waste and thermally hydrolyzed biowaste.
- Non-manure substrates gave elevated heavy metal concentrations that exceeded the CCME water guidelines for agricultural use (irrigation and livestock applications).
- Heavy metals were generally concentrated during anaerobic digestion due to loss of solids. When digestate was separated into liquid and solid fractions, most of the heavy metals were associated with the liquid fraction and not with the solid fraction, giving elevated levels in the liquid.
- Heavy metals from manures may be retained in large digester systems due to incomplete purging of substrates and precipitation. This should be investigated further.
- Electrical conductivity results or salt concentration for substrates, digestates and separated liquids were high using the 2% TS method.
- Manure substrates generally had higher salt concentrations than non-manure substrates associated with animals or vegetable/grain substrates.
- Vegetable/grain substrates had the lowest overall salt concentrations compared to manure and non-manure substrates associated with animals.

- Salt concentrations for separated solids were generally less than those for separated liquid. However, repeated application of separated solids to land without proper monitoring could lead to salt accumulation. More work needs to be done to reduce salts in the solid fraction and to determine salt accumulation for different soil types.

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PART B: EVALUATION OF PATHOGEN IN BIOWASTE AND DIGESTATE

1.0 Introduction

There are many different types of waste products that are used for anaerobic digestion, however, biowaste that contains manure has a high density of coliform bacteria (1,2,3,4,5,6). The coliform bacteria can include pathogens associated with human illness, such as *Salmonella* and other zoonotic pathogens such as *Campylobacter* and *Listeria* (7,8,9,10). Generally, methods used to denote contamination in waste use indicator organisms like fecal coliform bacteria. For water, detection and enumeration of this group of organisms are used to determine the suitability of water for domestic and industrial use (11). In the United States, sludge from wastewater treatment plants must fulfill the density requirements from the US Environmental Protection Agency (USEPA) for fecal coliform as an indicator or *Salmonella* as a pathogen (12).

In the discussion presented by Pell (13) on pathogenic microbes in manure, there is mention that in the past, most environmental concerns about biowaste management have focused on nutrient overload, water quality or odour problems. There are no regulations concerning pathogens in biowaste that are used for anaerobic digestion. With an emerging biogas industry in Alberta, large amounts of effluent from anaerobic digesters will be produced. There is a lack of information as to whether pathogens are present in anaerobic digester effluent and if present, whether they will pose a threat to public, animal and plant health. We have found no information on regulations for handling effluent from anaerobic digesters for Alberta, although there is information on wastewater systems (14). Alberta Agriculture and Rural Development guidelines mention that land application of digestate is under the Agricultural Operations Practices Act and Regulations as it applies to manure (15). The Canadian Council for the Ministers of the Environment (CCME), in their guidelines for organism content in compost containing only yard waste, mention that fecal coliform of fecal origin should be < 1000 Most Probable Number (MPN) /g of Total Solids (TS) calculated on a dry weight basis and *Salmonella* < 3 MPN/4 g TS (16) and compost containing other feedstock should contain fecal coliform at < 1000 MPN/g TS or *Salmonella*, < 3 MPN/4g TS. The compost with other feedstock must be exposed to temperatures of 55°C or higher for a specified time depending on the type of compost.

The USEPA have imposed regulations under Title 40 of the Code of Federal Regulations (CFR), Part 503 to control the use and disposal of biosolids (17). Biosolids are defined as the recyclable organic solid product produced during wastewater treatment processes. Part 503 of the rule gives the requirements for the use of biosolids in order to prevent contamination to the public and the environment. One requirement is for the control of pathogens or disease-causing organisms and the reduction of vector attraction to the biosolids. Pathogens can be bacteria, viruses and parasites and vectors include rodents, flies, mosquitoes and disease-carrying and transferring organisms. The rules described in Part 503 ensure that pathogen levels are safe for the biosolids to be land applied or surface disposed. The criteria for biosolid Class A are the same as the CCME guidelines for compost with other feedstock, with fecal coliform < 1000 MPN/ g TS or *Salmonella* < 3 MPN/4 g TS. A biosolid is considered Class B if pathogens are reduced to levels that do not pose a risk to the public and environment. Measures must be taken to prevent crop harvesting, animal grazing and public access to areas where Class B biosolid have been applied until the area is considered safe. The Class B biosolid requirements are that fecal coliform must be < 2 x 10⁶ MPN/g TS. For this biosolid, the fecal coliform is used as an indicator of average density of bacterial and viral pathogens.

We conducted a small-scale study on undigested biowaste and effluent after anaerobic digestion of biowaste using the USEPA microbiology testing methods for fecal coliform (18) and *Salmonella* (19) for biosolids and used the results to assess local biowaste samples. Due to time and funding restrictions, we were only able to perform selected analyses on chosen biowaste samples.

2.0 Objectives

- to assess the levels of fecal coliform used as a contamination indicator and *Salmonella* used as pathogen indicator for selected biowaste samples,
- to evaluate reduction of fecal coliform and *Salmonella* using thermophilic anaerobic digestion processes.

The results from this study will provide preliminary data for development of guidelines for handling and utilizing biowaste in Alberta.

3.0 Biowaste and Sample Collection

All samples were collected into sterile plastic bags or bottles and tested within 2-3 hours after collection, unless otherwise stated. All samples were collected specifically for this study except sample 3.4, which was collected and stored at ARC, Vegreville, Alberta. This sample was being used in the ARC fully automated anaerobic digestion system ARC Pilot Plant (referred to as ARC Pilot Plant from here on) at the time of this study. The digestion system operated at 55°C. All dairy and chicken manure samples were collected from the same farm in the winter months. The farm was chosen because of its close proximity to the testing laboratory, allowing valid testing of fecal coliform and *Salmonella* within the required time frame for the USEPA microbiological testing methods.

The following samples were tested in this study:

3.1	Dairy manure taken from within dairy cows. Three dairy manure samples collected on two occasions from 5 dairy cows. Sample 1 was a manure mixture from cows 1 and 2, and Sample 2 was a mixture from cows 3 and 4. Sample 3 was from cow 5. One sample was tested for <i>Salmonella</i> only.
3.2	Dairy manure from one cow that was collected from the barn and tested for <i>Salmonella</i> only.
3.3	Dairy manure collected from the general barn area. Some of the freshly collected manure was taken to the Edmonton ARC laboratory. The remainder of the manure was transported to Vegreville and digested in the ARC Pilot Plant. At this time the digester was running dairy manure at 55°C and 14-day hydraulic retention time. The freshly collected dairy manure was fed into the digester over 10 days. The last feeding of manure was 15 hours before the sample was taken for analysis.
3.4	Dairy manure that was used routinely for digestion at the ARC Pilot Plant. The dairy manure was collected from the same farm as samples 3.1 to 3.3 and stored for 2 months at 4°C. The stored sample and a random sample from the digester hopper were tested. The dairy manure from the hopper was diluted in the laboratory and left at 22°C for 1 hour. As well, after a 14-day hydraulic

	retention time in the ARC Pilot Plant, a digested sample from the dairy manure was collected and tested.
3.5	Chicken manure, collected from chicken cages in the barn.
3.6	Chicken manure, collected from the general barn area and included straw bedding.
3.7	Household kitchen waste, mostly vegetable and fruit waste collected daily over a 7-day period and held at 4-6°C until testing.
3.8	Broken eggs, including shell, collected at a grocery retail store that was close to the testing laboratory
3.9	Wet distillers grain from an ethanol production plant, collected in barrels and stored at -20°C until testing in the ARC Pilot Plant. This sample was collected for use in the ARC Pilot Plant and was chosen for pathogen analysis because it was a non-manure based biowaste. A diluted sample with 8% TS was taken for fecal coliform and <i>Salmonella</i> testing.

4.0 Testing Methods

All dehydrated culture media were purchased from Neogen (MI, USA) and testing was carried out in a Biolevel II lab, Alberta Research Council, Edmonton. A 5-tube MPN method was used as described in the USEPA methods to derive population estimates for the fecal coliform and *Salmonella*.

4.1 Total solid measurements of biowaste

Total solid analysis was done for biowaste using a forced-air oven-drying method at 70°C for 48 hours. The method assumes only water is removed. The results are reported as a percent of the sample's wet weight.

4.2 Testing for fecal coliform

The biowaste and anaerobic digester effluent were evaluated for fecal coliform using the USEPA Method 1680 (17). Briefly, the method uses a MPN procedure to derive a population estimate for fecal coliform bacteria, Lauryl-Tryptose broth and EC culture-

specific media and elevated temperature to isolate and enumerate fecal coliform organisms. The basis for the test is that fecal coliform bacteria, including *Escherichia coli* (*E. coli*), are commonly found in the feces of humans and other warm-blooded animals. These bacteria indicate the potential presence of other bacterial and viral pathogens. Total solids determination was done on the biowaste samples and used to calculate and report fecal coliform as MPN/g dry weight.

4.3 *Testing for Salmonella sp.*

The biowaste and anaerobic digester effluent were evaluated for *Salmonella* using the USEPA Method 1682 (18). Briefly, the method is for the detection and enumeration of *Salmonella* by enrichment with tryptic soy broth and selection with modified semisolid Rappaport-Vassiliadis medium. Presumptive identification was done using xylose-lysine desoxycholate agar and confirmation was done using lysine-iron agar, triple sugar iron agar and urea broth. Serological testing was done. Total solids were determined on a representative biowaste sample and used to calculate *Salmonella* density as MPN per 4 g dry weight.

4.4 *Quality control*

Milorganite (CAS 8049-99-8, Milwaukee Metropolitan Sewerage District, UNGRO Corp. ON), a heat-dried Class A biosolid proven by USEPA was used and spiked with appropriate control bacteria. *E.coli* (ATCC# 25922) was used as the positive control for the fecal coliform test and negative control for the *Salmonella* test. *Salmonella typhimurium* (ATCC# 14028) was used as the positive control for the *Salmonella* test. *Enterobacter aerogenes* (ATCC# 13048) and *Pseudomonas* (ATCC# 27853) were used as negative controls for the fecal coliform test.

5.0 Results and Discussion

Table 5.1 gives the total solid, fecal coliform and *Salmonella* MPN for the biowaste samples.

Table 5. 1: Summary of microbiology testing results of selected biowaste samples

Samples	Total solids (% of wet weight)	Fecal coliform (MPN/g TS)	<i>Salmonella</i> (MPN/4g TS)
# ^a			
3.1 Dairy manure taken from within dairy cows			
Sample 1	13	5.6×10^6	< 0.18
Sample 2	15	1.1×10^7	< 0.18
Sample 3	14 ^b	Not done	< 0.18
3.2 Dairy manure from general barn area	14 ^b	Not done	< 0.18
3.3 Dairy manure from general barn area	15	1.1×10^7	4.0×10^0
Anaerobic digestion effluent of dairy manure after 15 h digestion	10	< 0.18	< 0.18
3.4 Dairy manure used at ARC Pilot Plant			
Dairy manure stored for 2 months at 4°C	14	8.8×10^4	< 0.18
Dairy collected from ARC Pilot Plant hopper before anaerobic digestion	10	1.8×10^4	2.1×10^0
Anaerobic digestion effluent of dairy manure after 14 day hydraulic retention time	9	<0.18	< 0.18
3.5 Chicken manure from cages	37	4.3×10^6	< 0.18
3.6 Chicken manure from general barn area with straw bedding	78	2.1×10^6	< 0.18
3.7 Household kitchen waste	Not done	No growth	No growth
3.8 Broken eggs	Not done	No growth	No growth
3.9 Wet distillers grains	8	<0.18	< 0.18

^a Sample # relating to Section 3 ^b Estimated TS values

Dairy manure samples from the same facility were tested in this study. The samples were from the general barn area and taken from within cows. When tested, the density of fecal coliform that was found in all samples ranged from 8.8×10^4 MPN/g TS to 1.1×10^7 MPN/g TS. *Salmonella*, 4×10^0 MPN/4g TS, was found in one sample collected from the general barn area. Storage of the dairy manure at 4°C for 2 months decreased the fecal coliform 2- to 3-log. A random dairy manure sample was collected from the hopper of the ARC Pilot Plant and tested. It contained 1.8×10^4 MPN/g TS fecal coliform and 2.1×10^0 MPN/4g TS *Salmonella*. In both cases where dairy manure was digested at 55°C in the ARC Pilot Plant, the fecal coliform and *Salmonella* were decreased to below detection (<0.18 MPN/g TS for fecal coliform and <0.18 MPN/4g TS for *Salmonella*).

The chicken manures, kitchen waste, eggs and wet distillers grain were not put through digestion. Both chicken manure samples had fecal coliform, 4.3×10^6 and 2.1×10^6 MPN/g TS. No *Salmonella* was detected. There were no fecal coliform and *Salmonella* in the kitchen waste, eggs and wet distillers grains.

This brief study showed that bacteria common to manures were detected in the dairy and chicken manure samples. According to the USEPA guidelines for a Class A biosolid, the fecal coliform density was above the accepted level in all manure samples, and for a Class B biosolid, the fecal coliform density was above the accepted level in the freshly collected manure samples. The increased fecal coliform levels indicate that pathogenic bacteria could be present in these samples. This was verified by the fact that one fresh dairy sample contained 4.0×10^0 MPN/4g TS and a random hopper sample from the ARC Pilot Plant contained 2.1×10^0 MPN/4g TS *Salmonella*. The result from the hopper sample would have been considered acceptable according to the USEPA standards. Nevertheless, both samples of dairy manure were tested to contain below detection levels of both fecal coliform and *Salmonella* after anaerobic digestion at 55°C and 14-day hydraulic retention time in the ARC Pilot Plant digesters. For sample 3.3, based on the feeding style of the ARC Pilot Plant continuous stirred tank reactor digester system and the time the dairy manure was in the digester, there was about 15 hours of digestion between the last feed time and the time a sample was taken for analysis.

Bendixen (20) looked at the animal and human pathogen reduction in Danish biogas plants. It was reported that pathogen survival was greatly reduced at thermophilic digestion temperatures (50°C to 55°C) but not at low and mesophilic temperatures (5°C to 45°C). Biogas plant construction, function and management need to be monitored in order to assure pathogen destruction and policies need to be in place to classify the digested effluent for proper disposal.

The requirements in the USEPA standards (17) for sewage sludge use and disposal indicate that sewage sludge should be analyzed for enteric viruses and viable helminth ova. There are also requirements given for vector attraction reduction and reduction of volatile solids. As well, other pathogens should be investigated. For example, human norovirus strains have been found in livestock, indicating a route for zoonotic transmission (21). As well, policies have been made concerning plant pathogens that relate to anaerobic digestion facilities in Germany (22).

6.0 Summary

- Using the USEPA Class A biosolids and CCME guideline for compost of <1000 MPN/g TS for fecal coliform, all the freshly collected manures (dairy and chicken) were above the accepted level.
- Using the USEPA Class B biosolids guidelines of <2 x 10⁶ MPN/g TS for fecal coliform, all the freshly collected manure samples (dairy and chicken) were above the accepted level.
- For one fresh dairy manure, the *Salmonella* exceeded the USEPA Class A biosolids and CCME guideline for compost of <3 MPN/4 g TS.
- Storage of dairy manure at 4°C for 2 months decreased fecal coliform concentration.
- Anaerobic digestion at 55°C and 14-day hydraulic retention time in the ARC Pilot Plant reduced fecal coliform and *Salmonella* to below detection levels. Fifteen hours of digestion in a continuous stirred tank reactor system appeared to be inadequate for reduction.

- Household kitchen waste, broken eggs and wet distillers grains contained either no fecal coliform and *Salmonella* or levels below detection using the MPN method.

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